

# Package ‘syntenet’

September 19, 2024

**Title** Inference And Analysis Of Synteny Networks

**Version** 1.7.0

**Date** 2022-03-28

**Description** syntenet can be used to infer synteny networks from whole-genome protein sequences and analyze them. Anchor pairs are detected with the MCScanX algorithm, which was ported to this package with the Rcpp framework for R and C++ integration. Anchor pairs from synteny analyses are treated as an undirected unweighted graph (i.e., a synteny network), and users can perform: i. network clustering; ii. phylogenomic profiling (by identifying which species contain which clusters) and; iii. microsynteny-based phylogeny reconstruction with maximum likelihood.

**License** GPL-3

**URL** <https://github.com/almeidasilvaf/syntenet>

**BugReports** <https://support.bioconductor.org/t/syntenet>

**biocViews** Software, NetworkInference, FunctionalGenomics, ComparativeGenomics, Phylogenetics, SystemsBiology, GraphAndNetwork, WholeGenome, Network

**Encoding** UTF-8

**Roxygen** list(markdown = TRUE)

**RoxygenNote** 7.3.1

**Imports** Rcpp (>= 1.0.8), BiocParallel, GenomicRanges, rlang, Biostrings, rtracklayer, utils, methods, igraph, stats, grDevices, RColorBrewer, pheatmap, ggplot2, ggnetwork, intergraph, networkD3

**Suggests** BiocStyle, ggtree, labdsv, covr, knitr, rmarkdown, testthat (>= 3.0.0), xml2

**Config/testthat/edition** 3

**VignetteBuilder** knitr

**LinkingTo** Rcpp, testthat

**NeedsCompilation** yes

**Depends** R (>= 4.2)

**LazyData** false

**git\_url** <https://git.bioconductor.org/packages/syntenet>

**git\_branch** devel

**git\_last\_commit** 2b17e15

**git\_last\_commit\_date** 2024-04-30

**Repository** Bioconductor 3.20

**Date/Publication** 2024-09-18

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## Contents

syntenet-package . . . . .	3
angiosperm_phylogeny . . . . .	3
annotation . . . . .	4
binarize_and_transpose . . . . .	4
blast_list . . . . .	5
check_input . . . . .	5
clusters . . . . .	6
cluster_network . . . . .	7
collapse_protein_ids . . . . .	7
create_species_id_table . . . . .	8
diamond_is_installed . . . . .	9
edges . . . . .	10
export_sequences . . . . .	10
fasta2AAStringSetlist . . . . .	11
find_GS_clusters . . . . .	11
gff2GRangesList . . . . .	12
infer_microsynteny_phylogeny . . . . .	13
infer_syntenet . . . . .	14
interspecies_synteny . . . . .	15
intraspecies_synteny . . . . .	16
iqtree_is_installed . . . . .	17
iqtree_version . . . . .	18
last_is_installed . . . . .	18
network . . . . .	19
parse_collinearity . . . . .	19
phylogenomic_profile . . . . .	20
plot_network . . . . .	21
plot_profiles . . . . .	22
process_input . . . . .	24
profiles2phylip . . . . .	25
proteomes . . . . .	26
rcpp_mcscanx_file . . . . .	26
read_diamond . . . . .	28
run_diamond . . . . .	28
run_last . . . . .	29
scerevisiae_annot . . . . .	30

<i>syntenet-package</i>	3
<i>scerevisiae_diamond</i> . . . . .	31
<b>Index</b>	<b>32</b>

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<i>syntenet-package</i>	<i>syntenet: Inference And Analysis Of Synteny Networks</i>
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## Description

*syntenet* can be used to infer synteny networks from whole-genome protein sequences and analyze them. Anchor pairs are detected with the MCScanX algorithm, which was ported to this package with the Rcpp framework for R and C++ integration. Anchor pairs from synteny analyses are treated as an undirected unweighted graph (i.e., a synteny network), and users can perform: i. network clustering; ii. phylogenomic profiling (by identifying which species contain which clusters) and; iii. microsynteny-based phylogeny reconstruction with maximum likelihood.

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## See Also

Useful links:

- <https://github.com/almeidasilvaf/syntenet>
- Report bugs at <https://support.bioconductor.org/t/syntenet>

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<i>angiosperm_phylogeny</i>	<i>Microsynteny-based angiosperm phylogeny.</i>
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---

## Description

Original tree file obtained from Zhao et al., 2021. The tree is an object of class 'phylo', which can be created by reading the tree file with `treeio::read.tree()`.

## Usage

```
data(angiosperm_phylogeny)
```

## Format

An object of class 'phylo'.

**References**

Zhao, T., Zwaenepoel, A., Xue, J. Y., Kao, S. M., Li, Z., Schranz, M. E., & Van de Peer, Y. (2021). Whole-genome microsynteny-based phylogeny of angiosperms. *Nature Communications*, 12(1), 1-14.

**Examples**

```
data(angiosperm_phylogeny)
```

---

annotation	<i>Filtered genome annotation for <i>Ostreococcus</i> sp. species</i>
------------	---

---

**Description**

Data obtained from Pico-PLAZA 3.0. Only annotation data for primary transcripts were included, and only genes for chromosomes 1, 2, and 3.

**Usage**

```
data(annotation)
```

**Format**

A CompressedGRangesList containing the elements *O*lucimarinus, *O*sp\_RCC809, and *O*tauri.

**References**

Van Bel, M., Silvestri, F., Weitz, E. M., Kreft, L., Botzki, A., Coppens, F., & Vandepoele, K. (2021). PLAZA 5.0: extending the scope and power of comparative and functional genomics in plants. *Nucleic acids research*.

**Examples**

```
data(annotation)
```

---

binarize_and_transpose	<i>Binarize and transpose the phylogenomic profile matrix</i>
------------------------	---

---

**Description**

Binarize and transpose the phylogenomic profile matrix

**Usage**

```
binarize_and_transpose(profile_matrix = NULL)
```

**Arguments**

`profile_matrix` A matrix with phylogenomic profiles obtained with `phylogenomic_profile`.

