Package 'Uniquorn'

April 15, 2017

Title Identification of cancer cell lines based on their weighted mutational/ variational fingerprint

Version 1.2.0

Description This packages enables users to identify cancer cell lines. Cancer cell line misidentification and cross-contamination reprents a significant challenge for cancer researchers. The identification is vital and in the frame of this package based on the locations/ loci of somatic and germline mutations/ variations.
The input format is vcf/ vcf.gz and the files have to contain a single cancer cell line sample (i.e. a single member/genotype/gt column in the vcf file).
The implemented method is optimized for the Next-generation whole exome and whole genome DNA-sequencing technology. RNA-seq data is very likely to work as well but hasn't been rigiously tested yet. Panel-seq will require manual adjustment of thresholds

Imports DBI, stringr, RSQLite, R.utils, WriteXLS, stats

Depends R (>= 3.3)

License Artistic-2.0

LazyData TRUE

Type Package

Maintainer 'Raik Otto' <raik.otto@hu-berlin.de>

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Author Raik Otto

RoxygenNote 5.0.1

NeedsCompilation no

Suggests testthat, knitr, rmarkdown, BiocGenerics, RUnit

biocViews Software, StatisticalMethod, WholeGenome, ExomeSeq

VignetteBuilder knitr

R topics documented:

`add_custom_vcf_to_database` *Adds a custom vcf file to the three existing cancer cell line panels*

Description

Adds a custom vcf file to the three existing cancer cell line panels

Usage

```
add_custom_vcf_to_database(
 vcf_file_path,
ref_gen = "GRCH37",
name_cl = "",
safe_mode = FALSE,
test_mode = FALSE)
```

Arguments

`vcf_file_path`	Input vcf file. Only one sample column allowed.
`ref_gen`	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
`name_cl`	Name of the to-be-added cancer cell line sample. '_CUSTOM' will automatically be added as suffix.
`safe_mode`	Only add mutations to the database where there already are mutations found in the cannonical cancer cell lines. This is a safety mechanism against overfitting if there are too few custom training samples.
`test_mode`	Is this a test? Just for internal use

Value

Message if the adding has succeeded

Examples

```
HT29_vcf_file = system.file("extdata/HT29.vcf.gz", package="Uniquorn");
add_custom_vcf_to_database(
vcf_file_path = HT29_vcf_file,
name_cl = "",
ref_gen = "GRCH37",
safe_mode = FALSE,
test_mode = TRUE )
```

`create_bed_file` *create_bed_file*

Description

Creates BED files from the found and not found annotated mutations

Usage

```
create_bed_file(

sim_list,

vcf_fingerprint,

res_table,

output_file,

ref_gen,

manual_identifier

)
```

Arguments

`sim_list`	R table which contains the mutations from the training database for the cancer cell lines
`vcf_fingerprint`	contains the mutations that are present in the query cancer cell line's vcf file
`res_table`	Table containing the identification results
`output_file`	Path to output file
`ref_gen`	Reference genome version
`manual_identifier`	Manually enter a vector of CL name(s) whose bed files should be created, independently from them passing the detection threshold

Value

Returns a message which indicates if the BED file creation has succeeded

identify_vcf_file *identify_VCF_file*

Description

Identifies a cancer cell lines contained in a vcf file based on the pattern (start & length) of all contained mutations/ variations.

Usage

```
identify_vcf_file(
vcf_file,
output_file = "",
ref_gen = "GRCH37",
minimum_matching_mutations = 0,
mutational_weight_inclusion_threshold = 1.0,
only_first_candidate = FALSE,
write_xls = FALSE,
output_bed_file = FALSE,
manual_identifier_bed_file = "",
verbose = FALSE,
p_value = .05,
q_value = .05,
confidence_score = 10.0)
```

