

# synaptome.db

September 11, 2024

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findGeneByCompartmentPaperCnt

*Get gene table of frequently found genes within compartments*

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## Description

Get gene table and paper count for genes mentioned cnt or more times in different compartment-paper pairs.

## Usage

```
findGeneByCompartmentPaperCnt(cnt = 1)
```

## Arguments

cnt                    minimal number of times mentioned gene

## Value

data.frame with 9 columns: 8 specified in [getGenesByID](#) and Npmid column for the paper count.

## See Also

[getGenesByID](#)

Other Gene functions: [findGeneByPaperCnt\(\)](#), [findGeneByPapers\(\)](#), [findGenesByEntrez\(\)](#), [findGenesByName\(\)](#), [getAllGenes4Compartment\(\)](#), [getGenes4Compartment\(\)](#), [getGenesByID\(\)](#)

## Examples

```
cntT <- findGeneByPaperCnt(47)
head(cntT)
```

---

findGeneByPaperCnt      *Get gene table of frequently found genes*

---

**Description**

Get gene table and paper count for genes mentioned cnt or more times in different papers.

**Usage**

```
findGeneByPaperCnt(cnt = 1)
```

**Arguments**

cnt                      minimal number of papers that mentioned gene

**Value**

data.frame with 9 columns: 8 specified in [getGenesByID](#) and Npmid column for the paper count.

**See Also**

[getGenesByID](#)

Other Gene functions: [findGeneByCompartmentPaperCnt\(\)](#), [findGeneByPapers\(\)](#), [findGenesByEntrez\(\)](#), [findGenesByName\(\)](#), [getAllGenes4Compartment\(\)](#), [getGenes4Compartment\(\)](#), [getGenesByID\(\)](#)

**Examples**

```
cntT <- findGeneByPaperCnt(47)
head(cntT)
```

---

findGeneByPapers      *Get gene table of frequently found genes*

---

**Description**

Get gene table and paper count for genes mentioned cnt or more times in different papers.

**Usage**

```
findGeneByPapers(pmids, cnt = 1)
```

**Arguments**

pmids                    vector of PMIDs to search for genes  
cnt                      minimal number of papers that mentioned gene

**Value**

data.frame with 9 columns: 8 specified in [getGenesByID](#) and Npmid column for the paper count.

**See Also**

[getGenesByID](#)

Other Gene functions: [findGeneByCompartmentPaperCnt\(\)](#), [findGeneByPaperCnt\(\)](#), [findGenesByEntrez\(\)](#), [findGenesByName\(\)](#), [getAllGenes4Compartment\(\)](#), [getGenes4Compartment\(\)](#), [getGenesByID\(\)](#)

**Examples**

```
res<- getAllGenes4Compartment(compartmentID = 1)
gnt<-getGeneInfoByIDs(res$GeneID)
pmids<-names(sort(table(gnt$PaperPMID)))[1:5])
cntT <- findGeneByPapers(pmids,cnt=3)
head(cntT)
```

---

findGenesByEntrez      *Internal Gene representation for given list of gene Entrez IDs*

---

**Description**

Get internal gene representation for set of gene Entrez IDs. Function lookups for provided values in Human Entrez ID, Mouse Entrez ID and Rat Entrez ID columns and return following features for all found genes: GeneID (internal database ID), MGI ID, Human Entrez ID, Mouse Entrez ID, Rat Entrez ID, Human gene name, Mouse gene name and Rat gene name.

**Usage**

```
findGenesByEntrez(entrez)
```

**Arguments**

entrez                      vector of Entres IDs. Function accepts both integers and characters.

**Details**

Could be used as an intermediate step for building Protein-Protein interaction map from the list of Gene IDs returned in the first column. Also, this function provides a useful sanity check, e.g. how many Gene IDs correspond to the specific gene name or Entrez ID, which could be specie-specific.

**Value**

data.frame with columns specified above.

**See Also**

Other Lookup functions: [findGenesByName\(\)](#)

Other Gene functions: [findGeneByCompartmentPaperCnt\(\)](#), [findGeneByPaperCnt\(\)](#), [findGeneByPapers\(\)](#), [findGenesByName\(\)](#), [getAllGenes4Compartment\(\)](#), [getGenes4Compartment\(\)](#), [getGenesByID\(\)](#)

**Examples**

```
# get information for specific gene
t <- findGenesByEntrez(c(1742, 1741, 1739, 1740))
```

---

findGenesByName	<i>Find GeneIDs for names</i>
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---

**Description**

Get internal gene representation for set of gene names. Function lookups for provided values in Human Name, Mouse Name and Rat Name columns and return following features for all found genes: GeneID (internal database ID), MGI ID, Human Entrez ID, Mouse Entrez ID, Rat Entrez ID, Human gene name, Mouse gene name and Rat gene name.

**Usage**

```
findGenesByName(name)
```

**Arguments**

name                    vector of gene names.

