Package 'progeny'

January 21, 2025

```
Title Pathway RespOnsive GENes for activity inference from gene expression
```

Version 1.28.0

Description PROGENy is resource that leverages a large compendium of publicly available signaling perturbation experiments to yield a common core of pathway responsive genes for human and mouse. These, coupled with any statistical method, can be used to infer pathway activities from bulk or single-cell transcriptomics.

```
URL https://github.com/saezlab/progeny
BugReports https://github.com/saezlab/progeny/issues
Depends R (>= 3.6.0)
Imports Biobase, stats, dplyr, tidyr, ggplot2, ggrepel, gridExtra,
      decoupleR, reshape2
biocViews SystemsBiology, GeneExpression, FunctionalPrediction,
      GeneRegulation
License Apache License (== 2.0) | file LICENSE
LazyData true
LazyDataCompression bzip2
Encoding UTF-8
Suggests airway, biomaRt, BiocFileCache, broom, Seurat,
      SingleCellExperiment, DESeq2, BiocStyle, knitr, readr, readxl,
      pheatmap, tibble, rmarkdown, testthat (\geq 2.1.0)
VignetteBuilder knitr
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getModel Returns the Progeny Model

Description

This function is designed for getting a model matrix with top significant genes for each pathway

Usage

Index

```
getModel(organism = "Human", top = 100, decoupleR = F)
```

Arguments

organism	"Human" or "Mouse" taken from the main function's argument. Default to "Human"
top	Desired top number of genes for each pathway according to their significance(p.value). Default to 100
decoupleR	if TRUE, the model matrix is goign to be generated with a format makes it directlz compatible with the decoupleR package

Value

This function returns model matrix according to the top n significant

```
\label{thm:condition} \mbox{\tt \#getting a model matrix according to the desired top n significant} \\ \mbox{\tt model} <- \mbox{\tt getModel("Human", top=100)} \\
```

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model_human_full

The full human linear model underlying PROGENy

Description

HGNC gene symbols in rows, pathways in columns. Pathway activity inference works by matrix multiplication of gene expression with the model.

Format

The full human model contains 22479 genes, associated pathways, weight and the p-value.

```
gene gene names in HGNC symbolspathway names of PROGENy pathwaysweight z-scores for a given genep.value significance of gene in pathway
```

Source

```
https://www.nature.com/articles/s41467-017-02391-6
```

Examples

```
get("model_human_full", envir = .GlobalEnv)
```

model_mouse_full

The full mouse linear model underlying PROGENy

Description

MGI gene symbols in rows, pathways in columns. Pathway activity inference works by matrix multiplication of gene expression with the model.

Format

The full mouse model contains 17426 genes, associated pathways, weight and the p-value.

```
gene gene names in HGNC symbolspathway names of PROGENy pathwaysweight z-scores for a given genep.value significance of gene in a pathway
```

Source

```
https://www.ncbi.nlm.nih.gov/pubmed/31525460
```

```
get("model_mouse_full", envir = .GlobalEnv)
```

4 progeny

progeny

Calculate PROGENy pathway scores from gene expression

Description

This function uses the linear model of pathway-responsive genes underlying the PROGENy method. It transforms a gene expression matrix with HGNC/MGI gene symbols in rows and sample names in columns into a pathway score matrix with samples in rows and pathways in columns.

Usage

