

Package ‘IsoBayes’

January 28, 2025

Type Package

Title IsoBayes: Single Isoform protein inference Method via Bayesian Analyses

Version 1.4.0

Description IsoBayes is a Bayesian method to perform inference on single protein isoforms.

Our approach infers the presence/absence of protein isoforms, and also estimates their abundance; additionally, it provides a measure of the uncertainty of these estimates, via:

- i) the posterior probability that a protein isoform is present in the sample;
- ii) a posterior credible interval of its abundance.

IsoBayes inputs liquid chromatography mass spectrometry (MS) data, and can work with both PSM counts, and intensities.

When available, transcript isoform abundances (i.e., TPMs) are also incorporated:

TPMs are used to formulate an informative prior for the respective protein isoform relative abundance.

We further identify isoforms where the relative abundance of proteins and transcripts significantly differ.

We use a two-

layer latent variable approach to model two sources of uncertainty typical of MS data:

- i) peptides may be erroneously detected (even when absent);
- ii) many peptides are compatible with multiple protein isoforms.

In the first layer, we sample the presence/absence of each peptide based on its estimated probability

of being mistakenly detected, also known as PEP (i.e., posterior error probability).

In the second layer, for peptides that were estimated as being present, we allocate their abundance across the protein isoforms they map to.

These two steps allow us to recover the presence and abundance of each protein isoform.

biocViews StatisticalMethod, Bayesian, Proteomics, MassSpectrometry, AlternativeSplicing, Sequencing, RNASeq, GeneExpression, Genetics, Visualization, Software

License GPL-3

Depends R (>= 4.3.0)

Imports methods, Rcpp, data.table, glue, stats, doParallel, parallel, doRNG, foreach, iterators, ggplot2, HDInterval, SummarizedExperiment, S4Vectors

LinkingTo Rcpp, RcppArmadillo

Suggests knitr, rmarkdown, testthat, BiocStyle

SystemRequirements C++17

VignetteBuilder knitr**RoxygenNote** 7.3.2**ByteCompile** true**URL** <https://github.com/SimoneTiberi/IsoBayes>**BugReports** <https://github.com/SimoneTiberi/IsoBayes/issues>**git_url** <https://git.bioconductor.org/packages/IsoBayes>**git_branch** RELEASE_3_20**git_last_commit** 8e3242d**git_last_commit_date** 2024-10-29**Repository** Bioconductor 3.20**Date/Publication** 2025-01-27**Author** Jordy Bollon [aut],Simone Tiberi [aut, cre] (<<https://orcid.org/0000-0002-3054-9964>>)**Maintainer** Simone Tiberi <simone.tiberi@unibo.it>

Contents

IsoBayes-package	2
generate_SE	3
inference	4
input_data	6
plot_relative_abundances	8
plot_traceplot	9

Index	10
--------------	-----------

IsoBayes-package	<i>IsoBayes: Single Isoform protein inference Method via Bayesian Analyses</i>
------------------	--

Description

IsoBayes is a Bayesian method to perform inference on single protein isoforms. Our approach infers the presence/absence of protein isoforms, and also estimates their abundance; additionally, it provides a measure of the uncertainty of these estimates, via: i) the posterior probability that a protein isoform is present in the sample; ii) a posterior credible interval of its abundance. IsoBayes inputs liquid chromatography mass spectrometry (MS) data, and can work with both PSM counts, and intensities. When available, transcript isoform abundances (i.e., TPMs) are also incorporated: TPMs are used to formulate an informative prior for the respective protein isoform relative abundance. We further identify isoforms where the relative abundance of proteins and transcripts significantly differ. We use a two-layer latent variable approach to model two sources of uncertainty typical of MS data: i) peptides may be erroneously detected (even when absent); ii) many peptides are compatible with multiple protein isoforms. In the first layer, we sample the presence/absence of each peptide based on its estimated probability of being mistakenly detected, also known as PEP (i.e., posterior error probability). In the second layer, for peptides that were estimated as being present, we allocate their abundance across the protein isoforms they map to. These two steps allow us to recover the presence and abundance of each protein isoform.

Details

The DESCRIPTION file: This package was not yet installed at build time.

Questions relative to IsoBayes should be reported as a new issue at <https://github.com/SimoneTiberi/IsoBayes/issues>.

To access the vignettes, type: `browseVignettes("IsoBayes")`.

Index: This package was not yet installed at build time.

Author(s)

Jordy Bollon <jordy.bollon@iit.it>, Simone Tiberi <simone.tiberi@unibo.it>

generate_SE

Generate SummarizedExperiment object

Description

generate_SE converts the input files, required to run IsoBayes, into a SummarizedExperiment object. This object should then be passed to `input_data` function.

Usage

```
generate_SE(
  path_to_peptides_psm = NULL,
  path_to_peptides_intensities = NULL,
  input_type = NULL,
  abundance_type = NULL,
  PEP = TRUE,
  FDR_thd = 0.01
)
```

Arguments

`path_to_peptides_psm` a character string indicating the path to one of the following files:

- i) the psmtsv file from *MetaMorpheus* tool with PSM counts,
- ii) the idXML file from *OpenMS* toolkit, or
- iii) a `data.frame` or a path to a tsv file, formatted as explained in the "Input user-provided data" Section of the vignettes (only when `input_type = "other"`).