**Details**

Could be used as an intermediate step for building Protein-Protein interaction map from the list of Gene IDs returned in the first column. Also, this function provides a useful sanity check, e.g. how many Gene IDs correspond to the specific gene name or Entrez ID, which could be specie-specific.

**Value**

data.frame with columns specified above.

**See Also**

Other Lookup functions: [findGenesByEntrez\(\)](#)

Other Gene functions: [findGeneByCompartmentPaperCnt\(\)](#), [findGeneByPaperCnt\(\)](#), [findGeneByPapers\(\)](#), [findGenesByEntrez\(\)](#), [getAllGenes4Compartment\(\)](#), [getGenes4Compartment\(\)](#), [getGenesByID\(\)](#)

**Examples**

```
# Find GeneIDs for names
t <- findGenesByName(c("Src", "Srcin1", "Fyn"))
```

---

`getAllGenes4BrainRegion`*Get all genes for brain region for particular specie*

---

**Description**

Get all genes annotated for specific brain region for specific specie. Function lookups Brain Region ID and Specie Tax ID columns and returns main information like internal Gene IDs, MGI ID, Human Entrez ID, Human Gene Name, Mouse Entrez ID, Mouse Gene Name, Localisation (presynaptic, postsynaptic, synaptosomal), PaperPMID and BrainRegion.

**Usage**

```
getAllGenes4BrainRegion(brainRegion, taxID)
```

**Arguments**

<code>brainRegion</code>	ID for specific brain region
<code>taxID</code>	specie ID

**Value**

data.frame with the following columns:

- GeneID
- Localisation
- MGI
- HumanEntrez
- MouseEntrez
- HumanName
- MouseName
- PMID
- Paper
- Year
- SpeciesTaxID
- BrainRegion

**See Also**

Other BrainRegion functions: [getBrainRegions\(\)](#), [getGenes4BrainRegion\(\)](#), [getPPIbyIDs4BrainRegion\(\)](#)

Other BrainRegion Gene functions: [getGenes4BrainRegion\(\)](#)

**Examples**

```
gns <- getAllGenes4BrainRegion(brainRegion = "Striatum", taxID = 10090)
head(gns)
```

---

`getAllGenes4Compartment`*Extract all genes found in compartment*

---

**Description**

Get all genes annotated for specific compartment. Function returns main information like internal Gene IDs, MGI ID, Human Entrez ID, Human Gene Name, Mouse Entrez ID, Mouse Gene Name, Rat Entrez ID, Rat Gene Name.

**Usage**

```
getAllGenes4Compartment(compartmentID)
```

**Arguments**

compartmentID ID for specific compartment

**Value**

data.frame described in [getGenesByID\(\)](#)

**See Also**

Other Gene functions: [findGeneByCompartmentPaperCnt\(\)](#), [findGeneByPaperCnt\(\)](#), [findGeneByPapers\(\)](#), [findGenesByEntrez\(\)](#), [findGenesByName\(\)](#), [getGenes4Compartment\(\)](#), [getGenesByID\(\)](#)

Other Compartment functions: [getCompartments\(\)](#), [getGenes4Compartment\(\)](#), [getPPIbyIDs4Compartment\(\)](#)

**Examples**

```
G <- getAllGenes4Compartment(compartmentID = 1) # 5560 rows
```

---

`getBrainRegions`*Get list of all Brain regions in the database*

---

**Description**

Get full list of brain regions considered in the database. Returns table with regions and their respective IDs.

**Usage**

```
getBrainRegions()
```

**Value**

data.frame with following columns:

- ID: Brain region internal ID
- Name: name of the region
- Description: extended description of the region
- InterlexID
- ParentID: ID of the containing brain region

**See Also**

Other BrainRegion functions: [getAllGenes4BrainRegion\(\)](#), [getGenes4BrainRegion\(\)](#), [getPPIbyIDs4BrainRegion\(\)](#)

**Examples**

```
t <- getBrainRegions()
```

---

<code>getCompartments</code>	<i>Get full list of compartments</i>
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**Description**

Get list of synaptic compartments considered in the database. Returns table with compartments: “Presynaptic”, “Postsynaptic”, “Synaptosome”, and their internal IDs.

**Usage**

```
getCompartments()
```

**Value**

data.frame described above

**See Also**

Other Compartment functions: [getAllGenes4Compartment\(\)](#), [getGenes4Compartment\(\)](#), [getPPIbyIDs4Compartment\(\)](#)

**Examples**

```
c <- getCompartments()
```

---

`getGeneDiseaseByEntres`*Disease information for Human Entrez IDs*

---

**Description**

Get Human disease information (HDO provided) for the set of Human Entrez IDs. Function lookups for diseases associated with Human Entrez IDs and returns list of available diseases.

