

# Package ‘ChIPanalyser’

September 18, 2024

**Type** Package

**Title** ChIPanalyser: Predicting Transcription Factor Binding Sites

**Version** 1.27.0

**Date** 2017-09-01

**Author** Patrick C.N.Martin & Nicolae Radu Zabet

**Maintainer** Patrick C.N. Martin <pcnmartin@gmail.com>

**Citation** Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Description** ChIPanalyser is a package to predict and understand TF binding by utilizing a statistical thermodynamic model. The model incorporates 4 main factors thought to drive TF binding: Chromatin State, Binding energy, Number of bound molecules and a scaling factor modulating TF binding affinity. Taken together, ChIPanalyser produces ChIP-like profiles that closely mimic the patterns seen in real ChIP-seq data.

**License** GPL-3

**Collate** 2AllS4Class\_ProfileParameters.R 3AllGenerics.R 4AllMethods.R AllInitialize.R AllShowMethods.R computeChIPProfile.R computeOccupancy.R computeOptimal.R computePWMScore.R computeGenomeWide.R parallelInternalFunctionsDev.R GenomicProfileGenericFunctions.R plotOccupancy.R plotOptimalHeatMapDev.R DataPreprocessingDev.R DataPreprocessingGenericFunctionsDev.R profileAccuracyEstimateDev.R GAAnalysis.R GAGeneric.R

**Depends** R (>= 3.5.0), GenomicRanges, Biostrings, BSgenome, RcppRoll, parallel

**Imports** methods, IRanges, S4Vectors, grDevices, graphics, stats, utils, rtracklayer, ROCR, BiocManager, GenomeInfoDb, RColorBrewer

**Suggests** BSgenome.Dmelandogaster, UCSC.dm6, knitr, RUnit, BiocGenerics

**Encoding** UTF-8

**LazyData** true

**biocViews** Software, BiologicalQuestion, WorkflowStep, Transcription, Sequencing, ChipOnChip, Coverage, Alignment, ChIPSeq, SequenceMatching, DataImport, PeakDetection

**VignetteBuilder** knitr  
**RoxygenNote** 7.2.1  
**git\_url** <https://git.bioconductor.org/packages/ChIPAnalyser>  
**git\_branch** devel  
**git\_last\_commit** 6ba9b03  
**git\_last\_commit\_date** 2024-04-30  
**Repository** Bioconductor 3.20  
**Date/Publication** 2024-09-18

## Contents

ChIPAnalyser-package	5
averageExpPWMScore	6
averageExpPWMScore-methods	7
backgroundSignal	7
backgroundSignal-methods	8
backgroundSignal<-	9
backgroundSignal<-methods	10
boundMolecules	10
boundMolecules-methods	11
boundMolecules<-	11
boundMolecules<-methods	12
BPFrequency	12
BPFrequency-methods	13
BPFrequency<-	14
BPFrequency<-methods	15
ChIPAnalyserData	15
chipMean	16
chipMean-methods	17
chipMean<-	17
chipMean<-methods	18
ChIPScore-class	19
chipSd	20
chipSd-methods	21
chipSd<-	22
chipSd<-methods	23
chipSmooth	23
chipSmooth-methods	24
chipSmooth<-	24
chipSmooth<-methods	25
computeChIPProfile	25
computeGenomeWideScores	27
computeOccupancy	29
computeOptimal	30
computePWMScore	32
DNASequenceLength	34
DNASequenceLength-methods	35
drop	35
drop-methods	36

