

# Package ‘SubCellBarCode’

December 4, 2024

**Type** Package

**Title** SubCellBarCode: Integrated workflow for robust mapping and visualizing whole human spatial proteome

**Version** 1.23.0

**Author** Taner Arslan

**Maintainer** Taner Arslan <taner.arslan@ki.se>

**Description** Mass-Spectrometry based spatial proteomics have enabled the proteome-wide mapping of protein subcellular localization (Orre et al. 2019, Molecular Cell). SubCellBarCode R package robustly classifies proteins into corresponding subcellular localization.

**License** GPL-2

**Encoding** UTF-8

**LazyData** true

**Depends** R (>= 3.6)

**Suggests** knitr, rmarkdown, BiocStyle

**Imports** Rtsne, scatterplot3d, caret, e1071, ggplot2, gridExtra, networkD3, ggrepel, graphics, stats, org.Hs.eg.db, AnnotationDbi

**biocViews** Proteomics, MassSpectrometry, Classification

**RoxygenNote** 7.1.1

**VignetteBuilder** knitr

**git\_url** <https://git.bioconductor.org/packages/SubCellBarCode>

**git\_branch** devel

**git\_last\_commit** 260c964

**git\_last\_commit\_date** 2024-10-29

**Repository** Bioconductor 3.21

**Date/Publication** 2024-12-03

## Contents

applyThresholdCompartment . . . . .	2
applyThresholdNeighborhood . . . . .	3
calculateCoveredProtein . . . . .	4
calRowMean . . . . .	5
candidateRelocatedProteins . . . . .	6
compareCls . . . . .	7
computeThresholdCompartment . . . . .	8
computeThresholdNeighborhood . . . . .	9
convert2symbol . . . . .	10
hcc827Ctrl . . . . .	11
hcc827CtrlPSMCount . . . . .	11
hcc827exon . . . . .	12
hcc827GEF . . . . .	13
hcc827GEFClass . . . . .	13
hcc827GefPSMCount . . . . .	14
loadData . . . . .	14
markerProteins . . . . .	15
markerQualityControl . . . . .	15
mergeCls . . . . .	16
mergeProbability . . . . .	17
plotBarcode . . . . .	18
plotMultipleProtein . . . . .	20
replacePrediction . . . . .	20
sankeyPlot . . . . .	21
sumProbability . . . . .	22
svmClassification . . . . .	23
svmExternalData . . . . .	24
tsneVisualization . . . . .	25
<b>Index</b>	<b>26</b>

---

applyThresholdCompartment

*Apply thresholds to compartments*

---

### Description

Apply thresholds for all predictions to increase the true positive rate and remove poor classification.

### Usage

```
applyThresholdCompartment(all.repA, all.repB, threshold.df)
```

**Arguments**

all.repA            data.frame; all predictions and probability vectors for each protein in replicate A  
all.repB            data.frame; all predictions and probability vectors for each protein in replicate B  
threshold.df        data.frame; collection of precision and recall values for each compartment

**Value**

c.cls.df

**Examples**

```
{  
  df <- loadData(SubCellBarCode::hcc827Ctrl)  
  c.prots <- calculateCoveredProtein(rownames(df), markerProteins[,1])  
  set.seed(7)  
  c.prots <- sample(c.prots, 550)  
  cls <- svmClassification(c.prots, df, markerProteins)  
  test.A <- cls[[1]]$svm.test.prob.out  
  test.B <- cls[[2]]$svm.test.prob.out  
  t.c.df <- computeThresholdCompartment(test.A, test.B)  
  all.A <- cls[[1]]$all.prot.pred  
  all.B <- cls[[2]]$all.prot.pred  
  c.cls.df <- applyThresholdCompartment(all.A, all.B, t.c.df)  
}
```

---

applyThresholdNeighborhood

*Apply thresholds to neighborhood classification*

---

**Description**

Apply thresholds for all predictions at the neighborhood level to increase the true positive rate and remove poor classification.