**Usage**

```
getGeneDiseaseByEntres(entrez)
```

**Arguments**

entrez            vector of Human Entrez gene IDs

**Value**

data.frame with following columns:

- HumanEntrez
- HumanName
- HDOID
- Description

**See Also**

Other Disease functions: [getGeneDiseaseByIDs\(\)](#), [getGeneDiseaseByName\(\)](#)

**Examples**

```
t <- getGeneDiseaseByEntres(c(8573, 1742, 1739)) # (95 rows)
```

---

`getGeneDiseaseByIDs`*Disease information for internal gene IDs.*

---

**Description**

Get Human disease information (HDO provided) for the set of internal gene IDs. Function lookups for diseases associated with internal GeneIDs and returns list of available diseases.

**Usage**

```
getGeneDiseaseByIDs(ids)
```



**Arguments**

ids                    vector of gene IDs.

**Value**

data.frame with following columns:

- HumanEntrez
- HumanName
- HDOID
- Description

**See Also**

Other Disease functions: [getGeneDiseaseByEntres\(\)](#), [getGeneDiseaseByName\(\)](#)

**Examples**

```
t <- getGeneDiseaseByIDs(c(48, 585, 710))
```

---

*getGeneDiseaseByName*    *Disease information for Human Gene Names*

---

**Description**

Get Human disease information (HDO provided) for the set of Human Gene Names. Function lookups for diseases associated with internal Human gene names and returns list of available diseases.

**Usage**

```
getGeneDiseaseByName(names)
```

**Arguments**

names                    vector of Human gene names

**Value**

data.frame

**See Also**

Other Disease functions: [getGeneDiseaseByEntres\(\)](#), [getGeneDiseaseByIDs\(\)](#)

**Examples**

```
t <- getGeneDiseaseByName(c("CASK", "DLG2", "DLG1")) # (115 rows)
```

---

getGeneInfoByEntrez     *Gene information for given list of gene Entrez IDs*

---

### Description

Get gene information for set of gene Entrez IDs. Function lookup for name in Human Entrez ID and Mouse Entrez Id data and return following features for all found genes: GeneID (internal database ID), Localisation (presynaptic, postsynaptic, synaptosome), MGI (MGI ID), HumanEntrez (Human Entrez ID), MouseEntrez (Mouse Entrez ID), HumanName (Human gene name), MouseName (Mouse gene name), PaperPMID (PMID IDs for the publications where the genes were reported), Paper (papers where specific genes were reported in a format FIRS TAUTHOR\_YEAR), Year, SpeciesTaxID (specie the original experiment was performed on), BrainRegion (Brain region where the specific genes were identified, according to the paper)

### Usage

```
getGeneInfoByEntrez(entrez)
```

### Arguments

entrez                vector of Entres IDs. Function accepts both integers and characters.

### Value

data.frame with fields specified above.

### See Also

Other GeneInfo functions: [getGeneInfoByIDs\(\)](#), [getGeneInfoByName\(\)](#), [getGeneInfoByPapers\(\)](#)

### Examples

```
# get information for specific gene
t <- getGeneInfoByEntrez(1742)
# get information for specific character string Entres representation
t <- getGeneInfoByEntrez("1742")

# get information for the list of genes
t <- getGeneInfoByName(c(1741, 1742, 1739, 1740))
```

---

`getGeneInfoByIDs`*Get GeneInfo table for set of GeneIDs*

---

**Description**

Function lookup for internal GeneID values and return following features for all found genes: GeneID (internal database ID), Localisation (one of the following: presynaptic, postsynaptic, synaptosome), MGI (MGI ID), HumanEntrez (Human Entrez ID), MouseEntrez (Mouse Entrez ID), HumanName (Human gene name), MouseName (Mouse gene name), PaperPMID (PMID IDs for the publications where the genes were reported), Paper (papers where specific genes were reported in a format FIRS<sub>T</sub>AUTHOR\_YEAR), Year, SpeciesTaxID (specie the original experiment was performed on), BrainRegion (Brain region where the specific genes were identified, according to the paper)

**Usage**

```
getGeneInfoByIDs(ids)
```

**Arguments**

`ids`                    vector of Gene IDs.

**Details**

This function then returns following features for all found genes:

- GeneID,
- Localisation,
- MGI,
- HumanEntrez,
- MouseEntrez,
- HumanName,
- MouseName,
- PaperPMID,
- Paper,
- Year,
- SpeciesTaxID,
- BrainRegion

**Value**

`data.frame` with column specified above.

