

MSnbase development

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Abstract

This vignette describes the classes implemented in *MSnbase* package. It is intended as a starting point for developers or users who would like to learn more or further develop/extend mass spectrometry and proteomics data structures.

Contents

| | | |
|------|--|----|
| 1 | Introduction | 4 |
| 2 | <i>MSnbase</i> classes | 4 |
| 2.1 | <i>pSet</i> : a virtual class for raw mass spectrometry data and meta data | 4 |
| 2.2 | <i>MSnExp</i> : a class for MS experiments | 5 |
| 2.3 | <i>OnDiskMSnExp</i> : a on-disk implementation of the <i>MSnExp</i> class | 6 |
| 2.4 | <i>MSnSet</i> : a class for quantitative proteomics data 8 | |
| 2.5 | <i>MSnProcess</i> : a class for logging processing meta data. | 10 |
| 2.6 | <i>MIAPE</i> : Minimum Information About a Proteomics Experiment | 11 |
| 2.7 | <i>Spectrum et al.</i> : classes for MS spectra | 12 |
| 2.8 | <i>ReporterIons</i> : a class for isobaric tags. | 14 |
| 2.9 | <i>NAnnotatedDataFrame</i> : multiplexed <i>Annotated-DataFrames</i> | 15 |
| 2.10 | Other classes | 16 |
| 3 | Miscellaneous | 16 |
| 4 | Session information | 17 |

Foreword

MSnbase is under active development; current functionality is evolving and new features will be added. This software is free and open-source software. If you use it, please support the project by citing it in publications:

Laurent Gatto and Kathryn S. Lilley. *MSnbase - an R/Bioconductor package for isobaric tagged mass spectrometry data visualization, processing and quantitation*. *Bioinformatics* 28, 288-289 (2011).

Questions and bugs

You are welcome to contact me directly about *MSnbase*. For bugs, typos, suggestions or other questions, please file an issue in our tracking system (<https://github.com/lgatto/MSnbase/issues>) providing as much information as possible, a reproducible example and the output of `sessionInfo()`.

If you wish to reach a broader audience for general questions about proteomics analysis using R, you may want to use the Bioconductor support site: <https://support.bioconductor.org/>.

1 Introduction

This document is not a replacement for the individual manual pages, that document the slots of the *MSnbase* classes. It is a centralised high-level description of the package design.

MSnbase aims at being compatible with the *Biobase* infrastructure [1]. Many meta data structures that are used in *eSet* and associated classes are also used here. As such, knowledge of the *Biobase development and the new eSet* vignette would be beneficial; the vignette can directly be accessed with `vignette("BiobaseDevelopment", package="Biobase")`.

The initial goal is to use the *MSnbase* infrastructure for MS² labelled (iTRAQ [2] and TMT [3]) and label-free (spectral counting, index and abundance) quantitation - see the documentation for the `quantify` function for details.

2 MSnbase classes

All classes have a `.__classVersion__` slot, of class *Versioned* from the *Biobase* package. This slot documents the class version for any instance to be used for debugging and object update purposes. Any change in a class implementation should trigger a version change.

2.1 *pSet*: a virtual class for raw mass spectrometry data and meta data

This virtual class is the main container for mass spectrometry data, i.e spectra, and meta data. It is based on the *eSet* implementation for genomic data. The main difference with *eSet* is that the `assayData` slot is an environment containing any number of *Spectrum* instances (see section 2.7).

MSnbase level

One new slot is introduced, namely `processingData`, that contains one *MSnProcess* instance (see section 2.5). and the `experimentData` slot is now expected to contain *MIAPE* data (see section 2.6). The `annotation` slot has not been implemented, as no prior feature annotation is known in shotgun proteomics.

```
getClass("pSet")
```

```
Virtual Class "pSet" [package "MSnbase"]
```

```
Slots:
```

```
Name:          assayData          phenoData
Class:         environment NAnnotatedDataFrame
```

```
Name:          featureData          experimentData
Class:  AnnotatedDataFrame          MIAxE
```

```
Name:          protocolData          processingData
Class:  AnnotatedDataFrame          MSnProcess
```

```
Name:          .cache          .__classVersion__
Class:         environment          Versions
```

```
Extends: "Versioned"
```

```
Known Subclasses:
```

```
Class "MSnExp", directly
```

```
Class "OnDiskMSnExp", by class "MSnExp", distance 2, with explicit coerce
```