**Usage**

```
applyThresholdNeighborhood(all.repA, all.repB, threshold.df)
```

**Arguments**

all.repA            data.frame; all predictions and probability vectors for each protein in replicate A  
 all.repB            data.frame; all predictions and probability vectors for each protein in replicate B  
 threshold.df       data.frame; collection of precision and recall values for each neighborhood

**Value**

n.cls.df

**Examples**

```
{
df <- loadData(SubCellBarCode::hcc827Ctrl)

c.prots <- calculateCoveredProtein(rownames(df), markerProteins[,1])

set.seed(7)
c.prots <- sample(c.prots, 600)
cls <- svmClassification(c.prots, df, markerProteins)

test.A <- cls[[1]]$svm.test.prob.out
test.B <- cls[[2]]$svm.test.prob.out

t.n.df <- computeThresholdNeighborhood(test.A, test.B)

all.A <- cls[[1]]$all.prot.pred
all.B <- cls[[2]]$all.prot.pred

n.cls.df <- applyThresholdNeighborhood(all.A, all.B, t.n.df)
}
```

---

calculateCoveredProtein

*Evaluate marker protein coverage*

---

**Description**

Given the proteomics data, number of overlapped marker proteins is calculated. Bar plot for each compartment is plotted.

**Usage**

```
calculateCoveredProtein(proteinIDs, markerproteins)
```

**Arguments**

proteinIDs        character; gene symbol id  
 markerproteins   character; 3365 proteins gene symbol ids

**Value**

covered.proteins

**Examples**

```
{  
df <- loadData(SubCellBarCode::hcc827Ctrl)  
  
c.prots <- calculateCoveredProtein(rownames(df), markerProteins[,1])  
}
```

---

calRowMean

*Compute the means of replicates*

---

**Description**

Duplicated fractions A and B are summarized by taking their mean for each protein. After taking the mean, the data log2 transformed. Further, the 5 main fractions are used to check correlation between input datas. It is a helper function.

**Usage**

```
calRowMean(d.df)
```

**Arguments**

d.df                    data.frame; A data frame of 10 fraction profiles consisting of replicate A and B.

**Value**

r.df

**Examples**

```
{  
  
r.df <- calRowMean(SubCellBarCode::hcc827Ctrl)  
  
}
```

---

candidateRelocatedProteins

*Identify candidate relocated proteins*

---

### Description

Identify candidate condition-dependent relocated proteins by comparing neighborhood classifications with respect to protein-protein pearson correlation and mininum PSM, peptide spectrum matching, count.

### Usage

```
candidateRelocatedProteins(
  sampleCls1,
  s1PSM,
  s1Quant,
  sampleCls2,
  s2PSM,
  s2Quant,
  annotation = FALSE,
  min.psm = 2,
  pearson.cor = 0.8
)
```

### Arguments

sampleCls1	data.frame; merged classification, combination of compartment and neighborhood classification.
s1PSM	data.frame; minimum PSM count table across ten TMT channel
s1Quant	data.frame; fractionation quantification data
sampleCls2	data.frame; merged classification, combination of compartment and neighborhood classification.
s2PSM	data.frame; minimum PSM count table across ten TMT channel
s2Quant	data.frame; fractionation quantification data
annotation	boolean; labeling the selected proteins
min.psm	numeric; minimum psm, peptide spectra matching value
pearson.cor	numeric; pearson correlation threshold

### Value

candidate.df

**Examples**

```
{
  candidate.df <- candidateRelocatedProteins(hcc827GEFClass, hcc827GefPSMCount,
  hcc827GEF, hcc827GEFClass, hcc827GefPSMCount, hcc827GEF,
  annotation = FALSE)
}
```

compareCls

*Compare exon and gene centric classifications***Description**

Comparison of the gene centric and exon centric classification. Additionally, correlation analysis is performed using quantification data.

**Usage**

```
compareCls(geneCls, exonCls)
```

**Arguments**

geneCls	data frame gene centric classification output
exonCls	data frame exon centric classification output