`path_to_peptides_intensities` (optional) a character string indicating the path to the psmtsv file from *MetaMorpheus* with intensity values. Required if 'abundance_type' equals to "intensities" and `input_type` equals to "metamorpheus".

`input_type` a character string indicating the tool used to obtain the peptides file: "metamorpheus", "openMS" or "other".

`abundance_type` a character string indicating the type of input: "psm" or "intensities".

`PEP` logical; if TRUE (default), the algorithm will account for the probability that peptides are erroneously detected. If FALSE, PEP is ignored. We suggest using PEP with a weak FDR threshold of 0.1 (default parameters options). This is because peptides with $FDR > 0.1$ are usually unreliable, and associated to high error probabilities (e.g., $PEP > 0.9$).

FDR_thd a numeric value indicating the False Discovery Rate threshold to be used to discard unreliable peptides.

Value

A SummarizedExperiment object.

Author(s)

Jordy Bollon <jordy.bollon@iit.it> and Simone Tiberi <simone.tiberi@unibo.it>

See Also

[input_data](#)

Examples

```
# Load internal data to the package:
data_dir = system.file("extdata", package = "IsoBayes")

# Define the path to the AllPeptides.psmtsv file returned by *MetaMorpheus* tool
path_to_peptides_psm = paste0(data_dir, "/AllPeptides.psmtsv")

# Generate a SummarizedExperiment object
SE = generate_SE(path_to_peptides_psm = path_to_peptides_psm,
                 abundance_type = "psm",
                 input_type = "metamorpheus"
                 )

# For more examples see the vignettes:
# browseVignettes("IsoBayes")
```

inference

Run our two-layer latent variable Bayesian model

Description

inference runs our two-layer latent variable Bayesian model, taking as input the data created by [input_data](#).

Usage

```
inference(
  loaded_data,
  map_iso_gene = NULL,
  n_cores = 1,
  K = 2000,
  burn_in = 1000,
  thin = 1,
  traceplot = FALSE
)
```

Arguments

<code>loaded_data</code>	list of <code>data.frame</code> objects, returned by input_data .
<code>map_iso_gene</code>	(optional) a character string (indicating the path to a csv file with 2 columns), or a <code>data.frame</code> with 2 columns. In both cases, the 1st column must contain the isoform name/id, while the 2nd column has the gene name/id. This argument is required to return protein isoform relative abundances, normalized within each gene (i.e., adding to 1 within a gene), to plot results via plot_relative_abundances , and to return protein abundances aggregated by gene with HPD credible interval.
<code>n_cores</code>	the number of cores to use during algorithm execution. We suggest increasing the number of threads for large datasets only.
<code>K</code>	the number of MCMC iterations. Minimum 2000.
<code>burn_in</code>	the number of initial iterations to discard. Minimum 1000.
<code>thin</code>	thinning value to apply to the final MCMC chain. Useful for decreasing the memory (RAM) usage.
<code>traceplot</code>	a logical value indicating whether to return the posterior chain of the relative abundances of each protein isoform (i.e., "PI"). If TRUE, the posterior chains are stored in 'MCMC' object, and can be plotted via 'plot_traceplot' function.

Value

A list of three `data.frame` objects: 'isoform_results', and (only if 'map_iso_gene' is provided) 'normalized_isoform_results' (relative abundances normalized within each gene) and 'gene_abundance'. For more information about the results stored in the three `data.frame` objects, see the vignettes: `#browseVignettes("IsoBayes")`

Author(s)

Jordy Bollon <jordy.bollon@iit.it> and Simone Tiberi <simone.tiberi@unibo.it>

See Also

[input_data](#) and [plot_relative_abundances](#)