```
progeny(
  expr,
  scale = TRUE,
  organism = "Human",
  top = 100,
  perm = 1,
  verbose = FALSE,
  z_scores = FALSE,
  get_nulldist = FALSE,
  assay_name = "RNA",
  return_assay = FALSE,
  ...
)
```

Arguments

expr	A gene	expression	object with	n HGNC/MGI	symbols	in rows and	samples in

columns. In order to run PROGENy in single-cell RNAseq data, it also accepts Seurat and SingleCellExperiment object, taking the normalized counts for the

computation.

scale A logical value indicating whether to scale the scores of each pathway to have

a mean of zero and a standard deviation of one. It does not apply if we use

permutations.

organism - "Human" or "Mouse"

top The top n genes for generating the model matrix according to significance (p-

value)

perm An interger detailing the number of permutations. No permutations by default

(1). When Permutations larger than 1, we compute progeny pathway scores and assesses their significance using a gene sampling-based permutation strategy,

for a series of experimental samples/contrasts.

verbose A logical value indicating whether to display a message about the number of

genes used per pathway to compute progeny scores (i.e. number of genes present

in the progeny model and in the expression dataset)

z_scores Only applies if the number of permutations is greater than 1. A logical value.

TRUE: the z-scores will be returned for the pathway activity estimations. FALSE:

the function returns a normalized z-score value between -1 and 1.

get_nulldist Only applies if the number of permutations is greater than 1. A logical value.

TRUE: the null distributions generated to assess the signifance of the pathways

scores is also returned.

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assay_name	Only applies if the input is a Seurat object. It selects the name of the assay on which Progeny will be run. Default to: RNA, i.e. normalized expression values.
return_assay	Only applies if the input is a Seurat object. A logical value indicating whether to return progeny results as a new assay called Progeny in the Seurat object used as input. Default to FALSE.
	Additional arguments to be passed to the functions.

Details

The publication of the method is available at: https://www.nature.com/articles/s41467-017-02391-6

The supplied expression object has to contain HGNC/MGI symbols in rows. This will, in most cases (and how we originally used it), be either normalized gene expression of a microarray experiment or log-transformed (and possible variance-stabilized) counts from an RNA-seq experiment.

The human and mouse model matrices consists of 14 pathways and large set of genes with an associated p-value (p-value per gene and pathway) that accounts for the importance of each gene on each pathway upon perturbation. Its coefficients are non-zero if the gene-pathway pair corresponds to the top N genes (100 by default) that were up-regulated upon stimulation of the pathway in a wide range of experiments. The value corresponds to the fitted z-score across experiments in our model fit. Only rows with at least one non-zero coefficient were included, as the rest is not used to infer pathway activity.

Value

A matrix with samples in rows and pathways in columns. In case we run the method with permutations and the option get_nulldist to TRUE, we will get a list with two elements. The first element is the matrix with the pathway activity as before. The second elements is the null distributions that we generate to assess the signifiance of the pathways scores.

See Also

progenyPerm

Examples

progenyPerm

Compute progeny pathway scores and assesses significance based on permutations

Description

Compute progeny pathway scores and assesses significance based on permutations

6 progenyScatter

Usage

```
progenyPerm(
   df,
   weight_matrix,
   k = 10000,
   z_scores = TRUE,
   get_nulldist = FALSE
)
```

Arguments

df A data.frame of n*m+1 dimension, where n is the number of omic features to be considered and m is the number of samples/contrasts. The first column should be the identifiers of the omic features. These identifiers must be coherent with the identifiers of the weight matrix. weight_matrix A progeny coefficient matrix. the first column should be the identifiers of the omic features and should be coherent with the identifiers provided in df. k The number of permutations to be performed to generate the null-distribution used to estimate the significance of progeny scores. The default value is 10000. z_scores if true, the z-scores will be returned for the pathway activity estimations. Else, the function returns a normalized z-score value between -1 and 1. if true, the null score distribution used for normalization will be returned along get_nulldist

Value

This function returns a list of two elements. The first element is a data frame of p*m+1 dimensions, where p is the number of progeny pathways, and m is the number of samples/contrasts. Each cell represents the significance of a progeny pathway score for one sample/contrast. The significance ranges between -1 and 1. The significance is equal to x*2-1, x being the quantile of the progeny pathway score with respect to the null distribution. Thus, this significance can be interpreted as the equivalent of 1-p.value two-sided test over an empirical distribution) with the sign indicating the direction of the regulation. The second element is the null distribution list (a null distribution is generated for each sample/contrast).

Examples

```
# use example gene expression matrix
gene_expression <- as.matrix(read.csv(system.file("extdata",
   "human_input.csv", package = "progeny"), row.names = 1))