**Value**

```
c.df
```

**Examples**

```
{
  exon.cls <- data.frame(Protein = c("ENSE00000331854",
  "ENSE00000331855",
  "ENSE00000331859"),
  NeighborhoodCls = c("Cytosol",
  "Cytosol",
  "Cytosol"),
  CompartmentCls = c("C1", "C1", "C1"),
  Secretory = c(0.1, 0.1, 0.1),
  Nuclear = c(0.2, 0.2, 0.2),
  Cytosol = c(0.2, 0.2, 0.2),
  Mitochondria = c(0.2, 0.2, 0.2),
  S1 = c(0.2, 0.2, 0.2),
  S2 = c(0.2, 0.2, 0.2),
  S3 = c(0.2, 0.2, 0.2),
```

```

S4 = c(0.2, 0.2, 0.2),
N1 = c(0.2, 0.2, 0.2),
N2 = c(0.2, 0.2, 0.2),
N3 = c(0.2, 0.2, 0.2),
N4 = c(0.2, 0.2, 0.2),
C1 = c(0.2, 0.2, 0.2),
C2 = c(0.2, 0.2, 0.2),
C3 = c(0.2, 0.2, 0.2),
C4 = c(0.2, 0.2, 0.2),
C5 = c(0.2, 0.2, 0.2),
M1 = c(0.2, 0.2, 0.2),
M2 = c(0.2, 0.2, 0.2),
GeneSymbol = c("COPB1", "COPB1", "COPB1"),
PeptideCount = c(2, 4, 7))

gene.cls <- data.frame(Protein = c("COPB1"),
  NeighborhoodCls = c("Cytosol"),
  CompartmentCls = c("C1"),
  Secretory = c(0.1),
  Nuclear = c(0.2),
  Cytosol = c(0.2),
  Mitochondria = c(0.2),
  S1 = c(0.2),
  S2 = c(0.2),
  S3 = c(0.2),
  S4 = c(0.2),
  N1 = c(0.2),
  N2 = c(0.2),
  N3 = c(0.2),
  N4 = c(0.2),
  C1 = c(0.2),
  C2 = c(0.2),
  C3 = c(0.2),
  C4 = c(0.2),
  C5 = c(0.2),
  M1 = c(0.2),
  M2 = c(0.2))

comp.df <- compareCls(gene.cls, exon.cls)

}

```

---

```
computeThresholdCompartment
```

*Probability threshold for compartment classification*

---

### **Description**

Thresholds for each compartment are decided to get confident predictions.



**Usage**

```
computeThresholdCompartment(test.repA, test.repB)
```

**Arguments**

test.repA	data.frame; test predictions, observation and probability vectors for each protein in replicate A
test.repB	data.frame; test predictions, observation and probability vectors for each protein in replicate B

**Value**

```
threshold.compartment.df
```

**Examples**

```
{  
  df <- loadData(SubCellBarCode::hcc827Ctrl)  
  
  c.prots <- calculateCoveredProtein(rownames(df), markerProteins[,1])  
  
  set.seed(7)  
  c.prots <- sample(c.prots, 550)  
  cls <- svmClassification(c.prots, df, markerProteins)  
  
  test.A <- cls[[1]]$svm.test.prob.out  
  test.B <- cls[[2]]$svm.test.prob.out  
  
  t.c.df <- computeThresholdCompartment(test.A, test.B)  
}
```

---

```
computeThresholdNeighborhood
```

*Probability threshold for neighborhood classification*

---

**Description**

Thresholds for each neighborhood are decided to get confident predictions.

**Usage**

```
computeThresholdNeighborhood(test.repA, test.repB)
```

**Arguments**

test.repA	data.frame; test predictions, observation and probability vectors for each protein in replicate A
test.repB	data.frame; test predictions, observation and probability vectors for each protein in replicate B

**Value**

threshold.neighborhood.df

**Examples**

```
{  
df <- loadData(SubCellBarCode::hcc827Ctrl)  
  
c.prots <- calculateCoveredProtein(rownames(df), markerProteins[,1])  
  
set.seed(7)  
c.prots <- sample(c.prots, 600)  
cls <- svmClassification(c.prots, df, markerProteins)  
  
test.A <- cls[[1]]$svm.test.prob.out  
test.B <- cls[[2]]$svm.test.prob.out  
  
t.n.df <- computeThresholdNeighborhood(test.A, test.B)  
}
```

---

convert2symbol

*Convert identifier to gene symbol*

---

**Description**

Identifier for each feature should be converted into gene symbols unless they are not gene symbols

**Usage**

```
convert2symbol(df, id = "UNIPROT")
```

**Arguments**

df	data.frame; fractionated proteomics data where data contains 10 columns of duplicated 5 fractionations and rownames must be identifier e.g. UNIPROT, Entrez ID
id	caharacter; identifier id for each protein

**Value**

df

**Examples**

```
{  
  
df <- data.frame(Uniprot = c("A4D0S4", "A8TX70", "000305", "000337"),  
Organism = rep("Homo Sap.", 4))  
  
rownames(df) <- df$Uniprot  
}
```

---

hcc827Ctrl	<i>HCC827 Control Cell Line</i>
------------	---------------------------------

---

**Description**

Subcellular fractionated cell line.

