

Package ‘cfdnakit’

November 20, 2024

Title Fragmen-length analysis package from high-throughput sequencing of cell-free DNA (cfDNA)

Version 1.5.0

Description This package provides basic functions for analyzing shallow whole-genome sequencing (~0.3X or more) of cell-free DNA (cfDNA). The package basically extracts the length of cfDNA fragments and aids the visualization of fragment-length information. The package also extract fragment-length information per non-overlapping fixed-sized bins and used it for calculating ctDNA estimation score (CES).

License GPL-3

Encoding UTF-8

LazyData FALSE

Roxygen list(markdown = TRUE)

RoxygenNote 7.2.3

biocViews CopyNumberVariation, Sequencing, WholeGenome

BugReports <https://github.com/Pitithat-pu/cfdnakit/issues>

Imports Biobase, dplyr, GenomicRanges, GenomeInfoDb, ggplot2, IRanges, magrittr, PSCBS, QDNAseq, Rsamtools, utils, S4Vectors, stats, rlang

Depends R (>= 4.3)

Suggests rmarkdown, knitr, roxygen2, BiocStyle

VignetteBuilder knitr

git_url <https://git.bioconductor.org/packages/cfdnakit>

git_branch devel

git_last_commit 889853b

git_last_commit_date 2024-10-29

Repository Bioconductor 3.21

Date/Publication 2024-11-20

Author Pitithat Puranachot [aut, cre] (ORCID:
<<https://orcid.org/0000-0002-6786-9240>>)

Maintainer Pitithat Puranachot <pitithat@gmail.com>

Contents

cfdnakit-package	2
calculate_CES_score	4
call_cnv	5
create_blacklist_gr	6
create_PoN	6
extract_insert_size	7
filter_read_on_blacklist	8
fragment_dist	8
get_fragment_profile	9
get_segment_byresolution	10
get_solution_table	10
get_zscore_profile	11
GRCh2UCSCGRanges	12
if_exist_baifile	12
if_ucsc_chrformat	13
make_density_table	13
overlap_bin_with_segment	14
plot_cnv_solution	14
plot_distance_matrix	15
plot_fragment_dist	16
plot_sl_ratio	17
plot_transformed_sl	17
read_bamfile	18
read_PoN_files	19
segmentByPSCB	20
test_ysize_KolmogorovSmirnov	21
UCSC2GRChSampleBam	21
util.bias_correct	22
zscore_transform	22
%>%	23
Index	24

cfdnakit-package	<i>Fragmen-length analysis package from high-throughput sequencing of cell-free DNA (cfDNA)</i>
------------------	---

Description

This package provides basic functions for analyzing shallow whole-genome sequencing (~0.3X or more) of cell-free DNA (cfDNA). The package basically extracts the length of cfDNA fragments and aids the visualization of fragment-length information. The package also extract fragment-length information per non-overlapping fixed-sized bins and used it for calculating ctDNA estimation score (CES).

Details

This package provides functions for analyzing using shallow whole-genome sequencing data (~0.3X or more) of circulating cell-free DNA (cfDNA). The aim is to estimate circulating tumor DNA using its characteristic short-fragmented cfDNA. The package extracts length of each cfDNA and assists the visualization of fragment-length distribution. A short-fragment ratio is calculated per non-overlapping fixed-sized bins. Genome-wide copy-number alteration is estimated by the short-fragmented cfDNA. The ctDNA estimation score (CES) comprehensively estimates the circulating tumor DNA based on the short-fragment analysis.