Arguments

vcf_file	Input vcf file. Only one sample column allowed.
output_file	Path of the output file. If blank, autogenerated as name of input file plus '_uniquorn_ident.tab' suffix.
ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
minimum_matching_mutations	The minimum amount of mutations that has to match between query and training sample for a positive prediction
mutational_weight_inclusion_threshold	Include only mutations with a weight of at least x. Range: 0.0 to 1.0. 1= unique to CL. ~0 = found in many CL samples.
only_first_candidate	Only the CL identifier with highest score is predicted to be present in the sample
write_xls	Create identification results additionally as xls file for easier reading
output_bed_file	If BED files for IGV visualization should be created for the Cancer Cell lines that pass the threshold
manual_identifier_bed_file	Manually enter a vector of CL name(s) whose bed files should be created, independently from them passing the detection threshold
verbose	Print additional information

p_value	Required p-value for identification
q_value	Required q-value for identification
confidence_score	Threshold above which a positive prediction occurs default 25.0

Details

identify_vcf_file parses the vcf file and predicts the identity of the sample

Value

R table with a statistic of the identification result

Examples

```
HT29_vcf_file = system.file("extdata/HT29.vcf.gz", package="Uniquorn");

identification = identify_vcf_file( HT29_vcf_file )
```

---

initiate_canonical_databases

initiate_canonical_databases

Description

Parses data into r list variable

Usage

```
initiate_canonical_databases(
cosmic_file = "CosmicCLP_MutantExport.tsv",
ccle_file = "CCLE_hybrid_capture1650_hg19_NoCommonSNPs_CDS_2012.05.07.maf",
ref_gen = "GRCH37")
```

Arguments

cosmic_file	The path to the cosmic DNA genotype data file. Ensure that the right reference genome is used
ccle_file	The path to the ccle DNA genotype data file. Ensure that the right reference genome is used
ref_gen	Reference genome version

Value

Returns message if parsing process has succeeded

Examples

```
initiate_canonical_databases(
cosmic_file = "CosmicCLP_MutantExport.tsv",
ccle_file = "CCLE_hybrid_capture1650_hg19_NoCommonSNPs_CDS_2012.05.07.maf",
ref_gen = "GRCH37")
```

initiate_db_and_load_data
initiate_db_and_load_data

Description

Intern utility function, loads database and return the sim_list and sim_list_stats variables.

Usage

```
initiate_db_and_load_data(
ref_gen,
request_table,
load_default_db )
```

Arguments

ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
request_table	Names of the tables to be extracted from the database
load_default_db	Indicate whether the default db should be used as source for the data

Value

Returns the sim_list and sim_list_stats variable

parse_ccle_genotype_data
parse_ccle_genotype_data

Description

Parses ccle genotype data

Usage

```
parse_ccle_genotype_data(ccle_file, sim_list)
```

Arguments

ccle_file	Path to CCLE file on hard disk
sim_list	Variable containing mutations and cell line

Value

The R Table sim_list which contains the CCLE fingerprints

parse_cosmic_genotype_data *parse_cosmic_genotype_data*

Description

Parses cosmic genotype data

Usage

```
parse_cosmic_genotype_data(cosmic_file, sim_list)
```

Arguments

cosmic_file	Path to cosmic clp file in hard disk
sim_list	Variable containing mutations & cell line

Value

The R Table sim_list which contains the CoSMIC CLP fingerprints

parse_vcf_file *parse_vcf_file*

Description

Parses the vcf file and filters all information except for the start and length of variations/ mutations.

Usage

```
parse_vcf_file( vcf_file_path )
```

Arguments

vcf_file_path	Path to the vcf file on the operating system

Value

Loci-based DNA-mutational fingerprint of the cancer cell line as found in the input VCF file

`remove_custom_vcf_from_database` *Removes a cancer cell line training fingerprint (vcf file) from the database. The names of all training sets can be seen by using the function* `show_contained_cls`.

Description

Removes a cancer cell line training fingerprint (vcf file) from the database. The names of all training sets can be seen by using the function `show_contained_cls`.