**Value**

A binary and transposed version of the profiles matrix.

**Examples**

```
data(clusters)
profile_matrix <- phylogenomic_profile(clusters)
tmat <- binarize_and_transpose(profile_matrix)
```

---

blast\_list

*List of data frames containing BLAST-like tabular output*


---

**Description**

The object was created by running `run_diamond` on the protein sequences for the *Ostreococcus* algae available in the **proteomes** example data. Hits with <50% identity were filtered out. Code to recreate this data is available at the `script/` subdirectory.

**Usage**

```
data(blast_list)
```

**Format**

A list of data frames containing the pairwise comparisons between proteomes of *Ostreococcus* species.

**Examples**

```
data(blast_list)
```

---

check\_input

*Check if input objects are ready for further analyses*


---

**Description**

Check if input objects are ready for further analyses

**Usage**

```
check_input(seq = NULL, annotation = NULL, gene_field = "gene_id")
```

**Arguments**

seq	A list of <code>AAStringSet</code> objects, each list element containing protein sequences for a given species. This list must have names (not <code>NULL</code> ), and names of each list element must match the names of list elements in <b>annotation</b> .
annotation	A <code>GRangesList</code> , <code>CompressedGRangesList</code> , or list of <code>GRanges</code> with the annotation for the sequences in <b>seq</b> . This list must have names (not <code>NULL</code> ), and names of each list element must match the names of list elements in <b>seq</b> .
gene_field	Character, name of the column in the <code>GRanges</code> objects that contains gene IDs. Default: "gene_id".

**Details**

This function checks the input data for 3 required conditions:

1. Names of **seq** list (i.e., `names(seq)`) match the names of **annotation** `GRangesList/CompressedGRangesList` (i.e., `names(annotation)`)
2. For each species (list elements), the number of sequences in **seq** is not greater than the number of genes in **annotation**. This is a way to ensure users do not input the translated sequences for multiple isoforms of the same gene (generated by alternative splicing). Ideally, the number of sequences in **seq** should be equal to the number of genes in **annotation**, but this may not always stand true because of non-protein-coding genes.
3. For each species, sequence names (i.e., `names(seq[[x]])`, equivalent to FASTA headers) match gene names in **annotation**.

**Value**

TRUE if the objects pass the check.

**Examples**

```
data(annotation)
data(proteomes)
check_input(proteomes, annotation)
```

---

clusters

*Synteny network clusters of BUSCO genes for 25 eudicot species*

---

**Description**

Data obtained from Zhao & Schranz, 2019.

**Usage**

```
data(clusters)
```

**Format**

A 2-column data frame containing the following variables:

**Gene** Gene ID

**Cluster** Cluster ID

**References**

Zhao, T., & Schranz, M. E. (2019). Network-based microsynteny analysis identifies major differences and genomic outliers in mammalian and angiosperm genomes. *Proceedings of the National Academy of Sciences*, 116(6), 2165-2174.

**Examples**

```
data(clusters)
```

---

cluster_network	<i>Cluster the syntenic network using the Infomap algorithm</i>
-----------------	---

---

**Description**

Cluster the syntenic network using the Infomap algorithm

**Usage**

```
cluster_network(
  network = NULL,
  clust_function = igraph::cluster_infomap,
  clust_params = NULL
)
```

**Arguments**

network	A network represented as an edge list, which is a 2-column data frame with node 1 in the first column and node 2 in the second column. In a syntenic network, node 1 and node 2 are the anchor pairs.
clust_function	Function to be used to cluster the network. It must be one of the functions from the <b>cluster_*</b> family in the <b>igraph</b> package (e.g., cluster_infomap, cluster_leiden, etc). Default: igraph::cluster_infomap.
clust_params	A list with additional parameters (if any) to be passed to the igraph clustering function. Default: NULL (no additional parameters).

**Value**

A 2-column data frame with the following variables:

**Gene** Gene ID.

**Cluster** Cluster ID as identified by infomap.

**Examples**

```
data(network)
clusters <- cluster_network(network[1:500, ])
```

---

collapse_protein_ids	<i>Collapse protein IDs into gene IDs in sequence names of AAStringSet objects</i>
----------------------	--

---

**Description**

This function can be used if the sequence names of the AAStringSet objects contain protein IDs instead of gene IDs (what syntenet requires)

**Usage**

```
collapse_protein_ids(seq, protein2gene = NULL)
```

**Arguments**

seq	A list of AAStringSet objects, each list element containing protein sequences for a given species. This list must have names (not NULL), and names of each list element must match the names of list elements in <b>protein2gene</b> .
protein2gene	A list of 2-column data frames containing protein-to-gene ID correspondences, where the first column contains protein IDs, and the second column contains gene IDs. Names of list elements must match names of <b>seq</b> .

**Details**

For each species, this function will replace the protein IDs in sequence names with gene IDs using the protein-to-gene correspondence table in **protein2gene**. After replacing protein IDs with gene IDs, if there are multiple sequences with the same gene ID (indicating different isoforms of the same gene), only the longest sequence is kept, so that the number of sequences is not greater than the number of genes.

**Value**

A list of AAStringSet objects as in **seq**, but with protein IDs replaced with gene IDs.