Author(s)

Dr. rer. nat. Pitithat Puranachot

Examples

```
library(cfdnakit)
## Reading in a bamfile
sample_bamfile = system.file("extdata",
                             "ex.plasma.bam",
                             package = "cfdnakit")
plasma_SampleBam = read_bamfile(sample_bamfile,
                               apply_blacklist = FALSE)

## Plot a fragment-length distribution of a sample
plot_fragment_dist(list("Plasma.Sample"=plasma_SampleBam))

## Plot a fragment-length distribution of two samples
control_RDS_file =
  system.file("extdata", "BH01_CHR15.SampleBam.rds",
             package = "cfdnakit")
### Load example SampleBam of Healthy cfDNA
control_bins =
  readRDS(control_RDS_file)

comparing_list = list("Healthy.cfDNA"=control_bins,
                     "Patient.1"=plasma_SampleBam)
plot_fragment_dist(comparing_list)

## Derived and plot genome-wide short-fragment cfDNA
patient.SampleFragment =
  get_fragment_profile(plasma_SampleBam,
                     sample_id = "Patient.1")
plot_sl_ratio(patient.SampleFragment)

## Derived and plot normalized short-fragment cfDNA
PoN_rdsfile = system.file(
  "extdata",
  "ex.PoN.rds",
  package = "cfdnakit")
```

```

        ## Loading example PoN data
PoN.profiles = readRDS(PoN_rdsfile)

sample_zscore =
  get_zscore_profile(patient.SampleFragment,
                    PoN.profiles)
sample_zscore_segment = segmentByPSCB(sample_zscore)
plot_transformed_sl(sample_zscore, sample_zscore_segment)

## Estimate circulating tumor DNA
calculate_CES_score(sample_zscore_segment)

```

calculate_CES_score *Calculate CES Score from Segmentation*

Description

Calculate CES Score from Segmentation

Usage

```
calculate_CES_score(sample_segmentation)
```

Arguments

```
sample_segmentation
  Segmentation Dataframe
```

Value

Numeric; CES score

Examples

```

### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")

sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)

sample_zscore_segment <- segmentByPSCB(sample_zscore)

calculate_CES_score(sample_zscore_segment)

```

call_cnv	<i>Call Copy-number Variation from SLRatio and segmentation</i>
----------	---

Description

Call Copy-number Variation from SLRatio and segmentation

Usage

```
call_cnv(
  sample_segmentation,
  sample_zscore,
  callChr = seq_len(22),
  tfs = c(0, 0.7),
  ploidy = c(1.5, 3),
  MaxCN = 4
)
```

Arguments

sample_segmentation	segmentation dataframe from segmentByPSCBS
sample_zscore	zscore dataframe
callChr	chromosome to analysis : Default c(1:22)
tfs	range of fitting tumor fraction : Default c(0,0.8)
ploidy	range of fitting chromosomal ploidy : Default c(1.5,4)
MaxCN	maximum copy-number : Default 4

Value

List of cnvcalling solutions

Examples

```
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")

sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)

sample_zscore_segment <- segmentByPSCB(sample_zscore)

sample_cnv <- call_cnv(sample_zscore_segment, sample_zscore, tfs=c(0.1,0.3), ploidy=c(1.5,2), MaxCN=3)
```

```
plot_cnv_solution(sample_cnv, selected_solution = 1)
```

create_blacklist_gr *Create Blacklist regions GRanges object*

Description

Create Blacklist regions GRanges object

Usage

```
create_blacklist_gr(blacklist_files)
```

Arguments

blacklist_files
Character; Filepath to file containing blacklist regions

Value

GRanges object of blacklist regions

create_PoN *Create Panel-of-Normal (PoN) object*

Description

Create Panel-of-Normal (PoN) object

Usage

```
create_PoN(list_rdsfiles)
```

Arguments

list_rdsfiles Character; a file contains paths to Profile.Rdata per line

Value

Null

Examples

```
healthy.1 <- system.file("extdata","ex.healthy1.rds",package = "cfdnakit")
healthy.2 <- system.file("extdata","ex.healthy2.rds",package = "cfdnakit")

path_to_PoN_txt <- paste0(system.file("extdata",package = "cfdnakit"),"/temp.reference_healthy.listfile")
fileConn<-file(path_to_PoN_txt)
writeLines(c(healthy.1,healthy.2), fileConn)
close(fileConn)

PoN.profiles <- create_PoN(path_to_PoN_txt)
file.remove(path_to_PoN_txt)
```

extract_insert_size *Extract Insert size from SampleBam*

Description

Extract Insert size from SampleBam

Usage

```
extract_insert_size(readbam_bin, maximum_length = 600, minimum_length = 20)
```