2.2 MSnExp: a class for MS experiments

MSnExp extends *pSet* to store MS experiments. It does not add any new slots to *pSet*. Accessors and setters are all inherited from *pSet* and new ones should be implemented for *pSet*. Methods that manipulate actual data in experiments are implemented for *MSnExp* objects.

```
getClass("MSnExp")
```

MSnbase devel

```
Class "MSnExp" [package "MSnbase"]

Slots:

Name:          assayData          phenoData
Class:         environment NAnnotatedDataFrame

Name:          featureData        experimentData
Class:  AnnotatedDataFrame        MIAxE

Name:          protocolData        processingData
Class:  AnnotatedDataFrame        MSnProcess

Name:          .cache             .__classVersion__
Class:         environment        Versions

Extends:
Class "pSet", directly
Class "Versioned", by class "pSet", distance 2

Known Subclasses:
Class "OnDiskMSnExp", directly, with explicit coerce
```

2.3 *OnDiskMSnExp*: a on-disk implementation of the *MSnExp* class

The *OnDiskMSnExp* class extends *MSnExp* and inherits all of its functionality but is aimed to use as little memory as possible based on a balance between memory demand and performance. Most of the spectrum-specific data, like retention time, polarity, total ion current are stored within the object's *featureData* slot. The actual M/Z and intensity values from the individual spectra are, in contrast to *MSnExp* objects, not kept in memory (in the *assayData* slot), but are fetched from the original files on-demand. Because mzML files are indexed, using the *mzR* package to read the relevant spectrum data is fast and only moderately slower than for in-memory *MSnExp*¹.

¹The *benchmarking* vignette compares data size and operation speed of the two implementations.

MSnbase devel

To keep track of data manipulation steps that are applied to spectrum data (such as performed by methods `removePeaks` or `clean`) a *lazy execution* framework was implemented. Methods that manipulate or subset a spectrum's M/Z or intensity values can not be applied directly to a `OnDiskMSnExp` object, since the relevant data is not kept in memory. Thus, any call to a processing method that changes or subset M/Z or intensity values are added as `ProcessingStep` items to the object's `spectraProcessingQueue`. When the spectrum data is then queried from an `OnDiskMSnExp`, the spectra are read in from the file and all these processing steps are applied on-the-fly to the spectrum data before being returned to the user.

The operations involving extracting or manipulating spectrum data are applied on a per-file basis, which enables parallel processing. Thus, all corresponding method implementations for `OnDiskMSnExp` objects have an argument `BPPARAM` and users can set a `PARALLEL_THRESH` option flag ² that enables to define how and when parallel processing should be performed (using the `BiocParallel` package).

²see `?MSnbaseOptions` for details.

Note that all data manipulations that are not applied to M/Z or intensity values of a spectrum (e.g. sub-setting by retention time etc) are very fast as they operate directly to the object's `featureData` slot.

```
getClass("OnDiskMSnExp")
Class "OnDiskMSnExp" [package "MSnbase"]

Slots:

Name:  spectraProcessingQueue      backend
Class:                list         character

Name:  assayData                  phenoData
Class:  environment              NAnnotatedDataFrame

Name:  featureData                experimentData
Class:  AnnotatedDataFrame        MIAxE

Name:  protocolData              processingData
Class:  AnnotatedDataFrame        MSnProcess

Name:  .cache                     .__classVersion__
```

MSnbase level

```
Class:          environment          Versions

Extends:
Class "MSnExp", directly
Class "pSet", by class "MSnExp", distance 2
Class "Versioned", by class "MSnExp", distance 3
```

The distinction between *MSnExp* and *OnDiskMSnExp* is often not explicitly stated as it should not matter, from a user's perspective, which data structure they are working with, as both behave in equivalent ways. Often, they are referred to as *in-memory* and *on-disk MSnExp* implementations.

2.4 MSnSet: a class for quantitative proteomics data

This class stores quantitation data and meta data after running `quantify` on an *MSnExp* object or by creating an *MSnSet* instance from an external file, as described in the `MSnbase-io` vignette and in `?readMSnSet`, `readMzTabData`, etc. The quantitative data is in form of a $m \times n$ matrix, where m is the number of features/spectra originally in the *MSnExp* used as parameter in `quantify` and n is the number of reporter ions (see section 2.8). If read from an external file, n corresponds to the number of features (protein groups, proteins, peptides, spectra) in the file and m is the number of columns with quantitative data (samples) in the file.