**Examples**

```
# Load data
seq_path <- system.file(
  "extdata", "RefSeq_parsing_example", package = "syntenet"
)
seq <- fasta2AAStringSetlist(seq_path)
annot <- gff2GRangesList(seq_path)

# Clean sequence names
names(seq$Aalosa) <- gsub(" .*", "", names(seq$Aalosa))

# Create a correspondence data frame
cor_df <- as.data.frame(annot$Aalosa[annot$Aalosa$type == "CDS", ])
cor_df <- cor_df[, c("Name", "gene")]

# Create a list of correspondence data frames
protein2gene <- list(Aalosa = cor_df)

# Collapse IDs
new_seqs <- collapse_protein_ids(seq, protein2gene)
```

---

```
create_species_id_table
```

*Create a data frame of species IDs (3-5-character abbreviations)*

---

**Description**

Create a data frame of species IDs (3-5-character abbreviations)

**Usage**

```
create_species_id_table(species_names)
```



**Arguments**

**species\_names** A character vector of names extracted from the **seq** or **annotation** lists, which can be extracted with `names(seq)` or `names(annotation)`.

**Value**

A 2-column data frame with the following variables:

**species\_id** Character, species ID consisting of 3-5 characters.

**species\_name** Character, original names passed as input.

**Examples**

```
# Load 'seq' list (list of AAStringSet objects)
data(proteomes)

# Create ID table
create_species_id_table(names(proteomes))
```

---

`diamond_is_installed` *Check if DIAMOND is installed*

---

**Description**

Check if DIAMOND is installed

**Usage**

```
diamond_is_installed()
```

**Value**

Logical indicating whether DIAMOND is installed or not.

**Examples**

```
diamond_is_installed()
```

---

edges	<i>Synteny network of <i>Ostreococcus</i> genomes represented as an edge list</i>
-------	---

---

**Description**

The object was created by running `infer_syntenet` on the **blast\_list** example data. Code to recreate this data set is available at the `script/` subdirectory.

**Usage**

```
data(edges)
```

**Format**

A data frame containing anchor pairs between two *Ostreococcus* proteomes.

**Examples**

```
data(edges)
```

---

export_sequences	<i>Export processed sequences as FASTA files</i>
------------------	--

---

**Description**

Export processed sequences as FASTA files

**Usage**

```
export_sequences(seq = NULL, outdir = tempdir())
```

**Arguments**

seq	A processed list of <code>AAStringSet</code> objects as returned by <code>process_input()</code> .
outdir	Path to output directory where FASTA files will be stored.

**Value**

Path to exported FASTA files.

**Examples**

```
# Load data
data(proteomes)
data(annotation)

# Process data
pdata <- process_input(proteomes, annotation)

# Export data
outdir <- file.path(tempdir(), "example_test")
export_sequences(pdata$seq, outdir)
```

---

fasta2AAStringSetlist *Read FASTA files in a directory as a list of AAStringSet objects*

---

**Description**

Read FASTA files in a directory as a list of AAStringSet objects

**Usage**

```
fasta2AAStringSetlist(fasta_dir)
```

**Arguments**

fasta\_dir            Character indicating the path to the directory containing FASTA files.

**Value**

A list of AAStringSet objects, where each element represents a different FASTA file.

**Examples**

```
fasta_dir <- system.file("extdata", "sequences", package = "syntenet")
aastringsetlist <- fasta2AAStringSetlist(fasta_dir)
```

---

find\_GS\_clusters            *Find group-specific clusters based on user-defined species classification*

---

**Description**

Find group-specific clusters based on user-defined species classification

**Usage**

```
find_GS_clusters(
  profile_matrix = NULL,
  species_annotation = NULL,
  min_percentage = 50
)
```

**Arguments**

profile\_matrix    A matrix of phylogenomic profiles obtained with phylogenomic\_profile.

species\_annotation

A 2-column data frame with species IDs in the first column (same as column names of profile matrix), and species annotation (e.g., higher-level taxonomic information) in the second column.

**min\_percentage** Numeric scalar with the minimum percentage of species in a group to consider group specificity. For instance, if a given cluster is present in only 1 group of species, but in less than **min\_percentage** of the species for this group, it will not be considered a group-specific cluster. This filtering criterion is useful to differentiate group-specific clusters (e.g., family-specific) from subgroup-specific clusters (e.g., genus-specific). Default: 50.

### Value

A data frame with the following variables:

**Group** To which group of species the cluster is specific.

**Percentage** Percentage of species from the group that are represented by the cluster.

**Cluster** Cluster ID.

### Examples

```
data(clusters)
profile_matrix <- phylogenomic_profile(clusters)

# Species annotation
species_order <- c(
  "vra", "van", "pvu", "gma", "cca", "tpr", "mtr", "adu", "lja",
  "Lang", "car", "pmu", "ppe", "pbr", "mdo", "roc", "fve",
  "Mnot", "Zjuj", "hlu", "jcu", "mes", "rco", "lus", "ptr"
)
species_annotation <- data.frame(
  Species = species_order,
  Family = c(rep("Fabaceae", 11), rep("Rosaceae", 6),
    "Moraceae", "Ramnaceae", "Cannabaceae",
    rep("Euphorbiaceae", 3), "Linaceae", "Salicaceae")
)
gs_clusters <- find_GS_clusters(profile_matrix, species_annotation)
```

---

gff2GRangesList

*Read GFF/GTF files in a directory as a GRangesList object*

---

### Description

Read GFF/GTF files in a directory as a GRangesList object

### Usage

```
gff2GRangesList(gff_dir)
```

### Arguments

**gff\_dir** Character indicating the path to the directory containing GFF/GTF files.

### Value

A GRangesList object, where each element represents a different GFF/GTF file.