Arguments

readbam_bin SampleBam Object

maximum_length Int; Maximum length of fragment. cfDNA fragment longer than this value will not be considered; Default 600

minimum_length Int; Minimum length of fragment. cfDNA fragment shorter than this value will not be considered; Default 20

Value

Numeric Vector; Insert size of given sample

Examples

```
### Loading example SampleBam file
example_file <- system.file("extdata","example_patientcfDNA_SampleBam.RDS",package = "cfdnakit")
sample_bambin <- readRDS(example_file)
extract_insert_size(sample_bambin)
### Extract only insert size of fragment having specific size
extract_insert_size(sample_bambin,maximum_length=500, minimum_length = 50)
```

filter_read_on_blacklist

Filter out reads on blacklist regions

Description

Filter out reads on blacklist regions

Usage

```
filter_read_on_blacklist(sample_bin, blacklist_files = NULL, genome = "hg19")
```

Arguments

sample_bin	SampleBam; Object from function read_bamfile
blacklist_files	Character; Filepath to file containing blacklist regions
genome	Character; Abbreviation of reference genome; Either hg19 or mm10. default:hg19

Value

SampleBam after filtering out read on balck list regions

fragment_dist

Get insert-size distribution table

Description

Get insert-size distribution table

Usage

```
fragment_dist(readbam_bin, maximum_length = 600, minimum_length = 20)
```

Arguments

readbam_bin	SampleBam Object from function read_bamfile
maximum_length	Int; Maximum length of fragment. cfDNA fragment longer than this value will not be considered; Default 600
minimum_length	Int; Minimum length of fragment. cfDNA fragment shorter than this value will not be considered; Default 20

Value

Distribution table of fragment length

get_fragment_profile *Getting fragment-length information*

Description

Getting fragment-length information

Usage

```
get_fragment_profile(  
  readbam_bin,  
  sample_id,  
  genome = "hg19",  
  short_range = c(100, 150),  
  long_range = c(151, 250),  
  maximum_length = 600,  
  minimum_length = 20  
)
```

Arguments

readbam_bin	SampleBam Object
sample_id	Character; Given sample ID
genome	abbreviation of reference genome; namely hg19, mm10. default:hg19
short_range	Vector of 2 Int; Range of fragment length to be defined as short fragment; Default c(100,150)
long_range	Vector of 2 Int; Range of fragment length to be defined as long fragment; Default c(151,250)
maximum_length	Int; Maximum length of fragment. cfDNA fragment longer than this value will not be considered; Default 600
minimum_length	Int; Minimum length of fragment. cfDNA fragment shorter than this value will not be considered; Default 20

Value

SampleFragment Object; Fragment length information for quality check and downstream analysis per bin and summary of sample

Examples

```
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfDNAkit")  
sample_bam_bin <- readRDS(example_file)  
sample_profile <- get_fragment_profile(sample_bam_bin, sample_id = "Patient1")
```

```
get_segment_bysolution
```

Return CNV segmentation result from given all CNV solutions

Description

Return CNV segmentation result from given all CNV solutions

Usage

```
get_segment_bysolution(solution, sample_segmentation, SL_distance_df)
```

Arguments

solution solution dataframe
 sample_segmentation Segmentation dataframe
 SL_distance_df Distance matrix

Value

list of segmentation per solution

```
get_solution_table    Get summarised table of cnv solutions
```

Description

Get summarised table of cnv solutions

Usage

```
get_solution_table(cnv_solutions)
```

Arguments

cnv_solutions cnvcalling result from function call_cnv.R

Value

Dataframe of solution table

Examples

```

#'
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")

sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)

sample_zscore_segment <- segmentByPSCB(sample_zscore)

sample_cnv <- call_cnv(sample_zscore_segment, sample_zscore, tfs=c(0.1,0.3), ploidy=c(1.5,2), MaxCN=3)
get_solution_table(sample_cnv)