**See Also**

Other GeneInfo functions: [getGeneInfoByEntrez\(\)](#), [getGeneInfoByName\(\)](#), [getGeneInfoByPapers\(\)](#)

**Examples**

```
gdf <- getGeneInfoByIDs(c(46, 6, 15, 1))
```

---

getGeneInfoByName	<i>Get gene information for set of gene names.</i>
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---

**Description**

Function lookup for name in Human Gene name, Rat Gene name and Mouse Gene name data and return following features for all found genes: GeneID (internal database ID), Localisation (one of the following: presynaptic, postsynaptic, synaptosome), MGI (MGI ID), HumanEntrez (Human Entrez ID), MouseEntrez (Mouse Entrez ID), HumanName (Human gene name), MouseName (Mouse gene name), PaperPMID (PMID IDs for the publications where the genes were reported), Paper (papers where specific genes were reported in a format FIRSTAUTHOR\_YEAR), Year, SpeciesTaxID (specie the original experiment was performed on), BrainRegion (Brain region where the specific genes were identified, according to the paper)

**Usage**

```
getGeneInfoByName(name)
```

**Arguments**

name	vector of gene names
------	----------------------

**Details**

This function then returns following features for all found genes:

- GeneID,
- Localisation,
- MGI,
- HumanEntrez,
- MouseEntrez,
- HumanName,
- MouseName,
- PaperPMID,
- Paper,
- Year,
- SpeciesTaxID,
- BrainRegion

**Value**

data.frame with fields specified above.

**See Also**

Other GeneInfo functions: [getGeneInfoByEntrez\(\)](#), [getGeneInfoByIDs\(\)](#), [getGeneInfoByPapers\(\)](#)

**Examples**

```
# get information for specific gene
t <- getGeneInfoByName("CASK")

# get information for the list of genes
t <- getGeneInfoByName(c("CASK", "DLG2"))
```

---

getGeneInfoByPapers     *Get gene information for set of genes mentioned by certain papers.*

---

**Description**

Function lookup for specified PubMedIDs in the gene reference data and return following features for genes referenced by requested papers at least cnt times: GeneID (internal database ID), Localisation (one of the following: presynaptic, postsynaptic, synaptosome), MGI (MGI ID), HumanEntrez (Human Entrez ID), MouseEntrez (Mouse Entrez ID), HumanName (Human gene name), MouseName (Mouse gene name), PaperPMID (PMID IDs for the publications where the genes were reported if it is within pmids list), Paper (papers where specific genes were reported in a format FIRSAUTHOR\_YEAR), Year, SpeciesTaxID (specie the original experiment was performed on), BrainRegion (Brain region where the specific genes were identified, according to the paper)

**Usage**

```
getGeneInfoByPapers(pmids, cnt = 1)
```

**Arguments**

pmids	vector of PMIDs to search for genes
cnt	minimal number of papers that mentioned gene

**Details**

This function then returns following features for all found genes:

- GeneID,
- Localisation,
- MGI,
- HumanEntrez,

- MouseEntrez,
- HumanName,
- MouseName,
- PaperPMID,
- Paper,
- Year,
- SpeciesTaxID,
- BrainRegion

### Value

data.frame with fields specified above.

### See Also

Other GeneInfo functions: [getGeneInfoByEntrez\(\)](#), [getGeneInfoByIDs\(\)](#), [getGeneInfoByName\(\)](#)

### Examples

```
res<- getAllGenes4Compartment(compartmentID = 1)
gnt<-getGeneInfoByIDs(res$GeneID)
pmids<-names(sort(table(gnt$PaperPMID))[1:5])
cntT <- getGeneInfoByPapers(pmids,cnt=3)
head(cntT)
```

---

getGenes4BrainRegion *Select genes from the list that found in brain region of particular specie*

---

### Description

Select genes from your list annotated for specific brain region. Should be used with [findGenesByEntrez\(\)](#) or [findGenesByName\(\)](#) functions to obtain list of internal IDs for your list of genes. Function lookups the Gene table for specified localisation and returns main gene information like internal Gene IDs, MGI ID, Human Entrez ID, Human Gene Name, Mouse Entrez ID, Mouse Gene Name, PaperPMID, Localisation and BrainRegion.

### Usage

```
getGenes4BrainRegion(ids, brainRegion, taxID)
```

### Arguments

ids	internal IDs for list of genes
brainRegion	ID for the brain region of interest
taxID	taxon ID specie tax ID

**Value**

data.frame with the following columns:

- GeneID
- Localisation
- MGI
- HumanEntrez
- MouseEntrez
- HumanName
- MouseName
- PMID
- Paper
- Year
- SpeciesTaxID
- BrainRegion

**See Also**

Other BrainRegion functions: [getAllGenes4BrainRegion\(\)](#), [getBrainRegions\(\)](#), [getPPIbyIDs4BrainRegion\(\)](#)

Other BrainRegion Gene functions: [getAllGenes4BrainRegion\(\)](#)

**Examples**

```
Genes <- getGenes4BrainRegion(c(1, 15, 156, 1500, 3000, 7000),  
  brainRegion = "Striatum", taxID = 10090  
) # 5 rows
```

---

getGenes4Compartment *Select genes from the list that found in compartment*

---

**Description**

Select all genes from your list annotated for specific compartment. Should be used with [findGenesByEntrez\(\)](#) or [findGenesByName\(\)](#) functions to obtain list of internal IDs for your list of genes. Function lookups the Gene table for specified localisation and returns main gene information like internal Gene IDs, MGI ID, Human Entrez ID, Human Gene Name, Mouse Entrez ID, Mouse Gene Name, Rat Entrez ID, Rat Gene Name.