**Examples**

```
gff_dir <- system.file("extdata", "annotation", package = "syntenet")
grangeslist <- gff2GRangesList(gff_dir)
```

---

infer\_microsynteny\_phylogeny

*Infer microsynteny-based phylogeny with IQTREE*


---

**Description**

Infer microsynteny-based phylogeny with IQTREE

**Usage**

```
infer_microsynteny_phylogeny(
  transposed_profiles = NULL,
  bootr = 1000,
  alrtboot = 1000,
  threads = "AUTO",
  model = "MK+FO+R",
  outdir = tempdir(),
  outgroup = NULL,
  verbose = FALSE
)
```

**Arguments**

transposed_profiles	A binary and transposed profile matrix. The profile matrix can be obtained with <code>phylogenomic_profile()</code> .
bootr	Numeric scalar with the number of bootstrap replicates. Default: 1000.
alrtboot	Numeric scalar with the number of replicates for the SH-like approximate likelihood ratio test. Default: 1000.
threads	Numeric scalar indicating the number of threads to use or "AUTO", which allows IQTREE to automatically choose the best number of threads to use. Default: "AUTO".
model	Substitution model to use. If you are unsure, pick the default. Default: "MK+FO+R".
outdir	Path to output directory. By default, files are saved in a temporary directory, so they will be deleted when the R session closes. If you want to keep the files, specify a custom output directory.
outgroup	Name of outgroup clade to group the phylogeny. Default: NULL (unrooted phylogeny).
verbose	Logical indicating if progress messages should be prompted. Default: FALSE.

**Value**

A character vector of paths to output files.

**Examples**

```

data(clusters)
profile_matrix <- phylogenomic_profile(clusters)
tmat <- binarize_and_transpose(profile_matrix)

# Leave only some legumes and P. mume as an outgroup for testing purposes
included <- c("gma", "pvu", "vra", "van", "cca", "pmu")
tmat <- tmat[rownames(tmat) %in% included, ]

# Remove non-variable sites
tmat <- tmat[, colSums(tmat) != length(included)]

if(iqtree_is_installed()) {
  phylo <- infer_microsynteny_phylogeny(tmat, outgroup = "pmu",
                                       threads = 1)
}

```

infer\_syntenet

*Infer synteney network***Description**

Infer synteney network

**Usage**

```

infer_syntenet(
  blast_list = NULL,
  annotation = NULL,
  outdir = tempdir(),
  anchors = 5,
  max_gaps = 25,
  is_pairwise = TRUE,
  verbose = FALSE,
  bp_param = BiocParallel::SerialParam(),
  ...
)

```

**Arguments**

blast_list	A list of data frames, each data frame having the tabular output of BLASTp or similar programs, such as DIAMOND. This is the output of the function run_diamond(). If you performed pairwise comparisons on the command line, you can read the tabular output as data frames and combine them in a list. List names must be have species names separated by underscore. For instance, if the first list element is a data frame containing the comparison of speciesA (query) against speciesB (database), its name must be "speciesA_speciesB".
annotation	A processed GRangesList, CompressedGRangesList, or list of GRanges as returned by process_input().
outdir	Path to the output directory. Default: tempdir().

anchors	Numeric indicating the minimum required number of genes to call a syntenic block. Default: 5.
max_gaps	Numeric indicating the number of upstream and downstream genes to search for anchors. Default: 25.
is_pairwise	specify if only pairwise blocks should be reported Default: TRUE
verbose	Logical indicating if log messages should be printed on screen. Default: FALSE.
bp_param	BiocParallel back-end to be used. Default: BiocParallel::SerialParam().
...	Any additional arguments to the MCScanX algorithm. For a complete list of all available options, see the man page of rcpp_mcscanx_file().

### Value

A network represented as an edge list.

### Examples

```
# Load data
data(proteomes)
data(annotation)
data(blast_list)

# Create processed annotation list
annotation <- process_input(proteomes, annotation)$annotation

# Infer the syteny network
net <- infer_sytenet(blast_list, annotation)
```

---

interspecies\_syteny *Detect interspecies syteny*

---

### Description

Detect interspecies syteny

### Usage

```
interspecies_syteny(
  blast_inter = NULL,
  annotation = NULL,
  inter_dir = file.path(tempdir(), "inter"),
  anchors = 5,
  max_gaps = 25,
  is_pairwise = TRUE,
  verbose = FALSE,
  bp_param = BiocParallel::SerialParam(),
  ...
)
```

**Arguments**

blast_inter	A list of BLAST/DIAMOND data frames for interspecies comparisons as returned by run_diamond().
annotation	A processed GRangesList or CompressedGRangesList object as returned by process_input().
inter_dir	Path to output directory where .collinearity files will be stored.
anchors	Numeric indicating the minimum required number of genes to call a syntenic block. Default: 5.
max_gaps	Numeric indicating the number of upstream and downstream genes to search for anchors. Default: 25.
is_pairwise	specify if only pairwise blocks should be reported Default: TRUE.
verbose	Logical indicating if log messages should be printed on screen. Default: FALSE.
bp_param	BiocParallel back-end to be used. Default: BiocParallel::SerialParam().
...	Any additional arguments to the MCScanX algorithm. For a complete list of all available options, see the man page of rcpp_mcscanx_file().

**Value**

Paths to .collinearity files.

**Examples**

```
# Load data
data(proteomes)
data(blast_list)
data(annotation)

# Get DIAMOND and processed annotation lists
blast_inter <- blast_list[2]
annotation <- process_input(proteomes, annotation)$annotation

# Detect interspecies syteny
intersyn <- interspecies_syteny(blast_inter, annotation)
```

---

intraspecies\_syteny *Detect intraspecies syteny*

---

**Description**

Detect intraspecies syteny

**Usage**

```
intraspecies_syteny(
  blast_intra = NULL,
  annotation = NULL,
  intra_dir = file.path(tempdir(), "intra"),
  anchors = 5,
  max_gaps = 25,
```



```

    is_pairwise = TRUE,
    verbose = FALSE,
    bp_param = BiocParallel::SerialParam(),
    ...
  )

```

### Arguments

<code>blast_intra</code>	A list of BLAST/DIAMOND data frames for intraspecies comparisons as returned by <code>run_diamond()</code> .
<code>annotation</code>	A processed <code>GRangesList</code> or <code>CompressedGRangesList</code> object as returned by <code>process_input()</code> .
<code>intra_dir</code>	Path to output directory where <code>.collinearity</code> files will be stored.
<code>anchors</code>	Numeric indicating the minimum required number of genes to call a syntenic block. Default: 5.
<code>max_gaps</code>	Numeric indicating the number of upstream and downstream genes to search for anchors. Default: 25.
<code>is_pairwise</code>	Logical indicating if only pairwise blocks should be reported. Default: TRUE.
<code>verbose</code>	Logical indicating if log messages should be printed on screen. Default: FALSE.
<code>bp_param</code>	<code>BiocParallel</code> back-end to be used. Default: <code>BiocParallel::SerialParam()</code> .
<code>...</code>	Any additional arguments to the <code>MCSanX</code> algorithm. For a complete list of all available options, see the man page of <code>rcpp_mcscanx_file()</code> .