# calculate pathway activities
progeny(gene_expression, scale=TRUE, organism="Human", top=100, perm=10000)</pre>
```

with the actual normalized score data frame.

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Description

This function generate a series of scatter plot with marginal distribution (in the form of an arrange-Grob object), for each progeny pathway and sample/contrast. Each scatter plot has progeny weights as x-axis and the gene level stat used to compute progeny score as the y-axis. The marginal distribution of the gene level stats is displayed on the right of the plot to give visual support of the significance of each gene contributing to the progeny pathway score. The green and red colors represent the positive and negative contribution of genes to the progeny pathway, respectively. For each gene contribution, 4 cases are possible, as the combinations of the sign of the gene level stat and the sign of the gene level weight. Positive weight will lead to a positive(green)/negative(red) gene contribution if the gene level stat is positive/negative. Negative weight will lead to a negative(red)/positive(green) gene contribution if the gene level stat is positive/negative.

Usage

```
progenyScatter(
   df,
   weight_matrix,
   dfID = 1,
   weightID = 1,
   statName = "gene stats",
   verbose = FALSE
)
```

Arguments

df	an n*m data frame, where n is the number of omic features (genes). m isn't really important, as long as at least one column corresponds to a sample or contrast statistic. One of the columns should correspond to the gene symbols.
weight_matrix	A progeny coefficient matrix. the first column should be the identifiers of the omic features, and should be coherent with the identifiers provided in df.
dfID	an integer corresponding to the column number of the gene identifiers of df.
weightID	an integer corresponding to the column number of the gene identifiers of the weight matrix.
statName	The name of the stat used, to be displayed on the plot
verbose	Logical indicating whether we want to have the messages indicating the different computed weights.

Value

The function returns a list of list of arrangeGrob objects. The first level list elements correspond to samples/contrasts. The second level correspond to pathways. The plots can be saved in a pdf format using the saveProgenyPlots function.

```
# use example gene expression matrix
gene_expression <- read.csv(system.file("extdata",
   "human_input.csv", package = "progeny"))
# getting a model matrix with 100 top significant genes and converting to df
weight_matrix <- getModel("Human", top=100)</pre>
```

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```
weight_matrix <- data.frame(names = row.names(weight_matrix),
   row.names = NULL, weight_matrix)

#use progenyScatter function
plots <- progenyScatter(gene_expression, weight_matrix)</pre>
```

saveProgenyPlots

Function to save Progeny plots

Description

This function is designed to save the plots (in pdf format) of a nested (2 level) list of arrangeGrob objects, such as the one returned by the progenyScatter function.

Usage

```
saveProgenyPlots(plots, contrast_names, dirpath)
```

Arguments

plots a list of list of arrangeGrob object (such as the one returned by the progenyScat-

ter function.). The first level list elements correspond to samples/contrasts. The second level corresponds to pathways. The plots can be saved in a pdf format

using the saveProgenyPlots function.

contrast_names a vector of the same length as the first level of the plot list corresponding to the

names of each sample/contrast

dirpath the path to the directory where the plots should be saved

Value

This function produces the pdf files of plots taken from the progenyScatter function

```
#create plots using progneyScatter function
gene_expression <- read.csv(system.file("extdata",
   "human_input.csv", package = "progeny"))

# getting a weight_matrix
weight_matrix <- getModel("Human", top=100)
weight_matrix <- data.frame(names = row.names(weight_matrix),
   row.names = NULL, weight_matrix)
plots <- progenyScatter(gene_expression, weight_matrix)

#create a list with contrast names
contrast_names <- names(gene_expression[2:ncol(gene_expression)])

#assign a path to store your plots
dirpath <- "./progeny_plots/"

# save it
# saveProgenyPlots(plots, contrast_names, dirpath)</pre>
```

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vignette_data

The RNA data used in the progeny vignette

Description

List with three elements: the gene counts, the experimental design and the result of limma differential analysis

Format

List with three elements: the gene counts, the experimental design and the result of limma differential analysis

counts gene counts

design experiemental design

limma_ttop differential analysis result using limma

Source

https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE119931

Examples

data("vignette_data")

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```