Usage

```
remove_custom_vcf_from_database(
name_cl,
ref_gen = "GRCH37",
test_mode = FALSE)
```

Arguments

`name_cl`	name of the cancer cell line training fingerprintt
`ref_gen`	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
`test_mode`	Is this a test? Just for internal use

Value

Message that indicates if the removal was succesful

Examples

```
remove_custom_vcf_from_database(
name_cl = "HT29_CELLMINER",
ref_gen = "GRCH37",
test_mode = TRUE )
```

`re_calculate_cl_weights` *Re-calculate sim_list_weights*

Description

This function re-calculates the weights of mutation after a change of the training set

Usage

```
re_calculate_cl_weights(sim_list, ref_gen)
```

Arguments

sim_list	R Table which contains a mapping from mutations/ variations to their containing CLs
ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37

Value

A list containing both the sim_list at pos 1 and sim_list_stats at pos 2 data frames.

---

show_contained_cls *show_contained_cls*

Description

Show all cancer cell line identifier present in the database for a selected reference genome: This function shows the names, amount of mutations/ variations, overall weight of the mutations of all contained training CLs for a chosen reference genome.

Usage

```
show_contained_cls(
ref_gen)
```

Arguments

ref_gen	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37

Value

R table which contains the identifier of all cancer cell line samples with the specific reference genome and the weight of all mutations

Examples

```
contained_cls = show_contained_cls(
ref_gen = "GRCH37")
```

show_contained_mutations *show_contained_mutations*

Description

Show all mutations present in the database for a selected reference Genome: This function shows all training-set mutations for a selected reference genome, i.e. the mutations that are being used for identification of query cancer cell lines.

Usage

```
show_contained_mutations(
ref_gen )
```

Arguments

ref_gen	Reference genome version

Value

R Table which contains all mutations associated with a particular cancer cell line for a specified reference genome

Examples

```
contained_cls = show_contained_mutations( ref_gen = "GRCH37" )
```

show_contained_mutations_for_cl *show_contained_mutations_for_cl*

Description

Show all mutations present in the database for a selected cancer cell line and reference Genome

Usage

```
show_contained_mutations_for_cl(
name_cl,
ref_gen)
```

Arguments

name_cl	Name of the cancer cell line sample stored in the database
ref_gen	Reference genome version

Value

R table which contains all mutations associated with the defined cancer cell line and reference genome

Examples

```
SK_OV_3_CELLMINER_mutations = show_contained_mutations_for_cl(
name_cl = "SK_OV_3_CELLMINER_mutations",
ref_gen = "GRCH37")
```

`show_which_cls_contain_mutation` *show_which_cls_contain_mutation*

Description

Show all cancer cell lines in the database which contained the specified mutation and reference Genome. Closed interval coordinates. Format mutation: CHR_START_STOP, e.g. 1_123_123

Usage

```
show_which_cls_contain_mutation(
mutation_name,
ref_gen)
```

Arguments

`mutation_name`	Name of the mutation in the format CHROMOSOME_START_STOP, e.g. '11_244501_244510'
`ref_gen`	Reference genome version

Value

R table which contains all cancer cell line samples which contain the specified mutation with respect to the specified reference genome version

Examples

```
Cls_containing_mutations = show_which_cls_contain_mutation(
mutation_name = "10_103354427_103354427",
ref_gen = "GRCH37")
```

`split_add` *split_add*

Description

split_add

Usage

```
split_add(vcf_matrix_row)
```

Arguments

`vcf_matrix_row`	row of the vcf file

Value

Transformed entry of vcf file, reduced to start and length

`write_data_to_db` *write_data_to_db*

Description

Intern utility function, writes to database the sim_list and sim_list_stats variables

Usage

```
write_data_to_db(
content_table,
table_name,
ref_gen,
overwrite,
test_mode )
```

Arguments

`content_table`	Tables to be written in db
`table_name`	Name of the table to be written into the DB
`ref_gen`	Reference genome version. All training sets are associated with a reference genome version. Default: GRCH37
`overwrite`	Overwrite the potentially existing table
`test_mode`	Is this a test? Just for internal use

Value

the sim_list and sim_list_stats variable

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