**Usage**

```
hcc827Ctrl
```

**Format**

A data frame where 10480 protein gene-centric ids and 5 replicated subcellular fractions.

**References**

Orre et al. 2019 Cell 73, 1-17

**Examples**

```
{  
head(hcc827Ctrl)  
}
```

---

hcc827CtrlPSMCount	<i>Minimum PSM Count in HCC827Ctrl Cell Line.</i>
--------------------	---

---

**Description**

Minimum PSM, Peptide Sequence Match, Count table for HCC827Ctrl Cell Line.

**Usage**

```
hcc827CtrlPSMCount
```

**Format**

A data frame where 10480 protein gene-centric ids minimum PSM count.

**References**

Orre et al. 2019 Cell 73, 1-17

**Examples**

```
{  
head(hcc827Ctr1PSMCount)  
}
```

---

hcc827exon

*HCC827 Control Exon Cell Line*

---

**Description**

Exon-centric sub data of hcc827 fractionated data.

**Usage**

```
hcc827exon
```

**Format**

A data frame where 500 exon-centric ensemble identifiers, corresponding gene symbols, 5 replicated subcellular fractions and number of unique peptides matched to associated exon.

**References**

Orre et al. 2019 Cell 73, 1-17

**Examples**

```
{  
head(hcc827exon)  
}
```

---

`hcc827GEF`*Gefitinib treated HCC827 Cell Line*

---

**Description**

HCC827 cell line was treated with Gefitinib which is EGFR inhibition.

**Usage**`hcc827GEF`**Format**

A data frame where 10398 protein gene-centric ids and 5 replicated subcellular fractions with duplicates.

**References**

Orre et al. 2019 Cell 73, 1-17

**Examples**

```
{  
  head(hcc827GEF)  
}
```

---

`hcc827GEFClass`*Gefitinib treated HCC827 Cell Line Classification*

---

**Description**

Gefitinib treated HCC827 cell line classification contains both neighborhood and compartment level. The data will be used for the relocalization analysis.

**Usage**`hcc827GEFClass`**Format**

A data frame where 10398 protein gene-centric ids and corresponding compartment and neighborhood classification along with classification probabilities.

**References**

Orre et al. 2019 Cell 73, 1-17

**Examples**

```
{
  head(hcc827GEFC1ass)
}
```

---

hcc827GefPSMCount	<i>Minimum PSM Count in HCC827 Gefitinib Cell Line.</i>
-------------------	---

---

**Description**

Minimum PSM, Peptide Sequence Match, Count table for HCC827 Gefitinib Cell Line.

**Usage**

```
hcc827GefPSMCount
```

**Format**

A data frame where 10398 protein gene-centric ids minimum PSM count.

**References**

Orre et al. 2019 Cell 73, 1-17

**Examples**

```
{
  head(hcc827GefPSMCount)
}
```

---

loadData	<i>Load the fractionated proteomics data</i>
----------	--

---

**Description**

Sampled median normalized TMT ratios are checked if there is any "NA" value. If any, the corresponding row is filtered out. Later, the data is normalized by taking log2.

**Usage**

```
loadData(protein.data)
```

**Arguments**

protein.data	data.frame; fractionated proteomics data where data contains 10 columns of duplicated 5 fractionations and rownames must be gene-centric protein names
--------------	--

**Value**

protein.data.df

**Examples**

```
{  
df <- loadData(SubCellBarCode::hcc827Ctrl[1:20,])  
}
```

---

markerProteins	<i>Marker Proteins Source</i>
----------------	-------------------------------

---

**Description**

Data for the proteins whose localizations were well characterized. It also contains color codes for each compartment and median fractionation profiles for 5 fractions which are Cyto., Nsol., Nucl., Horg., Lorg., with replicates A and B. These fractionation profiles will be used for the marker protein quality control.

**Usage**

markerProteins

**Format**

A data frame of 3365 proteins as rows and 13 columns headers.

**References**

Orre et al. 2019 Cell 73, 1-17

---

markerQualityControl	<i>Evaluate the quality of the marker proteins</i>
----------------------	--

---

**Description**

Given the proteomics data, quality of the overlapped marker proteins are evaluated by correlating replicates of fractions.

**Usage**

markerQualityControl(coveredProteins, protein.data)

**Arguments**

coveredProteins character; list of marker proteins, gene symbols, that are covered in 3365 marker proteins.

protein.data data.frame; fractionated proteomics data, rownames are gene symbols associated protein.