**Usage**

```
getGenes4Compartment(ids, compartmentID)
```

**Arguments**

`ids`                    Gene IDs  
`compartmentID`    compartment ID

**Value**

data.frame described in [getGenesByID\(\)](#)

**See Also**

Other Gene functions: [findGeneByCompartmentPaperCnt\(\)](#), [findGeneByPaperCnt\(\)](#), [findGeneByPapers\(\)](#), [findGenesByEntrez\(\)](#), [findGenesByName\(\)](#), [getAllGenes4Compartment\(\)](#), [getGenesByID\(\)](#)

Other Compartment functions: [getAllGenes4Compartment\(\)](#), [getCompartments\(\)](#), [getPPIbyIDs4Compartment\(\)](#)

**Examples**

```
Genes <- getGenes4Compartment(c(1, 15, 156, 1500, 3000, 7000),
  compartmentID = 1
)
```

---

getGenesByID

*Get gene table from list of GeneIDs.*

---

**Description**

Takes internal gene IDs as input and return the following features for all found genes:

**GeneID** internal database ID

**MGI** MGI ID

**HumanEntrez** Human Entrez ID

**MouseEntrez** Mouse Entrez ID

**HumanName** Human gene name

**MouseName** Mouse gene name

**RatEntrez** Rat Entrez ID

**Rat Name** Rat gene name

**Usage**

```
getGenesByID(ids)
```

**Arguments**

`ids`                    vector of GeneID values.



**Value**

data.frame with 8 columns specified above.

**See Also**

Other Gene functions: [findGeneByCompartmentPaperCnt\(\)](#), [findGeneByPaperCnt\(\)](#), [findGeneByPapers\(\)](#), [findGenesByEntrez\(\)](#), [findGenesByName\(\)](#), [getAllGenes4Compartment\(\)](#), [getGenes4Compartment\(\)](#)

**Examples**

```
gdf <- getGenesByID(c(46, 6, 15, 1))
```

---

<code>getIGraphFromPPI</code>	<i>Get Igraph representation of PPI</i>
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---

**Description**

Combine information from PPI data.frame obtained with functions like [getPPIbyName](#) or [getPPIbyEntrez](#) with information about genes obtained from [getGenesByID](#) to make interpretable undirected PPI graph in [igraph](#) format. In this format network could be further analysed and visualized by algorithms in [igraph](#) package.

**Usage**

```
getIGraphFromPPI(ppi)
```

**Arguments**

`ppi` PPI data.frame with columns A and B, obtained from functions like [getPPIbyName](#)

**Value**

[igraph](#) object with specified PPI network.

**See Also**

[getPPIbyName\(\)](#), [getPPIbyEntrez\(\)](#) and [getPPIbyIDs\(\)](#) to get PPI data.frame, [getTableFromPPI\(\)](#) to get interpretable table representation of the PPI data.frame.

Other PPI functions: [getPPIbyEntrez\(\)](#), [getPPIbyIDs4BrainRegion\(\)](#), [getPPIbyIDs4Compartment\(\)](#), [getPPIbyIDs\(\)](#), [getPPIbyName\(\)](#), [getTableFromPPI\(\)](#)

**Examples**

```
library(igraph)
g <- getIGraphFromPPI(
  getPPIbyIDs(c(48, 129, 975, 4422, 5715, 5835), type = "lim")
)
plot(g, vertex.label = V(g)$RatName, vertex.size = 35)
```

---

`getMutations4DiseaseByEntres`*Get mutational information for Gene and Disease*

---

**Description**

Function looks up the information for particular set of Entrez IDs and disease HDOID.

**Usage**

```
getMutations4DiseaseByEntres(entrez, hdoid)
```

**Arguments**

entrez	list of Entrez IDs for genes to select
hdoid	disease HDOID to get mutational association.

**Value**

data.frame as described in [getMutations4DiseaseByIDs](#).

**See Also**

[findGenesByEntrez](#)

Other Mutation functions: [getMutations4DiseaseByIDs\(\)](#), [getMutations4DiseaseByName\(\)](#)

**Examples**

```
hdoid<-'D0ID:0060041'  
entrez<-c("23859", "17754", "18673", "268566", "12293", "320840", "24012")  
mdf<-getMutations4DiseaseByEntres(entrez, hdoid)  
is.null(mdf)
```

---

`getMutations4DiseaseByIDs`*Get mutational information for Gene and Disease*

---

**Description**

Function looks up the information for particular set of GeneIDs and disease HDOID.