### Value

Paths to `.collinearity` files.

### Examples

```

# Load data
data(scerevisiae_annot)
data(scerevisiae_diamond)

# Detect intragenome synteny
intra_syn <- intraspecies_synteny(
  scerevisiae_diamond, scerevisiae_annot
)

```

---

`iqtree_is_installed`    *Check if IQTREE is installed*

---

### Description

Check if IQTREE is installed

### Usage

```
iqtree_is_installed()
```

**Value**

Logical indicating whether IQTREE is installed or not.

**Examples**

```
iqtree_is_installed()
```

---

<code>iqtree_version</code>	<i>Get IQ-TREE version</i>
-----------------------------	----------------------------

---

**Description**

Get IQ-TREE version

**Usage**

```
iqtree_version()
```

**Value**

Numeric indicating IQ-TREE version, with either 1 or 2.

**Examples**

```
iqtree_version()
```

---

<code>last_is_installed</code>	<i>Check if last is installed</i>
--------------------------------	-----------------------------------

---

**Description**

Check if last is installed

**Usage**

```
last_is_installed()
```

**Value**

Logical indicating whether last is installed or not.

**Examples**

```
last_is_installed()
```

---

network	<i>Syteny network of BUSCO genes for 25 eudicot species</i>
---------	---

---

**Description**

Data obtained from Zhao & Schranz, 2019.

**Usage**

```
data(network)
```

**Format**

An edgelist (i.e., a 2-column data frame with node 1 in column 1 and node 2 in column 2).

**References**

Zhao, T., & Schranz, M. E. (2019). Network-based microsynteny analysis identifies major differences and genomic outliers in mammalian and angiosperm genomes. *Proceedings of the National Academy of Sciences*, 116(6), 2165-2174.

**Examples**

```
data(network)
```

---

parse_collinearity	<i>Parse .collinearity files obtained with MCScan</i>
--------------------	---

---

**Description**

The .collinearity files can be obtained with `intraspecies_syteny` and `interspecies_syteny`, which execute a native version of the MCScan algorithm.

**Usage**

```
parse_collinearity(collinearity_paths = NULL, as = "anchors")
```

**Arguments**

`collinearity_paths`

Character vector of paths to .collinearity files.

`as`

Character specifying what to extract. One of "anchors" (default), "blocks", or "all".

**Value**

If **as** is "anchors", a data frame with variables "Anchor1", and "Anchor2". If **as** is "blocks", a data frame with variables "Block", "Block\_score", "Chr", and "Orientation". If **as** is "all", a data frame with all aforementioned variables, which indicate:

**Block** Numeric, syntenic block ID

**Block\_score** Numeric, score of syntenic block.

**Chr** Character, query and target chromosome of the syntenic block formatted as "&".

**Orientation** Character, the orientation of genes within blocks, with "plus" indicating that genes are in the same direction, and "minus" indicating that genes are in opposite directions.

**Anchor1** Character, gene ID of anchor 1.

**Anchor2** Character, gene ID of anchor 2.

**Examples**

```
collinearity_paths <- system.file(
  "extdata", "Scerevisiae.collinearity", package = "syntenet"
)
net <- parse_collinearity(collinearity_paths)
```

---

phylogenomic\_profile *Perform phylogenomic profiling for syntenic network clusters*

---

**Description**

Perform phylogenomic profiling for syntenic network clusters

**Usage**

```
phylogenomic_profile(clusters = NULL)
```

**Arguments**

**clusters** A 2-column data frame with variables **Gene** and **Cluster** as returned by `cluster_network`.

**Value**

A matrix of *i* rows and *j* columns containing the number of genes in cluster *i* for each species *j*. The number of rows is equal to the number of clusters in **clusters**, and the number of columns is equal to the number of species in **clusters**.

**Examples**

```
data(clusters)
profiles <- phylogenomic_profile(clusters)
```

---

plot_network	<i>Plot network</i>
--------------	---------------------

---

### Description

Plot network

### Usage

```
plot_network(
  network = NULL,
  clusters = NULL,
  cluster_id = NULL,
  color_by = "cluster",
  interactive = FALSE,
  dim_interactive = c(600, 600)
)
```

### Arguments

network	The syntenic network represented as an edge list, which is a 2-column data frame with each member of the anchor pair in a column.
clusters	A 2-column data frame with the variables <b>Gene</b> and <b>Cluster</b> representing gene ID and cluster ID, respectively, exactly as returned by <code>cluster_network</code> .
cluster_id	Character scalar or vector with cluster ID. If more than one cluster is passed as input, clusters are colored differently.
color_by	Either "cluster" or a 2-column data frame with gene IDs in the first column and variable to be used for coloring (e.g., taxonomic information) in the second column.
interactive	Logical scalar indicating whether to display an interactive network or not. Default: FALSE.
dim_interactive	Numeric vector of length 2 with the window dimensions of the interactive plot. If <b>interactive</b> is set to FALSE, this parameter is ignored.

### Value

A ggplot object with the network.