```

get_zscore_profile *Transform SLRatio with PoN Fragment profile*

Description

Transform SLRatio with PoN Fragment profile

Usage

```
get_zscore_profile(fragment_profile, pon_profile)
```

Arguments

```

fragment_profile
                Sample Profile
pon_profile     PoN Profiles

```

Value

Dataframe of robust transformed SLratio

Examples

```

### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)

### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")

```

```
sample_zscore <- get_zscore_profile(sample_profile,pon_profiles)
sample_zscore_segment <- segmentByPSCB(sample_zscore)
```

GRCh2UCSCGRanges *Convert GRCh chromosome format to UCSC style*

Description

Convert GRCh chromosome format to UCSC style

Usage

```
GRCh2UCSCGRanges(which)
```

Arguments

which GRanges object;

Value

GRanges; GRanges after chromosome format conversion

if_exist_baifile *Check if bai file exist from given bam*

Description

Check if bai file exist from given bam

Usage

```
if_exist_baifile(bamfile)
```

Arguments

bamfile Character; Path to sample bamfile

Value

Boolean if the bai file exist

if_ucsc_chrformat	<i>Check UCSC chromosomes format for input bam file</i>
-------------------	---

Description

Check UCSC chromosomes format for input bam file

Usage

```
if_ucsc_chrformat(bamfile_path)
```

Arguments

bamfile_path Character; Path to sample bamfile

Value

Boolean; if the input bam file is UCSC format, chr prefix

make_density_table	<i>Make Fragment-length density table</i>
--------------------	---

Description

Make Fragment-length density table

Usage

```
make_density_table(readbam_bin, minimum_length, maximum_length)
```

Arguments

readbam_bin List; A list containing SampleBam object/objects from the read_bamfile function

minimum_length numeric;

maximum_length numeric

Value

data.frame

overlap_bin_with_segment
Overlap and merge bin data frame with segmentation dataframe

Description

Overlap and merge bin data frame with segmentation dataframe

Usage

```
overlap_bin_with_segment(per_bin_profile, sample_segmentation)
```

Arguments

```
per_bin_profile      bin dataframe
sample_segmentation  segmentation dataframe
```

Value

dataframe of overlapping bin and segmentation

plot_cnv_solution *Plot Fragment-length profile with CNV calling result*

Description

Plot Fragment-length profile with CNV calling result

Usage

```
plot_cnv_solution(
  cnvcall,
  selected_solution = 1,
  genome = "hg19",
  ylim = c(-30, 30)
)
```

Arguments

```
cnvcall      solution results from call_cnv function
selected_solution  solution rank to plot
genome       Character; version of reference genome (default hg19)
ylim         Vector of 2 Int; ylim of plot (default c(-20,20))
```

Value

ggplot object plot Genomics CNV profile of selected solution

Examples

```
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")

sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)
sample_zscore_segment <- segmentByPSCB(sample_zscore)

sample_cnv <- call_cnv(sample_zscore_segment, sample_zscore, tfs=c(0.1,0.3), ploidies=c(1.5,2), MaxCN=3)
plot_cnv_solution(sample_cnv, selected_solution = 1)
```

plot_distance_matrix *Plot Distance Matrix from CNVCalling*

Description

Plot Distance Matrix from CNVCalling

Usage

```
plot_distance_matrix(cnvcall)
```

Arguments

cnvcall cnvcalling result from function call_cnv.R

Value

ggplot object ; distance matrix per cnvcalling solution

Examples

```
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")
```

```

sample_zscore <- get_zscore_profile(sample_profile,pon_profiles)
sample_zscore_segment <- segmentByPSCB(sample_zscore)

sample_cnv <- call_cnv(sample_zscore_segment,sample_zscore, tfs=c(0.1,0.3),ploidies=c(1.5,2), MaxCN=3)
plot_distance_matrix(sample_cnv)

```

plot_fragment_dist *Plot Fragment-length Distribution*

Description

Plot Fragment-length Distribution

Usage

```
plot_fragment_dist(readbam_list, maximum_length = 550, minimum_length = 20)
```