**Usage**

```
getMutations4DiseaseByIDs(ids, hdoid)
```

**Arguments**

<code>ids</code>	internal GeneIDs to filter mutation data
<code>hdoid</code>	disease HDOID to get mutational association.

**Details**

This function then returns following features for all found genes:

- GeneID,
- MGI,
- MouseEntrez,
- MouseName,
- HumanName,
- HumanEntrez,
- HDOID,
- Disease,
- Chromosome,
- Position,
- Variant,
- FunctionClass,
- cDNAvariant,
- ProteinVariant,
- DENOVO,
- SFARI,
- EpilepsyGene,
- ClinVar,
- PMID,
- Paper

**Value**

data.frame with fields specified above.

**See Also**

Other Mutation functions: [getMutations4DiseaseByEntres\(\)](#), [getMutations4DiseaseByName\(\)](#)

**Examples**

```
hdoid<-'DOID:0060041'  
ids<-c(6,32,127,181,240,267,558)  
mdf<-getMutations4DiseaseByIDs(ids, hdoid)  
is.null(mdf)
```

---

```
getMutations4DiseaseByName
```

*Get mutational information for Gene and Disease*

---

### Description

Function lookups for provided values in Human Name, Mouse Name and Rat Name columns, extract known mutations, filter requested HDOID and return data.frame in the format described in [getMutations4DiseaseByIDs](#).

### Usage

```
getMutations4DiseaseByName(name, hdoid)
```

### Arguments

name	gene names to look for
hdoid	disease HDOID to get mutational association.

### Value

data.frame as described in [getMutations4DiseaseByIDs](#).

### See Also

[findGenesByName](#)

Other Mutation functions: [getMutations4DiseaseByEntres\(\)](#), [getMutations4DiseaseByIDs\(\)](#)

### Examples

```
hdoid<-'DOID:0060041'
name<-c("Dlg2", "Map1a", "Phb", "Gphn", "Cacna2d1", "Negr1", "Rgs7")
mdf<-getMutations4DiseaseByName(name, hdoid)
is.null(mdf)
```

---

```
getPPIbyEntrez
```

*Extract the PPIs for my list of genes defined by Entrez IDs*

---

### Description

Get Protein-Protein interactions (PPIs) for the set of ID. Function lookups for PPIs for the list of Entrez IDs and returns either “induced” (all available interactions for these genes) or “limited” (only interactions between genes specified in the query) table of interacting genes A and B, where A and B are respective Internal IDs.

**Usage**

```
getPPIbyEntrez(entrez, type = c("induced", "limited"))
```

**Arguments**

entrez	Entrez IDs
type	type of the PPI network should be either induced (for all the PPIs for specific genes, including external genes) or limited (for PPIs between the genes specified in the query). Type could be shortened to recognizable minimum like 'ind' or 'lim'.

**Value**

data.frame with interactors internal GeneID in columns A and B

**See Also**

[getPPIbyName\(\)](#) and [getPPIbyIDs\(\)](#) to get PPI data.frame, [getIGraphFromPPI\(\)](#) to get igraph representation of the PPI data.frame and [getTableFromPPI\(\)](#) to get interpretable table representation of the PPI data.frame.

Other PPI functions: [getIGraphFromPPI\(\)](#), [getPPIbyIDs4BrainRegion\(\)](#), [getPPIbyIDs4Compartment\(\)](#), [getPPIbyIDs\(\)](#), [getPPIbyName\(\)](#), [getTableFromPPI\(\)](#)

**Examples**

```
t <- getPPIbyEntrez(c(1739, 1740, 1742, 1741), type = "ind")
```

---

```
getPPIbyIDs
```

*Extract the PPIs for my list of genes defined by GeneID*

---

**Description**

Get Protein-Protein interactions (PPIs) for the set of internal GeneIDs. Function lookups for PPIs for specific GeneID and returns either "induced" (all interaction for this GeneID) or "limited" (only interactions between GeneIDs specified in the query) table of A and B interacting genes, where A and B are respective GeneIDs.

**Usage**

```
getPPIbyIDs(ids, type = c("induced", "limited"))
```

**Arguments**

ids	Gene IDs
type	type of the PPI network should be either induced (for all the PPIs for specific genes, including external genes) or limited (for PPIs between the genes specified in the query). Type could be shortened to recognizable minimum like 'ind' or 'lim'.

**Value**

data.frame with interactors internal GeneID in columns A and B

**See Also**

[getPPIbyName\(\)](#) and [getPPIbyEntrez\(\)](#) to get PPI data.frame, [getIGraphFromPPI\(\)](#) to get igraph representation of the PPI data.frame and [getTableFromPPI\(\)](#) to get interpretable table representation of the PPI data.frame.