This prompted to keep a similar implementation as the *ExpressionSet* class, while adding the proteomics-specific annotation slot introduced in the *pSet* class, namely `processingData` for objects of class *MSnProcess* (see section 2.5).

```
getClass("MSnSet")

Class "MSnSet" [package "MSnbase"]

Slots:
```


MSnbase level

```
Name:      experimentData      processingData      qual
Class:      MIAPE              MSnProcess          data.frame

Name:      assayData          phenoData          featureData
Class:      AssayData AnnotatedDataFrame AnnotatedDataFrame

Name:      annotation          protocolData      ___classVersion__
Class:      character AnnotatedDataFrame      Versions

Extends:
Class "eSet", directly
Class "VersionedBiobase", by class "eSet", distance 2
Class "Versioned", by class "eSet", distance 3
```

The *MSnSet* class extends the virtual *eSet* class to provide compatibility for *ExpressionSet*-like behaviour. The experiment meta-data in `experimentData` is also of class *MIAPE* (see section 2.6). The `annotation` slot, inherited from *eSet* is not used. As a result, it is easy to convert *ExpressionSet* data from/to *MSnSet* objects with the coercion method `as`.

```
data(msnset)
class(msnset)

[1] "MSnSet"
attr(,"package")
[1] "MSnbase"

class(as(msnset, "ExpressionSet"))

[1] "ExpressionSet"
attr(,"package")
[1] "Biobase"

data(sample.ExpressionSet)
class(sample.ExpressionSet)

[1] "ExpressionSet"
attr(,"package")
[1] "Biobase"

class(as(sample.ExpressionSet, "MSnSet"))
```

MSnbase devel

```
[1] "MSnSet"  
attr(,"package")  
[1] "MSnbase"
```

2.5 *MSnProcess*: a class for logging processing meta data

This class aims at recording specific manipulations applied to *MSnExp* or *MSnSet* instances. The `processing` slot is a `character` vector that describes major processing. Most other slots are of class `logical` that indicate whether the data has been centroided, smoothed, ... although many of the functionality is not implemented yet. Any new processing that is implemented should be documented and logged here.

It also documents the raw data file from which the data originates (`files` slot) and the *MSnbase* version that was in use when the *MSnProcess* instance, and hence the *MSnExp/MSnSet* objects, were originally created.

```
getClass("MSnProcess")  
  
Class "MSnProcess" [package "MSnbase"]  
  
Slots:  
  
Name:          files          processing          merged  
Class:         character      character          logical  
  
Name:          cleaned        removedPeaks       smoothed  
Class:         logical        character          logical  
  
Name:          trimmed        normalised         MSnbaseVersion  
Class:         numeric        logical           character  
  
Name:          ___classVersion___  
Class:         Versions  
  
Extends: "Versioned"
```

2.6 *MIAPE*: Minimum Information About a Proteomics Experiment

The Minimum Information About a Proteomics Experiment [4, 5] *MIAPE* class describes the experiment, including contact details, information about the mass spectrometer and control and analysis software.

```
getClass("MIAPE")  
  
Class "MIAPE" [package "MSnbase"]  
  
Slots:  
  
Name:          title          url  
Class:         character       character  
  
Name:          abstract       pubMedIds  
Class:         character       character  
  
Name:          samples        preprocessing  
Class:         list           list  
  
Name:          other          dateStamp  
Class:         list           character  
  
Name:          name           lab  
Class:         character       character  
  
Name:          contact        email  
Class:         character       character  
  
Name:          instrumentModel instrumentManufacturer  
Class:         character       character  
  
Name: instrumentCustomisations softwareName  
Class:         character       character  
  
Name:          softwareVersion switchingCriteria  
Class:         character       character
```

MSnbase devel

```
Name:      isolationWidth      parameterFile
Class:      numeric            character

Name:      ionSource          ionSourceDetails
Class:      character          character

Name:      analyser           analyserDetails
Class:      character          character

Name:      collisionGas        collisionPressure
Class:      character          numeric

Name:      collisionEnergy      detectorType
Class:      character          character

Name:      detectorSensitivity  ___classVersion__
Class:      character          Versions
```