### Examples

```
data(network)
data(clusters)
# Option 1: 1 cluster
cluster_id <- 25
plot_network(network, clusters, cluster_id)

# Option 2: 2 clusters
cluster_id <- c(25, 1089)
plot_network(network, clusters, cluster_id)
```

```

# Option 3: custom annotation for coloring
species_order <- c(
  "vra", "van", "pvu", "gma", "cca", "tpr", "mtr", "adu", "lja",
  "Lang", "car", "pmu", "ppe", "pbr", "mdo", "roc", "fve",
  "Mnot", "Zjuj", "jcu", "mes", "rco", "lus", "ptr"
)
species_annotation <- data.frame(
  Species = species_order,
  Family = c(rep("Fabaceae", 11), rep("Rosaceae", 6),
    "Moraceae", "Ranunculaceae", rep("Euphorbiaceae", 3),
    "Linaceae", "Salicaceae")
)
genes <- unique(c(network$node1, network$node2))
gene_df <- data.frame(
  Gene = genes,
  Species = unlist(lapply(strsplit(genes, "_"), head, 1))
)
gene_df <- merge(gene_df, species_annotation)[, c("Gene", "Family")]

plot_network(network, clusters, cluster_id = 25, color_by = gene_df)

```

---

plot\_profiles

*Plot a heatmap of phylogenomic profiles*


---

## Description

Plot a heatmap of phylogenomic profiles

## Usage

```

plot_profiles(
  profile_matrix = NULL,
  species_annotation = NULL,
  palette = "Greens",
  dist_function = stats::dist,
  dist_params = list(method = "euclidean"),
  clust_function = stats::hclust,
  clust_params = list(method = "ward.D"),
  cluster_species = FALSE,
  show_colnames = FALSE,
  discretize = TRUE,
  ...
)

```

## Arguments

**profile\_matrix** A matrix of phylogenomic profiles obtained with `phylogenomic_profile`.

**species\_annotation**

A 2-column data frame with species IDs in the first column (same as column names of profile matrix), and species annotation (e.g., higher-level taxonomic information) in the second column.

**palette**

A character vector of colors or a character scalar with the name of an RColorBrewer palette. Default: "RdYIBu".

<code>dist_function</code>	Function to use to calculate a distance matrix for synteny clusters. Popular examples include <code>stats::dist</code> , <code>labdsv::dsvdis</code> , and <code>vegan::vegdist</code> . Default: <code>stats::dist</code> .
<code>dist_params</code>	A list with parameters to be passed to the function specified in parameter <b><code>dist_function</code></b> . Default: <code>list(method = "euclidean")</code> .
<code>clust_function</code>	Function to use to cluster the distance matrix returned by the function specified in <code>dist_function</code> . Examples include <code>stats::hclust</code> and <code>Rclusterpp::Rclusterpp.hclust</code> . Default: <code>stats::hclust</code> .
<code>clust_params</code>	A list with additional parameters (if any) to be passed to the function specified in parameter <b><code>clust_function</code></b> . Default: <code>list(method = "ward.D")</code> .
<code>cluster_species</code>	Either a logical scalar (TRUE or FALSE) or a character vector with the order in which species should be arranged. TRUE or FALSE indicate whether hierarchical clustering should be applied to rows (species). Ideally, the character vector should contain the order of species in a phylogenetically meaningful way. If users pass a named vector, vector names will be used to rename species. If users have a species tree, they can read it with <code>treeio::read.tree()</code> , plot it with <code>ggtree::ggtree()</code> , and get the species order from the <code>ggtree</code> object with <code>ggtree::get_taxa_name()</code> . Default: FALSE.
<code>show_colnames</code>	Logical indicating whether to show column names (i.e., cluster IDs) or not. Showing cluster IDs can be useful when visualizing a small subset of them. When visualizing all clusters, cluster IDs are impossible to read. Default: FALSE.
<code>discretize</code>	Logical indicating whether to discretize clusters in 4 categories: 0, 1, 2, and 3+. If FALSE, counts will be log2 transformed. Default: TRUE.
...	Additional parameters to <code>pheatmap::pheatmap()</code> .

**Value**

A pheatmap object.

**Examples**

```
data(clusters)
profile_matrix <- phylogenomic_profile(clusters)
species_order <- c(
  "vra", "van", "pvu", "gma", "cca", "tpr", "mtr", "adu", "lja",
  "Lang", "car", "pmu", "ppe", "pbr", "mdo", "roc", "fve",
  "Mnot", "Zjuj", "jcu", "mes", "rco", "lus", "ptr"
)
species_names <- c(
  "V. radiata", "V. angularis", "P. vulgaris", "G. max", "C. cajan",
  "T. pratense", "M. truncatula", "A. duranensis", "L. japonicus",
  "L. angustifolius", "C. arietinum", "P. mume", "P. persica",
  "P. bretschneideri", "M. domestica", "R. occidentalis",
  "F. vesca", "M. notabilis", "Z. jujuba", "J. curcas",
  "M. esculenta", "R. communis", "L. usitatissimum", "P. trichocarpa"
)
names(species_order) <- species_names
species_annotation <- data.frame(
  Species = species_order,
  Family = c(rep("Fabaceae", 11), rep("Rosaceae", 6),
             "Moraceae", "Ramnaceae", rep("Euphorbiaceae", 3),
             "Linaceae", "Salicaceae")
)
```

```

)
p <- plot_profiles(profile_matrix, species_annotation,
                  cluster_species = species_order)

p <- plot_profiles(profile_matrix, species_annotation,
                  cluster_species = species_order,
                  discretize = FALSE)

```

---

process\_input

*Process sequence data*

---

## Description

Process sequence data

## Usage

```

process_input(
  seq = NULL,
  annotation = NULL,
  gene_field = "gene_id",
  filter_annotation = FALSE
)

```

## Arguments

seq	A list of AAStringSet objects, each list element containing protein sequences for a given species. This list must have names (not NULL), and names of each list element must match the names of list elements in <b>annotation</b> .
annotation	A GRangesList, CompressedGRangesList, or list of GRanges with the annotation for the sequences in <b>seq</b> . This list must have names (not NULL), and names of each list element must match the names of list elements in <b>seq</b> .
gene_field	Character, name of the column in the GRanges objects that contains gene IDs. Default: "gene_id".
filter_annotation	Logical indicating whether <b>annotation</b> should be filtered to keep only genes that are also in <b>seq</b> . This is particularly useful if users want to remove information on non-protein coding genes from <b>annotation</b> , since such genes are typically not present in sets of whole-genome protein sequences. Default: FALSE.