**Value**

robustMarkers

**Examples**

```
{
df <- loadData(SubCellBarCode::hcc827Ctrl)
c.prots <- calculateCoveredProtein(rownames(df), markerProteins[,1])
r.markers <- markerQualityControl(c.prots[1:5], df)
}
```

---

mergeCls

*Merge compartment and neighborhood classification*

---

**Description**

Compartment and neighborhood classifications are merged for the single output.

**Usage**

```
mergeCls(compartmentCls, neighborhoodCls)
```

**Arguments**

compartmentCls data.frame; all predictions, including unclassified as well, and probability vectors for each protein in compartment classification

neighborhoodCls data.frame; all predictions, including unclassified as well, and probability vectors for each protein in neighborhood classification

**Value**

cls.df



**Examples**

```
{  
  
  #create mock data  
  com.df <- data.frame(Proteins = "TP53",  
    svm.pred = "N1",  
    S1 = as.numeric(0.02),  
    S2 = as.numeric(0.02),  
    S3 = as.numeric(0.02),  
    S4 = as.numeric(0.02),  
    N1 = as.numeric(0.72),  
    N2 = as.numeric(0.02),  
    N3 = as.numeric(0.02),  
    N4 = as.numeric(0.02),  
    C1 = as.numeric(0.02),  
    C2 = as.numeric(0.02),  
    C3 = as.numeric(0.02),  
    C4 = as.numeric(0.02),  
    C5 = as.numeric(0.02),  
    M1 = as.numeric(0.02),  
    M2 = as.numeric(0.02))  
  
  rownames(com.df) <- "TP53"  
  
  neig.df <- data.frame(Proteins = "TP53",  
    svm.pred.all = "Nuclear",  
    Secretary = as.numeric(0.01),  
    Nuclear = as.numeric(0.95),  
    Cytosol = as.numeric(0.02),  
    Mitochondria = as.numeric(0.02))  
  
  rownames(neig.df) <- "TP53"  
  
  cls.df <- mergeCls(com.df, neig.df)  
  
}
```

---

**mergeProbability***Merge compartment probabilities to neighborhood probabilities*

---

**Description**

Compartment levels classifications are summed up to associated neighborhood levels. It is a helper function.

**Usage**

```
mergeProbability(df)
```

**Arguments**

df                    data.frame; all predictions at the neighborhood level and probability vectors for each protein

**Value**

merged.df

**Examples**

```
{  
  
#create mock data  
df <- data.frame(Protein = "TP53",  
S1 = as.numeric(0.02),  
S2 = as.numeric(0.02),  
S3 = as.numeric(0.02),  
S4 = as.numeric(0.02),  
N1 = as.numeric(0.72),  
N2 = as.numeric(0.02),  
N3 = as.numeric(0.02),  
N4 = as.numeric(0.02),  
C1 = as.numeric(0.02),  
C2 = as.numeric(0.02),  
C3 = as.numeric(0.02),  
C4 = as.numeric(0.02),  
C5 = as.numeric(0.02),  
M1 = as.numeric(0.02),  
M2 = as.numeric(0.02))  
  
rownames(df) <- "TP53"  
  
merged.df <- mergeProbability(df)  
  
}
```

---

plotBarcode

*Visualize the SubCellBarCode*

---

**Description**

Stacked bar plot are plotted for compartment and neighborhood level with respect to classification probabilities.

**Usage**

```
plotBarcode(sampleClassification, protein, s1PSM)
```

**Arguments**

sampleClassification	data.frame; merged classification, combination of compartment and neighborhood classification.
protein	character; protein gene symbol name
s1PSM	data.frame; minimum PSM count table. Row names should be gene centric protein id.