Other PPI functions: [getIGraphFromPPI\(\)](#), [getPPIbyEntrez\(\)](#), [getPPIbyIDs4BrainRegion\(\)](#), [getPPIbyIDs4Compartment\(\)](#), [getPPIbyName\(\)](#), [getTableFromPPI\(\)](#)

**Examples**

```
t <- getPPIbyIDs(c(48, 585, 710), type = "limited") # (16 rows)
t <- getPPIbyIDs(c(48, 585, 710), type = "induced") # 306 rows
```

---

```
getPPIbyIDs4BrainRegion
```

*Prepare induced or limited network for brain region*

---

**Description**

Prepare PPI network for genes from your list annotated for specific brain region. Should be used with [findGenesByEntrez\(\)](#) or [findGenesByName\(\)](#) functions to obtain list of internal IDs for your list of genes. Could be used with [getAllGenes4BrainRegion](#) functions to obtain all genes belonging to respective brain region. Function lookups the PPI table for gene IDs from the list and returns “limited” or “induced” interactors GeneIDs for the specified region.

**Usage**

```
getPPIbyIDs4BrainRegion(
  ids,
  brainRegion,
  taxID,
  type = c("induced", "limited")
)
```

**Arguments**

ids	gene IDs
brainRegion	region ID
taxID	taxon ID
type	type of the PPI network should be either induced (for all the PPIs for specific genes, including external genes) or limited (for PPIs between the genes specified in the query). Type could be shortened to recognizable minimum like 'ind' or 'lim'.

**Value**

data.frame with interactors internal GeneID in columns A and B

**See Also**

Other PPI functions: [getIGraphFromPPI\(\)](#), [getPPIbyEntrez\(\)](#), [getPPIbyIDs4Compartment\(\)](#), [getPPIbyIDs\(\)](#), [getPPIbyName\(\)](#), [getTableFromPPI\(\)](#)

Other BrainRegion functions: [getAllGenes4BrainRegion\(\)](#), [getBrainRegions\(\)](#), [getGenes4BrainRegion\(\)](#)

**Examples**

```
# getting all genes for mouse Striatum
gns <- getAllGenes4BrainRegion(brainRegion = "Striatum", taxID = 10090)
head(gns)

# getting full PPI network for postsynaptic compartment
ppi <- getPPIbyIDs4BrainRegion(
  gns$GeneID,
  brainRegion = "Striatum",
  taxID = 10090,
  type = "limited"
)
head(ppi)
```

---

```
getPPIbyIDs4Compartment
```

*Prepare induced or limited network for compartment*

---

**Description**

Prepare PPI network for genes from your list annotated for specific compartment. Should be used with `findGenesByEntrez` or `findGenesByName` functions to obtain list of internal IDs for your list of genes. Could be used with `getAllGenes4Compartment` functions to obtain all genes belonging to respective compartment. Function lookups the PPI table for gene IDs from the list and returns “limited” or “induced” interactors GeneIDs from the same compartment.

**Usage**

```
getPPIbyIDs4Compartment(ids, compartmentID, type = c("induced", "limited"))
```

**Arguments**

<code>ids</code>	internal gene IDs
<code>compartmentID</code>	ID for specific compartment
<code>type</code>	type of the PPI network should be either ‘induced’ (for all the PPIs for specific genes, including external genes) or ‘limited’ (for PPIs between the genes specified in the query). Type could be shortened to recognizable minimum like ‘ind’ or ‘lim’.

**Value**

data.frame with interactors internal GeneID in columns A and B

**See Also**

Other PPI functions: [getIGraphFromPPI\(\)](#), [getPPIbyEntrez\(\)](#), [getPPIbyIDs4BrainRegion\(\)](#), [getPPIbyIDs\(\)](#), [getPPIbyName\(\)](#), [getTableFromPPI\(\)](#)

Other Compartment functions: [getAllGenes4Compartment\(\)](#), [getCompartments\(\)](#), [getGenes4Compartment\(\)](#)

**Examples**

```
ppi <- getPPIbyIDs4Compartment(c(1, 15, 156, 1500, 3000, 7000),
  compartmentID = 1, type = "induced"
) # 201 rows
```

---

getPPIbyName

*Extract the PPIs for my list of genes defined by Gene name*

---

**Description**

Get Protein-Protein interactions (PPIs) for the set of gene names. Function lookups for PPIs for the list of GeneIDs and returns either “induced” (all interaction for this GeneID) or “limited” (only interactions between GeneIDs specified in the query) table of interacting genes A and B, where A and B are respective gene names.

**Usage**

```
getPPIbyName(name, type = c("induced", "limited"))
```

**Arguments**

name	Gene names
type	type of the PPI network should be either induced (for all the PPIs for specific genes, including external genes) or limited (for PPIs between the genes specified in the query). Type could be shortened to recognizable minimum like 'ind' or 'lim'.

**Value**

data.frame with interactors internal GeneID in columns A and B

**See Also**

[getPPIbyEntrez\(\)](#) and [getPPIbyIDs\(\)](#) to get PPI data.frame, [getIGraphFromPPI\(\)](#) to get igrph representation of the PPI data.frame and [getTableFromPPI\(\)](#) to get interpretable table representation of the PPI data.frame.