Arguments

readbam_list	List; A list containing SampleBam object/objects from the read_bamfile function
maximum_length	Int; Maximum length of fragment. cfDNA fragment longer than this value will not be considered; Default 550
minimum_length	Int; Minimum length of fragment. cfDNA fragment shorter than this value will not be considered; Default 20

Value

distribution plot

Examples

```

example_file <- system.file("extdata","example_patientcfDNA_SampleBam.RDS",package = "cfdnakit")
sample_bambin <- readRDS(example_file)

### adding more samples to the plot
example_file2 <- system.file("extdata","BH01_CHR15.SampleBam.rds",package = "cfdnakit")
control_bambin <- readRDS(example_file2)
readbam_list <- list(plasma1 = sample_bambin, Healthy.blood.plasma=control_bambin)
plot_fragment_dist(readbam_list)

```

plot_sl_ratio *Plot Short/Long-fragment Ratio*

Description

Plot Short/Long-fragment Ratio

Usage

```
plot_sl_ratio(fragment_profile, ylim = c(0, 0.4), genome = "hg19")
```

Arguments

fragment_profile	list
ylim	plot y-axis limit
genome	Character; version of reference genome (default hg19)

Value

plot

Examples

```
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bam_bin <- readRDS(example_file)
sample_profile <- get_fragment_profile(sample_bam_bin, sample_id = "Patient1")
plot_sl_ratio(fragment_profile = sample_profile)

### change plot y-axis
plot_sl_ratio(fragment_profile = sample_profile, ylim=c(0.1,0.5))

### change reference genome
plot_sl_ratio(fragment_profile = sample_profile, genome="hg38")
```

plot_transformed_sl *Plot z-transformed Short/Long-fragment Ratio*

Description

Plot z-transformed Short/Long-fragment Ratio

Usage

```
plot_transformed_sl(
  sample_transformed_sl,
  sample_segment_df = NULL,
  ylim = c(-30, 30),
  genome = "hg19"
)
```

Arguments

```
sample_transformed_sl      Dataframe z-transformed SLRatio from get_zscore_profile
sample_segment_df         Dataframe segmenation from segmentByPSCB
ylim                       plot y-axis limit
genome                     Character; version of reference genome (default hg19)
```

Value

Genome-wide plot of z-transformed SLRatio

Examples

```
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")

sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)
sample_zscore_segment <- segmentByPSCB(sample_zscore)
plot_transformed_sl(sample_zscore, sample_zscore_segment)
## Change reference genome
plot_transformed_sl(sample_zscore, sample_zscore_segment, genome="hg38")
```

read_bamfile	<i>Read a bam file Read a bam file from give path. Alignment and sequencing read information will be binned into non-overlapping size</i>
--------------	---

Description

Read a bam file Read a bam file from give path. Alignment and sequencing read information will be binned into non-overlapping size

Usage

```
read_bamfile(
  bamfile_path,
  binsize = 1000,
  blacklist_files = NULL,
  genome = "hg19",
  target_bedfile = NULL,
  min_mapq = 20,
  apply_blacklist = TRUE
)
```

Arguments

bamfile_path	Character; Path to sample bamfile
binsize	Int; Size of non-overlapping windows in KB. Only 100,500 and 1000 is available; Default 1000
blacklist_files	Character; Filepath to file containing blacklist regions
genome	Character; abbreviation of reference genome; available genome: hg19,hg38, mm10. default:hg19
target_bedfile	Character; Path to exon/target bedfile; Default NULL
min_mapq	Int; minimum read mapping quality; Default 20
apply_blacklist	Logical; To exclude read on the blacklist regions Default TRUE

Value

SampleBam Object; A list object containing read information from the BAM file.