Extends:

Class "MIAxE", directly

Class "Versioned", by class "MIAxE", distance 2

2.7 *Spectrum et al.*: classes for MS spectra

Spectrum is a virtual class that defines common attributes to all types of spectra. MS1 and MS2 specific attributes are defined in the *Spectrum1* and *Spectrum2* classes, that directly extend *Spectrum*.

```
getClass("Spectrum")
```

```
Virtual Class "Spectrum" [package "MSnbase"]
```

Slots:

```
Name:      msLevel      peaksCount      rt
Class:      integer      integer          numeric

Name:      acquisitionNum  scanIndex      tic
```

MSnbase level

```
Class:          integer          integer          numeric

Name:           mz              intensity      fromFile
Class:          numeric          numeric        integer

Name:           centroided      smoothed      polarity
Class:          logical          logical        integer

Name:  ___classVersion__
Class:          Versions

Extends: "Versioned"

Known Subclasses: "Spectrum2", "Spectrum1"
```

```
getClass("Spectrum1")
```

```
Class "Spectrum1" [package "MSnbase"]
```

```
Slots:
```

```
Name:          msLevel          peaksCount          rt
Class:          integer          integer              numeric

Name:  acquisitionNum          scanIndex          tic
Class:          integer          integer              numeric

Name:           mz              intensity      fromFile
Class:          numeric          numeric        integer

Name:           centroided      smoothed      polarity
Class:          logical          logical        integer

Name:  ___classVersion__
Class:          Versions
```

```
Extends:
```

```
Class "Spectrum", directly
```

```
Class "Versioned", by class "Spectrum", distance 2
```

MSnbase devel

```
getClass("Spectrum2")
Class "Spectrum2" [package "MSnbase"]

Slots:

Name:          merged          precScanNum          precursorMz
Class:         numeric         integer              numeric

Name: precursorIntensity  precursorCharge  collisionEnergy
Class:         numeric         integer              numeric

Name:          msLevel          peaksCount          rt
Class:         integer         integer              numeric

Name:          acquisitionNum  scanIndex          tic
Class:         integer         integer              numeric

Name:          mz              intensity          fromFile
Class:         numeric         numeric             integer

Name:          centroided     smoothed          polarity
Class:         logical         logical             integer

Name:  ___classVersion___
Class:  Versions

Extends:
Class "Spectrum", directly
Class "Versioned", by class "Spectrum", distance 2
```

2.8 ReporterIons: a class for isobaric tags

The iTRAQ and TMT (or any other peak of interest) are implemented *ReporterIons* instances, that essentially defines an expected MZ position for the peak and a width around this value as well a names for the reporters.

MSnbase devel

```
getClass("ReporterIons")
Class "ReporterIons" [package "MSnbase"]

Slots:

Name:          name      reporterNames      description
Class:         character  character          character

Name:          mz        col                width
Class:         numeric   character          numeric

Name:  .__classVersion__
Class:  Versions

Extends: "Versioned"
```

2.9 *NAnnotatedDataFrame*: multiplexed *AnnotatedDataFrames*

The simple expansion of the *AnnotatedDataFrame* classes adds the `multiplex` and `multiLabel` slots to document the number and names of multiplexed samples.

```
getClass("NAnnotatedDataFrame")
Class "NAnnotatedDataFrame" [package "MSnbase"]

Slots:

Name:          multiplex      multiLabels      varMetadata
Class:         numeric       character        data.frame

Name:          data          dimLabels  .__classVersion__
Class:         data.frame    character   Versions

Extends:
Class "AnnotatedDataFrame", directly
```

MSnbase devel

```
Class "Versioned", by class "AnnotatedDataFrame", distance 2
```

2.10 Other classes

Lists of *MSnSet* instances

When several *MSnSet* instances are related to each other and should be stored together as different objects, they can be grouped as a list into an *MSnSetList* object. In addition to the actual *list* slot, this class also has basic logging functionality and enables iteration over the *MSnSet* instances using a dedicated `lapply` methods.

```
getClass("MSnSetList")  
Class "MSnSetList" [package "MSnbase"]  
  
Slots:  
  
Name:                x                log  ___classVersion__  
Class:                list            list          Versions  
  
Extends: "Versioned"
```

3 Miscellaneous

Unit tests *MSnbase* implements unit tests with the *testthat* package.