## Details

This function processes the input sequences and annotation to:

1. Remove whitespace and anything after it in sequence names (i.e., names(seq[[x]]), which is equivalent to FASTA headers), if there is any.
2. Add a unique species identifier to sequence names. The species identifier consists of the first 3-5 strings of the element name. For instance, if the first element of the **seq** list is named "Athaliana", each sequence in it will have an identifier "Atha\_" added to the beginning of each gene name (e.g., Atha\_AT1G01010).
3. If sequences have an asterisk (\*) representing stop codon, remove it.



4. Add a unique species identifier (same as above) to gene and chromosome names of each element of the **annotation** GRangesList/CompressedGRangesList.
5. Filter each element of the **annotation** GRangesList/CompressedGRangesList to keep only seqnames, ranges, and gene ID.

### Value

A list of 2 elements:

**seq** The processed list of AAStringSet objects from **seq**.

**annotation** The processed GRangesList or CompressedGRangesList object from **annotation**.

### Examples

```
data(annotation)
data(proteomes)
seq <- proteomes
clean_data <- process_input(seq, annotation)
```

---

profiles2phylip	<i>Save the transposed binary profiles matrix to a file in PHYLIP format</i>
-----------------	--

---

### Description

Save the transposed binary profiles matrix to a file in PHYLIP format

### Usage

```
profiles2phylip(transposed_profiles = NULL, outdir = tempdir())
```

### Arguments

transposed_profiles	A binary and transposed profile matrix. The profile matrix can be obtained with <code>phylogenomic_profile()</code> .
outdir	Path to output directory. By default, files are saved in a temporary directory, so they will be deleted when the R session closes. If you want to keep the files, specify a custom output directory.

### Value

Character specifying the path to the PHYLIP file.

### Examples

```
data(clusters)
profile_matrix <- phylogenomic_profile(clusters)
tmat <- binarize_and_transpose(profile_matrix)
profiles2phylip(tmat)
```

---

 proteomes

*Filtered proteomes of Ostreococcus sp. species*


---

### Description

Data obtained from Pico-PLAZA 3.0. Only the translated sequences of primary transcripts were included, and only genes from chromosomes 1, 2, and 3.

### Usage

```
data(proteomes)
```

### Format

A list of AAStringSet objects containing the elements *Olucimarinus*, *Osp\_RCC809*, and *Otauri*.

### References

Van Bel, M., Silvestri, F., Weitz, E. M., Kreft, L., Botzki, A., Coppens, F., & Vandepoele, K. (2021). PLAZA 5.0: extending the scope and power of comparative and functional genomics in plants. *Nucleic acids research*.

### Examples

```
data(proteomes)
```

---

 rcpp\_mcscanx\_file

*rcpp\_mcscanx\_file*


---

### Description

MCSCanX provides a clustering module for viewing the relationship of collinear segments in multiple genomes (or heavily redundant genomes). It takes the predicted pairwise segments from dynamic programming (DAGchainer in particular) and then tries to build consensus segments from a set of related, overlapping segments.

### Usage

```
rcpp_mcscanx_file(
  blast_file,
  gff_file,
  prefix = "out",
  outdir = "",
  match_score = 50L,
  gap_penalty = -1L,
  match_size = 5L,
  e_value = 1e-05,
  max_gaps = 25L,
  overlap_window = 5L,
  is_pairwise = FALSE,
```

```

    in_synteny = 0L,
    species_id_length = 3L,
    verbose = FALSE
)

```

### Arguments

<code>blast_file</code>	Character indicating the path to the BLAST/DIAMOND output file.
<code>gff_file</code>	Character indicating the path to the "gff" file, which is a tab-delimited file with 4 columns indicating the chromosome name, gene id, gene start position, and gene end position, respectively.
<code>prefix</code>	Character indicating the prefix to output files. Default: "out".
<code>outdir</code>	Character indicating the path to the output directory. Default: "".
<code>match_score</code>	Numeric indicating the match score. Default: 50.
<code>gap_penalty</code>	Numeric indicating the gap penalty. Default: -1.
<code>match_size</code>	Numeric indicating the minimum number of genes required to call synteny. Default: 5.
<code>e_value</code>	Numeric indicating the minimum e-value allowed. Default: 1e-5.
<code>max_gaps</code>	Numeric indicating the maximum number of gaps between genes allowed. The unit measure of gaps is number of genes, so <code>max_gaps = 20</code> indicates that a maximum of 20 genes can exist between two homologous genes for synteny to be called. Default: 25.
<code>overlap_window</code>	Numeric indicating the overlap window. Default: 5.
<code>is_pairwise</code>	Logical indicating whether only pairwise blocks should be reported. Default: FALSE.
<code>in_synteny</code>	Numeric indicating the patterns of collinear blocks, where 0 indicates intra and interspecies comparisons, 1 indicates intraspecies comparisons, and 2 indicates interspecies comparisons. Default: 0.
<code>species_id_length</code>	Integer indicating the length of the species IDs. Default: 3. 0: intra- and inter-species (default); 1: intra-species; 2: inter-species
<code>verbose</code>	Logical indicating whether to print progress messages to the screen. Default: FALSE.

### Value

NULL, and a .collinearity file is created in the directory specified in `outdir`.

### Author(s)

Kristian K Ullrich and Fabricio Almeida-Silva

### References

- Wang et al. (2012) MCScanX: a toolkit for detection and evolutionary analysis of gene synteny and collinearity. *Nucleic acids research*. **40.7**, e49-e49.
- Haas et al. (2004) DAGchainer: a tool for mining segmental genome duplications and synteny. *Bioinformatics*. **20.18** 3643-3646.