evolve . . . . .	37
generateStartingPopulation . . . . .	38
genomicProfiles . . . . .	39
genomicProfiles-class . . . . .	41
genomicProfilesInternal-class . . . . .	43
getHighestFitnessSolutions . . . . .	45
getTestingData . . . . .	46
getTrainingData . . . . .	46
GRList-class . . . . .	47
initialize-methods . . . . .	48
lambdaPWM . . . . .	48
lambdaPWM-methods . . . . .	49
lambdaPWM<- . . . . .	49
lambdaPWM<-methods . . . . .	50
loci . . . . .	50
loci-class . . . . .	51
loci-methods . . . . .	52
lociWidth . . . . .	52
lociWidth-methods . . . . .	53
lociWidth<- . . . . .	53
lociWidth<-methods . . . . .	54
maxPWMScore . . . . .	54
maxPWMScore-methods . . . . .	55
maxSignal . . . . .	56
maxSignal-methods . . . . .	57
maxSignal<- . . . . .	57
maxSignal<-methods . . . . .	58
minPWMScore . . . . .	58
minPWMScore-methods . . . . .	59
naturalLog . . . . .	59
naturalLog-methods . . . . .	60
naturalLog<- . . . . .	61
naturalLog<-methods . . . . .	62
noiseFilter . . . . .	62
noiseFilter-methods . . . . .	63
noiseFilter<- . . . . .	63
noiseFilter<-methods . . . . .	64
noOfSites . . . . .	64
noOfSites-methods . . . . .	65
noOfSites<- . . . . .	65
noOfSites<-methods . . . . .	66
nos-class . . . . .	67
parameterOptions . . . . .	67
parameterOptions-class . . . . .	69
PFMFormat . . . . .	72
PFMFormat-methods . . . . .	73
PFMFormat<- . . . . .	73
PFMFormat<-methods . . . . .	74
ploidy . . . . .	74
ploidy-methods . . . . .	75
ploidy<- . . . . .	75
ploidy<-methods . . . . .	76

plotOccupancyProfile . . . . .	76
plotOptimalHeatMaps . . . . .	78
PositionFrequencyMatrix . . . . .	80
PositionFrequencyMatrix-methods . . . . .	81
PositionFrequencyMatrix<- . . . . .	81
PositionFrequencyMatrix<-methods . . . . .	82
PositionWeightMatrix . . . . .	83
PositionWeightMatrix-methods . . . . .	84
PositionWeightMatrix<- . . . . .	84
PositionWeightMatrix<-methods . . . . .	85
processingChIP . . . . .	85
profileAccuracyEstimate . . . . .	86
profiles-methods . . . . .	88
PWMPseudocount . . . . .	88
PWMPseudocount-methods . . . . .	89
PWMPseudocount<- . . . . .	90
PWMPseudocount<-methods . . . . .	91
PWMThreshold . . . . .	91
PWMThreshold-methods . . . . .	92
PWMThreshold<- . . . . .	92
PWMThreshold<-methods . . . . .	93
removeBackground . . . . .	93
removeBackground-methods . . . . .	94
removeBackground<- . . . . .	94
removeBackground<-methods . . . . .	95
scores . . . . .	96
scores-methods . . . . .	96
searchSites . . . . .	97
setChromatinStates . . . . .	98
show-methods . . . . .	99
singleRun . . . . .	100
splitData . . . . .	101
stepSize . . . . .	102
stepSize-methods . . . . .	103
stepSize<- . . . . .	103
stepSize<-methods . . . . .	104
strandRule . . . . .	104
strandRule-methods . . . . .	105
strandRule<- . . . . .	105
strandRule<-methods . . . . .	106
whichstrand . . . . .	106
whichstrand-methods . . . . .	107
whichstrand<- . . . . .	108
whichstrand<-methods . . . . .	109

---

ChIPAnalyser-package    *ChIPAnalyser: Predicting Transcription Factor Binding Sites*

---

## Description

ChIPAnalyser is a package to predict and understand TF binding by utilizing a statistical thermodynamic model. The model incorporates 4 main factors thought to drive TF binding: Chromatin State, Binding energy, Number of bound molecules and a scaling factor modulating TF binding affinity. Taken together, ChIPAnalyser produces ChIP-like profiles that closely mimic the patterns seen in real ChIP-seq data.

## Details

The DESCRIPTION file: This package was not yet installed at build time.