Examples

```
f1 <- system.file("extdata","ex.plasma.bam",package = "cfdnakit")
### read bam file with default params (hg19, 1000K binsize)
sample.bam <-read_bamfile(f1, apply_blacklist=FALSE)
```

read_PoN_files	<i>Read Fragment Profile from a list of rds file</i>
----------------	--

Description

Read Fragment Profile from a list of rds file

Usage

```
read_PoN_files(list_rdsfiles)
```

Arguments

list_rdsfiles path to file containing list of rds file

Value

list containing content of rds file

segmentByPSCB	<i>Segmentation data with PSCBS</i>
---------------	-------------------------------------

Description

Segmentation data with PSCBS

Usage

```
segmentByPSCB(sample_transformed_sl)
```

Arguments

sample_transformed_sl
dataframe of z-transformed SLRatio

Value

Dataframe of segmentation result

Examples

```
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")

sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)
sample_zscore_segment <- segmentByPSCB(sample_zscore)
```

```
test_ysize_KolmogorovSmirnov
      KolmogorovSmirnov test for insert size
```

Description

KolmogorovSmirnov test for insert size

Usage

```
test_ysize_KolmogorovSmirnov(control_insert_size, sample_insert_size)
```

Arguments

```
control_insert_size
      Vector of insert size of a control sample
sample_insert_size
      Vector of insert size of a testing sample
```

Value

KS.Test result

Examples

```
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
control_rds <- "BH01_CHR15.SampleBam.rds"
control_RDS_file <- system.file("extdata", control_rds, package = "cfdnakit")
control_fragment_profile <- readRDS(control_RDS_file)
sample.isize <- extract_insert_size(sample_bambin)
healthy.isize <- extract_insert_size(control_fragment_profile)
test_ysize_KolmogorovSmirnov(sample.isize, healthy.isize)
```

```
UCSC2GRChSampleBam      Convert UCSC chromosome format to GRCh style from a list of alignment information
```

Description

Convert UCSC chromosome format to GRCh style from a list of alignment information

Usage

```
UCSC2GRChSampleBam(sample.bam)
```

Arguments

sample.bam list of alignment information from function read_bamfile

Value

List; list of alignment information after conversion

util.bias_correct *Correct GC Bias readcount*

Description

Correct GC Bias readcount

Usage

```
util.bias_correct(readcount, bias)
```

Arguments

readcount numeric
bias numeric

Value

numeric

zscore_transform *zscore_transform transforms SLRatio profile into z-score*

Description

zscore_transform transforms SLRatio profile into z-score

Usage

```
zscore_transform(per_bin_profile)
```

Arguments

per_bin_profile
SampleFragment from function get_fragment_profile

Value

dataframe of z-score per bin

`%>%`*Pipe operator*

Description

See `magrittr::%>%` for details.

Arguments

<code>lhs</code>	A value or the <code>magrittr</code> placeholder.
<code>rhs</code>	A function call using the <code>magrittr</code> semantics.

Value

The result of calling `rhs(lhs)`.

Index

- * **internal**
 - %>%, [23](#)
- * **package cf-DNA**
 - cfdnakit-package, [2](#)
 - %>%, [23](#), [23](#)
- calculate_CES_score, [4](#)
- call_cnv, [5](#)
- cfdnakit (cfdnakit-package), [2](#)
- cfdnakit-package, [2](#)
- create_blacklist_gr, [6](#)
- create_PoN, [6](#)

- extract_insert_size, [7](#)

- filter_read_on_blacklist, [8](#)
- fragment_dist, [8](#)

- get_fragment_profile, [9](#)
- get_segment_byresolution, [10](#)
- get_solution_table, [10](#)
- get_zscore_profile, [11](#)
- GRCh2UCSCGRanges, [12](#)

- if_exist_baifile, [12](#)
- if_ucsc_chrformat, [13](#)

- make_density_table, [13](#)

- overlap_bin_with_segment, [14](#)

- plot_cnv_solution, [14](#)
- plot_distance_matrix, [15](#)
- plot_fragment_dist, [16](#)
- plot_sl_ratio, [17](#)
- plot_transformed_sl, [17](#)

- read_bamfile, [18](#)
- read_PoN_files, [19](#)

- segmentByPSCB, [20](#)

- test_ysize_KolmogorovSmirnov, [21](#)

- UCSC2GRChSampleBam, [21](#)
- util.bias_correct, [22](#)

- zscore_transform, [22](#)