**Value**

proteinPlot

**Examples**

```
{
#create mock data
plot.df <- data.frame(Protein = "TP53",
NeighborhoodCls = "Nuclear",
CompartmentCls = "N1",
Secretory = as.numeric(0.01),
Nuclear = as.numeric(0.95),
Cytosol = as.numeric(0.02),
Mitochondria = as.numeric(0.02),
S1 = as.numeric(0.02),
S2 = as.numeric(0.02),
S3 = as.numeric(0.02),
S4 = as.numeric(0.02),
N1 = as.numeric(0.72),
N2 = as.numeric(0.02),
N3 = as.numeric(0.02),
N4 = as.numeric(0.02),
C1 = as.numeric(0.02),
C2 = as.numeric(0.02),
C3 = as.numeric(0.02),
C4 = as.numeric(0.02),
C5 = as.numeric(0.02),
M1 = as.numeric(0.02),
M2 = as.numeric(0.02))

rownames(plot.df) <- "TP53"

psm.df <- data.frame(Protein = "TP53",
PSMs.for.quant = as.numeric(31))

rownames(psm.df) <- "TP53"

proteinPlot <- plotBarcode(plot.df, "TP53", psm.df)
}
```

---

plotMultipleProtein     *Visualization of multiple protein localizations*

---

### Description

Distributions of subcellular localizations of multiple proteins both at the compartment and neighborhood level are plotted.

### Usage

```
plotMultipleProtein(sampleClassification, proteinList)
```

### Arguments

sampleClassification     data.frame; merged classification, combination of compartment and neighborhood classifications per protein.

proteinList     vector; protein gene symbol names.

### Value

multipleProt.df

### Examples

```
{
  proteasome26s <- c("PSMA7", "PSMC3", "PSMB1", "PSMA1", "PSMA3", "PSMA4",
    "PSMA5", "PSMB4", "PSMB6", "PSMB5", "PSMC2", "PSMC4", "PSMB3", "PSMB2",
    "PSMD4", "PSMA6", "PSMC1", "PSMC5", "PSMC6", "PSMB7", "PSMD13")

  exp.cls.df <- SubCellBarCode::hcc827GEFCClass

  multipleProt.df <- plotMultipleProtein(exp.cls.df, proteasome26s )
}
```

---

replacePrediction     *Replace compartment predictions to neighborhood predictions*

---

### Description

Compartment level classifications are replaced with neighborhood level assignment. It is a helper function.

**Usage**

```
replacePrediction(df, column = c("svm.pred.all", "Observation", "svm.pred"))
```

**Arguments**

`df` data.frame; all predictions at the compartment level and probability vectors for each protein

`column` character; selected column in the data frame, `df`

**Value**

replaced.df

**Examples**

```
{
#define mock data frame
df <- data.frame(svm.pred.all = c("S1","S2","S3","S4",
"N1","N2","N3","N4",
"C1","C2","C3","C4","C5",
"M1","M2"))

df$svm.pred.all <- as.character(df$svm.pred.all)
df$Prob <- "1"

df <- replacePrediction(df, column = "svm.pred.all")
}
```

---

sankeyPlot

*Sankey plot for condition-dependent protein relocation*

---

**Description**

Identify candidate condition-dependent relocated proteins by comparing neighborhood classifications.

**Usage**

```
sankeyPlot(sampleCls1, sampleCls2)
```

**Arguments**

`sampleCls1` data.frame; merged classification, combination of compartment and neighborhood classification.

`sampleCls2` data.frame; merged classification, combination of compartment and neighborhood classification.

**Value**

label.link.df

**Examples**

```
{
  exp.cls.df <- SubCellBarCode::hcc827GEFCClass
  sankeyData <- sankeyPlot(exp.cls.df, exp.cls.df)
}
```

---

sumProbability	<i>Sum compartment test data probabilities to neighborhood probabilities</i>
----------------	--

---

**Description**

Compartment levels classifications on the test data are summed up to associated neighborhood levels. It is a helper function.

**Usage**

```
sumProbability(df)
```

**Arguments**

df                    data.frame; test data classifications at the neighborhood level and probability vectors for each protein.

**Value**

summed.df

**Examples**

```
{
  #create mock data
  df <- data.frame(Protein = "TP53",
    svm.pred = "N1",
    S1 = as.numeric(0.02),
    S2 = as.numeric(0.02),
    S3 = as.numeric(0.02),
    S4 = as.numeric(0.02),
    N1 = as.numeric(0.72),
    N2 = as.numeric(0.02),
    N3 = as.numeric(0.02),
```

```
N4 = as.numeric(0.02),
C1 = as.numeric(0.02),
C2 = as.numeric(0.02),
C3 = as.numeric(0.02),
C4 = as.numeric(0.02),
C5 = as.numeric(0.02),
M1 = as.numeric(0.02),
M2 = as.numeric(0.02))

rownames(df) <- "TP53"

sum.df <- sumProbability(df)

}
```

---

svmClassification	<i>Protein subcellular localization classification</i>
-------------------	--