Processing methods Methods that process raw data, i.e. spectra should be implemented for *Spectrum* objects first and then `eapply`'ed (or similar) to the `assayData` slot of an *MSnExp* instance in the specific method.

4 Session information

- R version 3.3.2 (2016-10-31), x86_64-pc-linux-gnu
- Locale: LC_CTYPE=en_US.UTF-8, LC_NUMERIC=C, LC_TIME=en_US.UTF-8, LC_COLLATE=C, LC_MONETARY=en_US.UTF-8, LC_MESSAGES=en_US.UTF-8, LC_PAPER=en_US.UTF-8, LC_NAME=C, LC_ADDRESS=C, LC_TELEPHONE=C, LC_MEASUREMENT=en_US.UTF-8, LC_IDENTIFICATION=C
- Base packages: base, datasets, grDevices, graphics, grid, methods, parallel, stats, stats4, utils
- Other packages: AnnotationDbi 1.36.0, Biobase 2.34.0, BiocGenerics 0.20.0, BiocParallel 1.8.1, BiocStyle 2.2.1, IRanges 2.8.1, MLInterfaces 1.54.0, MSnbase 2.0.2, ProtGenerics 1.6.0, Rcpp 0.12.8, RcppClassic 0.9.6, Rdisop 1.34.0, S4Vectors 0.12.1, XML 3.98-1.5, annotate 1.52.1, cluster 2.0.5, ggplot2 2.2.1, gplots 3.0.1, knitr 1.15.1, microbenchmark 1.4-2.1, msdata 0.14.0, mzR 2.8.0, pRoloc 1.14.5, pRolocdata 1.12.0, pryr 0.1.2, reshape2 1.4.2, zoo 1.7-14
- Loaded via a namespace (and not attached):
BiocInstaller 1.24.0, DBI 0.5-1, DEoptimR 1.0-8, FNN 1.1, KernSmooth 2.23-15, MALDIquant 1.16, MASS 7.3-45, Matrix 1.2-7.1, MatrixModels 0.4-1, ModelMetrics 1.1.0, R6 2.2.0, RColorBrewer 1.1-2, RCurl 1.95-4.8, RSQLite 1.1-1, SparseM 1.74, TH.data 1.0-7, affy 1.52.0, affyio 1.44.0, assertthat 0.1, backports 1.0.4, base64enc 0.1-3, biomaRt 2.30.0, bitops 1.0-6, caTools 1.17.1, car 2.1-4, caret 6.0-73, class 7.3-14, codetools 0.2-15, colorspace 1.3-2, dendextend 1.3.0, digest 0.6.11, diptest 0.75-7, doParallel 1.0.10, dplyr 0.5.0, e1071 1.6-7, evaluate 0.10, flexmix 2.3-13, foreach 1.4.3, fpc 2.1-10, gbm 2.1.1, gdata 2.17.0, genefilter 1.56.0, ggvis 0.4.3, gridExtra 2.2.1, gtable 0.2.0, gtools 3.5.0, highr 0.6, htmltools 0.3.5, htmlwidgets 0.8, httpuv 1.3.3, hwriter 1.3.2, impute 1.48.0, iterators 1.0.8, jsonlite 1.2, kernlab 0.9-25, labeling 0.3, lattice 0.20-34, lazyeval 0.2.0, limma 3.30.7, lme4 1.1-12, lpSolve 5.6.13, magrittr 1.5, mclust 5.2.1, memoise 1.0.0,

MSnbase devel

mgcv 1.8-16, mime 0.5, minqa 1.2.4, mlbench 2.1-1,
modeltools 0.2-21, multcomp 1.4-6, munsell 0.4.3,
mvtnorm 1.0-5, mzID 1.12.0, nlme 3.1-128, nloptr 1.0.4,
nnet 7.3-12, pbkrtest 0.4-6, pcaMethods 1.66.0, pls 2.6-0,
plyr 1.8.4, prabclus 2.2-6, preprocessCore 1.36.0, proxy 0.4-16,
quantreg 5.29, randomForest 4.6-12, rda 1.0.2-2,
rmarkdown 1.3, robustbase 0.92-7, rpart 4.1-10, rprojroot 1.1,
sampling 2.8, sandwich 2.3-4, scales 0.4.1, sfsmisc 1.1-0,
shiny 0.14.2, splines 3.3.2, stringi 1.1.2, stringr 1.1.0,
survival 2.40-1, threejs 0.2.2, tibble 1.2, tools 3.3.2,
trimcluster 0.1-2, vsn 3.42.3, whisker 0.3-2, xtable 1.8-2,
yaml 2.1.14, zlibbioc 1.20.0