Examples

```
# Load internal data to the package:
data_dir = system.file("extdata", package = "IsoBayes")

# Define the path to the AllPeptides.psmtsv file returned by MetaMorpheus tool
path_to_peptides_psm = paste0(data_dir, "/AllPeptides.psmtsv")

# Generate a SummarizedExperiment object
SE = generate_SE(path_to_peptides_psm = path_to_peptides_psm,
                abundance_type = "psm",
                input_type = "metamorpheus"
                )
# Define the path to the jurkat_isoform_kallisto.tsv with mRNA relative abundance
tpm_path = paste0(data_dir, "/jurkat_isoform_kallisto.tsv")

# Load and process SE object
data_loaded = input_data(SE, path_to_tpm = tpm_path)
```

```

# Define the path to the map_iso_gene.csv file.
# Alternatively a data.frame can be used (see documentation).
path_to_map_iso_gene = paste0(data_dir, "/map_iso_gene.csv")

# Run the algorithm
set.seed(169612)
results = inference(data_loaded, map_iso_gene = path_to_map_iso_gene, traceplot = TRUE)

# Results is a list of 3 data.frames:
names(results)

# Main results:
head(results$isoform_results)

# Results normalized within genes
# (relative abundances add to 1 within each gene):
# useful to study alternative splicing within genes:
head(results$normalized_isoform_results)

# Gene abundance
head(results$gene_abundance)

# results normalized within genes (total abundance of each gene),
# useful to study alternative splicing within genes:
head(results$normalized_isoform_results)

# Plotting results, normalizing within genes
# (relative abundances add to 1 within each gene):
plot_relative_abundances(results,
  gene_id = "TUBB",
  normalize_gene = TRUE)

# Plotting results, NOT normalized
# (relative abundances add to 1 across all isoforms in the dataset):
plot_relative_abundances(results,
  gene_id = "TUBB",
  normalize_gene = FALSE)

# Visualize MCMC chain for isoforms "TUBB-205", "TUBB-206", and "TUBB-208"
# To visualize traceplots, set "traceplot" to TRUE when running "inference" function
plot_traceplot(results, "TUBB-205")
plot_traceplot(results, "TUBB-206")
plot_traceplot(results, "TUBB-208")

# For more examples see the vignettes:
# browseVignettes("IsoBayes")

```

input_data

Load and process input data

Description

input_data reads and processes a SummarizedExperiment object collecting input data and meta-data required to run IsoBayes model.

Usage

```
input_data(SE, path_to_tpm = NULL)
```

Arguments

SE	a SummarizedExperiment object created by generate_SE function. Alternatively, this object can be created by the user, following the structure specified in the "Input user-provided data" Section of the vignettes
path_to_tpm	(optional) a data.frame object or a character string indicating the path to a tsv file with mRNA isoform TPMs. The tsv file must have 1 row per isoform, and 2 columns: i) 'isoname': a character string indicating the isoform name; ii) 'tpm': a numeric variable indicating the Transcripts Per Million (TPM) count. Column names must be 'isoname' and 'tpm'.

Value

A list of data.frame objects, with the data needed to run [inference](#) function.

Author(s)

Jordy Bollon <jordy.bollon@iit.it> and Simone Tiberi <simone.tiberi@unibo.it>

See Also

[generate_SE](#), [inference](#)

Examples

```
# Load internal data to the package:
data_dir = system.file("extdata", package = "IsoBayes")

# Define the path to the AllPeptides.psmtsv file returned by *MetaMorpheus* tool
path_to_peptides_psm = paste0(data_dir, "/AllPeptides.psmtsv")

# Generate a SummarizedExperiment object
SE = generate_SE(path_to_peptides_psm = path_to_peptides_psm,
                 abundance_type = "psm",
                 input_type = "metamorpheus"
                )

# Load and process SE object
data_loaded = input_data(SE)

# For more examples see the vignettes:
# browseVignettes("IsoBayes")
```

plot_relative_abundances

Plot isoform results

Description

plot_relative_abundances plots protein isoforms results, obtained by [inference](#), for a specific gene, together with transcripts abundances if available.

Usage

```
plot_relative_abundances(  
  res_inference,  
  gene_id,  
  plot_CI = TRUE,  
  normalize_gene = TRUE  
)
```

Arguments

`res_inference` list of two data.frame objects returned by [inference](#).

`gene_id` a character string indicating the gene to be plotted.

`plot_CI` logical; if TRUE (default), plot 0.95 level Credibility Intervals for each isoform.

`normalize_gene` logical; if TRUE (default), plot isoform relative abundances, normalized within the specified gene (they add to 1 within a gene).

Value

A ggplot object, showing isoform relative abundances for a specific gene.

Author(s)

Jordy Bollon <jordy.bollon@iit.it> and Simone Tiberi <simone.tiberi@unibo.it>

See Also

[inference](#)

Examples

```
# see the example of inference function:  
help(inference)
```

plot_traceplot	<i>Traceplot of the (thinned) posterior chain of the relative abundance of each protein isoform (i.e., pi).</i>
----------------	---

Description

plot_traceplot plots the traceplot of the (thinned) posterior chain of the relative abundance of each protein isoform (i.e., pi). The vertical grey dashed line indicates the burn-in (the iterations on the left side of the burn-in are discarded in posterior analyses).

Usage

```
plot_traceplot(results, protein_id)
```

Arguments

results	a list of data.frame objects, computed via inference .
protein_id	a character, indicating the protein isoform to plot.

Value

A gtable object.

Author(s)

Simone Tiberi <simone.tiberi@unibo.it>

See Also

[inference](#)

Examples

```
# see the example of inference function:  
help(inference)
```

Index

* **package**

IsoBayes-package, [2](#)

data.frame, [9](#)

generate_SE, [3](#), [7](#)

inference, [4](#), [7–9](#)

input_data, [3–5](#), [6](#)

IsoBayes (IsoBayes-package), [2](#)

IsoBayes-package, [2](#)

plot_relative_abundances, [5](#), [8](#)

plot_traceplot, [9](#)