---

### Description

Support Vector Machine classifier is trained and used for prediction of protein subcellular localization

### Usage

```
svmClassification(markerProteins, protein.data, markerprot.df)
```

### Arguments

markerProteins character; robust marker proteins along with subcellular localization that are present in the given data.

protein.data data.frame; fractionated proteomics data

markerprot.df data.frame; collection of marker proteins along with corresponding subcellular localization

### Value

all.classifications

### Examples

```
{

df <- loadData(SubCellBarCode::hcc827Ctrl)

c.prots <- calculateCoveredProtein(rownames(df), markerProteins[,1])

set.seed(7)
c.prots <- sample(c.prots, 500)
```

```
cls <- svmClassification(c.prots, df, markerProteins)
}
```

---

svmExternalData

*Peptide/exon/transcript centric or PTM enriched classification*

---

### Description

Peptide/exon/transcript centric or PTM enriched classification is applied to predict localization of them.

### Usage

```
svmExternalData(df, modelA, modelB)
```

### Arguments

df	data frame fractionated additional data
modelA	model for the replicate A classification
modelB	model for the replicate B classification

### Value

c.cls.df

### Examples

```
{
df <- loadData(SubCellBarCode::hcc827Ctrl)

c.prots <- calculateCoveredProtein(rownames(df), markerProteins[,1])

set.seed(7)
c.prots <- sample(c.prots, 550)
cls <- svmClassification(c.prots, df, markerProteins)
modelA <- cls[[1]]$model
modelB <- cls[[2]]$model

exon.cls <- svmExternalData(SubCellBarCode::hcc827exon,
modelA = modelA, modelB = modelB)
}
```



---

tsneVisualization	<i>Visualization of marker proteins by t-SNE map</i>
-------------------	--

---

**Description**

The marker proteins are visualized in 3D t-SNE map to see the distributions of the marker proteins.

**Usage**

```
tsneVisualization(protein.data, markerProteins, dims, theta, perplexity)
```

**Arguments**

protein.data	data.frame; fractionated proteomics data
markerProteins	character; robust marker proteins, gene symbols, that are present in the given data and overlapped with package's marker protein list.
dims	integer; dimensionality
theta	numeric; Speed/accuracy trade-off ,increase for less accuracy
perplexity	integer; Perplexity parameter

**Value**

tsneMap.df

**Examples**

```
{  
  
df <- loadData(SubCellBarCode::hcc827Ctrl)  
  
c.prots <- calculateCoveredProtein(rownames(df), markerProteins[,1])  
  
set.seed(21)  
tsneMap.df <- tsneVisualization(protein.data = df,  
markerProteins = c.prots[1:20],  
dims = 2, theta = c(0.4), perplexity = c(5))  
}
```

# Index

## \* datasets

- [hcc827Ctrl](#), [11](#)
- [hcc827CtrlPSMCount](#), [11](#)
- [hcc827exon](#), [12](#)
- [hcc827GEF](#), [13](#)
- [hcc827GEFClass](#), [13](#)
- [hcc827GefPSMCount](#), [14](#)
- [markerProteins](#), [15](#)

- [applyThresholdCompartment](#), [2](#)
- [applyThresholdNeighborhood](#), [3](#)

- [calculateCoveredProtein](#), [4](#)
- [calRowMean](#), [5](#)
- [candidateRelocatedProteins](#), [6](#)
- [compareCls](#), [7](#)
- [computeThresholdCompartment](#), [8](#)
- [computeThresholdNeighborhood](#), [9](#)
- [convert2symbol](#), [10](#)

- [hcc827Ctrl](#), [11](#)
- [hcc827CtrlPSMCount](#), [11](#)
- [hcc827exon](#), [12](#)
- [hcc827GEF](#), [13](#)
- [hcc827GEFClass](#), [13](#)
- [hcc827GefPSMCount](#), [14](#)

- [loadData](#), [14](#)

- [markerProteins](#), [15](#)
- [markerQualityControl](#), [15](#)
- [mergeCls](#), [16](#)
- [mergeProbability](#), [17](#)

- [plotBarcode](#), [18](#)
- [plotMultipleProtein](#), [20](#)

- [replacePrediction](#), [20](#)

- [sankeyPlot](#), [21](#)
- [sumProbability](#), [22](#)

- [svmClassification](#), [23](#)

- [svmExternalData](#), [24](#)

- [tsneVisualization](#), [25](#)