---

read_diamond	<i>Read DIAMOND/BLAST tables as a list of data frames</i>
--------------	---

---

**Description**

Read DIAMOND/BLAST tables as a list of data frames

**Usage**

```
read_diamond(diamond_dir = NULL)
```

**Arguments**

`diamond_dir` Path to directory containing the tabular output of DIAMOND or similar programs (e.g., BLAST).

**Value**

A list of data frames with the tabular DIAMOND output.

**Examples**

```
# Path to output directory
diamond_dir <- system.file("extdata", package = "syntenet")

# Read output
l <- read_diamond(diamond_dir)
```

---

run_diamond	<i>Wrapper to run DIAMOND from an R session</i>
-------------	---

---

**Description**

Wrapper to run DIAMOND from an R session

**Usage**

```
run_diamond(
  seq = NULL,
  top_hits = 5,
  verbose = FALSE,
  outdir = tempdir(),
  threads = NULL,
  compare = "all",
  ...
)
```

**Arguments**

seq	A processed list of AAStringSet objects as returned by process_input().
top_hits	Number of top hits to keep in DIAMOND search. Default: 5.
verbose	Logical indicating if progress messages should be printed. Default: FALSE.
outdir	Output directory for DIAMOND results. By default, output files are saved to a temporary directory.
threads	Number of threads to use. Default: let DIAMOND auto-detect and use all available virtual cores on the machine.
compare	Character scalar indicating which comparisons should be made when running DIAMOND. Possible modes are "all" (all-vs-all comparisons), "intraspecies" (intraspecies comparisons only), or "interspecies" (interspecies comparisons only). Alternatively, users can pass a 2-column data frame as input with the names of species to be compared.
...	Any additional arguments to diamond blastp.

**Value**

A list of data frames containing DIAMOND's tabular output for each pairwise combination of species. For  $n$  species, the list length will be  $n^2$ .

**Examples**

```
data(proteomes)
data(annotation)
seq <- process_input(proteomes, annotation)$seq[1:2]
if(diamond_is_installed()) {
  diamond_results <- run_diamond(seq)
}
```

---

run\_last

*Wrapper to run last from an R session*


---

**Description**

Wrapper to run last from an R session

**Usage**

```
run_last(
  seq = NULL,
  verbose = FALSE,
  outdir = tempdir(),
  threads = 1,
  compare = "all",
  lastD = 1e+06,
  ...
)
```

**Arguments**

seq	A processed list of AAStringSet objects as returned by process_input().
verbose	Logical indicating if progress messages should be printed. Default: FALSE.
outdir	Output directory for last results. By default, output files are saved to a temporary directory.
threads	Number of threads to use. Default: 1.
compare	Character scalar indicating which comparisons should be made when running last. Possible modes are "all" (all-vs-all comparisons), "intraspecies" (intraspecies comparisons only), or "interspecies" (interspecies comparisons only). Alternatively, users can pass a 2-column data frame as input with the names of species to be compared.
lastD	last option D: query letters per random alignment. Default: 1e6.
...	Any additional arguments to lastal.

**Value**

A list of data frames containing last's tabular output for each pairwise combination of species. For  $n$  species, the list length will be  $n^2$ .

**Examples**

```
data(proteomes)
data(annotation)
seq <- process_input(proteomes, annotation)$seq[1:2]
if(last_is_installed()) {
  last_results <- run_last(seq)
}
```

---

scerevisiae\_annot      *Genome annotation of the yeast species S. cerevisiae*

---

**Description**

Data obtained from Ensembl Fungi. Only annotation data for primary transcripts were included.

**Usage**

```
data(scerevisiae_annot)
```

**Format**

A GRangesList as returned by process\_input() containing the element **Scerevisiae**.

**Examples**

```
data(scerevisiae_annot)
```

---

scerevisiae\_diamond    *Intraspecies DIAMOND output for S. cerevisiae*

---

**Description**

List obtained with `run_diamond()`.

**Usage**

```
data(scerevisiae_diamond)
```

**Format**

A list of data frames (length 1) containing the whole paranome of *S. cerevisiae* resulting from intragenome similarity searches.

**Examples**

```
data(scerevisiae_diamond)
```

# Index

## \* datasets

- angiosperm\_phylogeny, 3
- annotation, 4
- blast\_list, 5
- clusters, 6
- edges, 10
- network, 19
- proteomes, 26
- scerevisiae\_annot, 30
- scerevisiae\_diamond, 31

## \* internal

- syntenet-package, 3

- angiosperm\_phylogeny, 3
- annotation, 4

- binarize\_and\_transpose, 4
- blast\_list, 5

- check\_input, 5
- cluster\_network, 7
- clusters, 6
- collapse\_protein\_ids, 7
- create\_species\_id\_table, 8

- diamond\_is\_installed, 9

- edges, 10
- export\_sequences, 10

- fasta2AAStringSetlist, 11
- find\_GS\_clusters, 11

- gff2GRangesList, 12

- infer\_microsynteny\_phylogeny, 13
- infer\_syntenet, 14
- interspecies\_synteny, 15
- intraspecies\_synteny, 16
- iqtree\_is\_installed, 17
- iqtree\_version, 18

- last\_is\_installed, 18

- network, 19

- parse\_collinearity, 19
- phylogenomic\_profile, 20
- plot\_network, 21
- plot\_profiles, 22
- process\_input, 24
- profiles2phylip, 25
- proteomes, 26

- rcpp\_mcscanx\_file, 26
- read\_diamond, 28
- run\_diamond, 28
- run\_last, 29

- scerevisiae\_annot, 30
- scerevisiae\_diamond, 31
- syntenet (syntenet-package), 3
- syntenet-package, 3