Other PPI functions: [getIGraphFromPPI\(\)](#), [getPPIbyEntrez\(\)](#), [getPPIbyIDs4BrainRegion\(\)](#), [getPPIbyIDs4Compartment\(\)](#), [getPPIbyIDs\(\)](#), [getTableFromPPI\(\)](#)



**Examples**

```
t <- getPPIbyName(  
  c("CASK", "DLG4", "GRIN2A", "GRIN2B", "GRIN1"),  
  type = "lim"  
)
```

---

getPapers

*Get synaptome papers overview*

---

**Description**

Get synaptome papers overview

**Usage**

```
getPapers()
```

**Value**

data.frame with following columns:

- PaperPMID
- SpeciesTaxID
- Year
- Name
- Localisation
- BrainRegion
- Method
- Ngenes

**Examples**

```
p <- getPapers()  
head(p)
```

---

`getTableFromPPI`*Get table representation of the PPI.*

---

**Description**

Combine information from PPI data .frame obtained with functions like [getPPIbyName](#) or [getPPIbyEntrez](#) with information about genes obtained from [getGenesByID](#) to make interpretable table representation.

**Usage**

```
getTableFromPPI(ppi)
```

**Arguments**

`ppi` PPI data .frame with columns A and B, obtained from functions like [getPPIbyName](#)

**Value**

table with the following columns added for both interactors (A and B):

- GeneID internal database ID
- MGI MGI ID
- HumanEntrez Human Entrez ID
- MouseEntrez Mouse Entrez ID
- HumanName Human gene name
- MouseName Mouse gene name
- RatEntrez Rat Entrez ID
- Rat Name Rat gene name

**See Also**

[getPPIbyName\(\)](#), [getPPIbyEntrez\(\)](#) and [getPPIbyIDs\(\)](#) to get PPI data .frame, [getIGraphFromPPI\(\)](#) to get igraph representation of the PPI data .frame.

Other PPI functions: [getIGraphFromPPI\(\)](#), [getPPIbyEntrez\(\)](#), [getPPIbyIDs4BrainRegion\(\)](#), [getPPIbyIDs4Compartment\(\)](#), [getPPIbyIDs\(\)](#), [getPPIbyName\(\)](#)

**Examples**

```
tbl <- getTableFromPPI(getPPIbyIDs(c(48, 585, 710), type = "limited"))
```

---

`graphFromSynaptomeByEntrez`*Utility function to create network from [synaptome.db](#) data*

---

**Description**

Utility function to create network from [synaptome.db](#) data

**Usage**

```
graphFromSynaptomeByEntrez(entrez, LCC = TRUE, simplify = TRUE)
```

**Arguments**

<code>entrez</code>	vector of EntrezIDs for network vertices
<code>LCC</code>	if TRUE only largest connected component is returned
<code>simplify</code>	if TRUE loops and multiple edges will be removed

**Value**

network defined by the gene table

**Examples**

```
library(synaptome.db)
cid<-match('Presynaptic', getCompartments())$Name)
geneTable<-getAllGenes4Compartment(cid)
gg<-graphFromSynaptomeByEntrez(geneTable$HumanEntrez)
```

---

`graphFromSynaptomeGeneTable`*Utility function to create network from [synaptome.db](#) data*

---

**Description**

Utility function to create network from [synaptome.db](#) data

**Usage**

```
graphFromSynaptomeGeneTable(
  geneTable,
  LCC = FALSE,
  simplify = FALSE,
  type = c("induced", "limited")
)
```

**Arguments**

geneTable	data.frame described in <a href="#">getGenesByID</a>
LCC	if TRUE only largest connected component is returned
simplify	if TRUE loops and multiple edges will be removed
type	type of the PPI network should be either 'induced' (for all the PPIs for specific genes, including external genes) or 'limited' (for PPIs between the genes specified in the query). Type could be shortened to recognizable minimum like 'ind' or 'lim'.

**Value**

network defined by the gene table

**See Also**

[[getPPIbyIDs\(\)](#)]

**Examples**

```
library(synaptome.db)
cid<-match('Presynaptic', getCompartments())$Name)
geneTable<-getAllGenes4Compartment(cid)
gg<-graphFromSynaptomeGeneTable(geneTable)
```

---

synaptome.db	<i>synaptome.db: programmatic access to the Synaptic proteome database</i>
--------------	--

---

**Description**

The package obtains a local copy of the Synaptic proteome database from AnnotationHub package [synaptome.data](#) and provides a set of utility R functions to query and analyse its content.

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**References**

\insertRefSorokina:2021hlsynaptome.db

**See Also**

Useful links:

- Report bugs at <https://github.com/lptolik/synaptome.db/issues/>

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