References

- [1] Robert C. Gentleman, Vincent J. Carey, Douglas M. Bates, Ben Bolstad, Marcel Dettling, Sandrine Dudoit, Byron Ellis, Laurent Gautier, Yongchao Ge, Jeff Gentry, Kurt Hornik, Torsten Hothorn, Wolfgang Huber, Stefano Iacus, Rafael Irizarry, Friedrich Leisch, Cheng Li, Martin Maechler, Anthony J. Rossini, Gunther Sawitzki, Colin Smith, Gordon Smyth, Luke Tierney, Jean Y. H. Yang, and Jianhua Zhang. Bioconductor: open software development for computational biology and bioinformatics. *Genome Biol*, 5(10):–80, 2004. URL: <http://dx.doi.org/10.1186/gb-2004-5-10-r80>, [doi:10.1186/gb-2004-5-10-r80](https://doi.org/10.1186/gb-2004-5-10-r80).
- [2] Philip L. Ross, Yulin N. Huang, Jason N. Marchese, Brian Williamson, Kenneth Parker, Stephen Hattan, Nikita Khainovski, Sasi Pillai, Subhakar Dey, Scott Daniels, Subhasish Purkayastha, Peter Juhasz, Stephen Martin, Michael Bartlet-Jones, Feng He, Allan Jacobson, and Darryl J. Pappin. Multiplexed protein quantitation in *saccharomyces cerevisiae* using amine-reactive isobaric tagging reagents. *Mol Cell Proteomics*, 3(12):1154–1169, Dec 2004. URL: <http://dx.doi.org/10.1074/mcp.M400129-MCP200>, [doi:10.1074/mcp.M400129-MCP200](https://doi.org/10.1074/mcp.M400129-MCP200).

- [3] Andrew Thompson, Jürgen Schäfer, Karsten Kuhn, Stefan Kienle, Josef Schwarz, Günter Schmidt, Thomas Neumann, R Johnstone, A Karim A Mohammed, and Christian Hamon. Tandem mass tags: a novel quantification strategy for comparative analysis of complex protein mixtures by MS/MS. *Anal. Chem.*, 75(8):1895–904, 2003.
- [4] Chris F. Taylor, Norman W. Paton, Kathryn S. Lilley, Pierre-Alain Binz, Randall K. Julian, Andrew R. Jones, Weimin Zhu, Rolf Apweiler, Ruedi Aebersold, Eric W. Deutsch, Michael J. Dunn, Albert J. R. Heck, Alexander Leitner, Marcus Macht, Matthias Mann, Lennart Martens, Thomas A. Neubert, Scott D. Patterson, Peipei Ping, Sean L. Seymour, Puneet Souda, Akira Tsugita, Joel Vandekerckhove, Thomas M. Vondriska, Julian P. Whitelegge, Marc R. Wilkins, Ioannis Xenarios, John R. Yates, and Henning Hermjakob. The minimum information about a proteomics experiment (miape). *Nat Biotechnol*, 25(8):887–893, Aug 2007. URL: <http://dx.doi.org/10.1038/nbt1329>, doi:10.1038/nbt1329.
- [5] Chris F Taylor, Pierre-Alain Binz, Ruedi Aebersold, Michel Affolter, Robert Barkovich, Eric W Deutsch, David M Horn, Andreas HÄijhmer, Martin Kussmann, Kathryn Lilley, Marcus Macht, Matthias Mann, Dieter MÄijller, Thomas A Neubert, Janice Nickson, Scott D Patterson, Roberto Raso, Kathryn Resing, Sean L Seymour, Akira Tsugita, Ioannis Xenarios, Rong Zeng, and Randall K Julian. Guidelines for reporting the use of mass spectrometry in proteomics. *Nat. Biotechnol.*, 26(8):860–1, 2008. doi:10.1038/nbt0808-860.