Index: This package was not yet installed at build time.

## Author(s)

Patrick C.N. Martin <pm16057@essex.ac.uk>

And

Nicolae Radu Zabet <nzabet@essex.ac.uk>

Maintainer: Patrick C.N. Martin <pcnmartin@gmail.com>

## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

## Examples

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR",BPFfrequency=DNASequenceSet)

chip<-processingChIP(chip,top)
# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(DNASequenceSet = DNASequenceSet,
  genomicsProfiles = GPP)
```

```
#Compute PWM Scores
PWMScores <- computePWMscore(genomicsProfiles = GenomeWide,
  DNASequenceSet = DNASequenceSet,
  loci = top, chromatinState = Access)
#Compute Occupancy
Occupancy <- computeOccupancy(genomicsProfiles = PWMScores,
  parameterOptions = OPP)

#Compute CHIP profiles
chipProfile <- computeChIPProfile(genomicProfiles = Occupancy,
  loci = top,
  parameterOptions = OPP)
#Estimating accuracy estimate
AccuracyEstimate <- profileAccuracyEstimate(genomicProfiles = chipProfile,
  ChIPScore = chip,
  parameterOptions = OPP)
```

---

averageExpPWMScore     *Accessor for averageExpPWMscore slot in a [genomicProfiles](#) object.*

---

## Description

Extract or Access averageExpPWMscore slot in a [genomicProfiles](#)

## Usage

```
averageExpPWMscore(object)
```

## Arguments

object                object is a [genomicProfiles](#)

## Details

As a general rule, averageExpPWMscore is computed and updated internally by [computeGenomeWideScores](#). Ideally, this slot should not be updated by user. The averageExpPWMscore is the sum of the exponential of every PWM score for a given DNA sequence and divided by the length of the said DNA sequence ([DNASequenceLength](#)). This can either be the full length sequence or only the accessible sequence (see [computeGenomeWideScores](#)).

## Value

Returns the averageExpPWMscore of a [genomicProfiles](#) when computed.

## Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Accessing Data
data(ChIPanalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata", package="ChIPanalyser"), "BEAF-32.pfm")
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM, PFMFormat="JASPAR")
# Extracting AllSitesAboveThreshold slot
averageExpPWMScore(GPP)

## Note this slot is now empty as nothing has yet been computed
```

---

averageExpPWMScore-methods

~~ *Methods for Function averageExpPWMScore* ~~

---

**Description**

~~ Methods for function averageExpPWMScore ~~

**Methods:**

signature(object = "genomicProfilesInternal")

---

backgroundSignal      *Accessor method for the backgroundSignal slot in a parameterOptions object.*

---

**Description**

Extract or access the backgroundSignal slot in a [parameterOptions](#) object.

**Usage**

```
backgroundSignal(object)
```

**Arguments**

object                  object is an [parameterOptions](#)

## Details

Default Value: 0

When computing `computeOccupancy`, a ChIP-seq background signal is used to scale Occupancy by considering both a `backgroundSignal` and a `maxSignal`. The `backgroundSignal` is also used to normalise occupancies against `maxOccupancy`. The `backgroundSignal` usually comes from experimental data and is provided by user. As a general rule, if ChIP-seq data is available and will be used in `computeChIPProfile`, `profileAccuracyEstimate` or `plotOccupancyProfile`, it is advised to use the `backgroundSignal` from this data. We strongly encourage to set values when building a `parameterOptions` object.

## Value

Returns a `backgroundSignal` of a `parameterOptions` object.

## Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

## Examples

```
# Building occupancyProfileParameters object
OPP <- parameterOptions()
#Viewing single value in object
backgroundSignal(OPP)
```

---

backgroundSignal-methods

~~ *Methods for Function backgroundSignal* ~~

---

## Description

~~ Methods for function backgroundSignal ~~

## Methods:

signature(object = "parameterOptions")



---

backgroundSignal<-      *Setter method for backgroundSignal slot in a [parameterOptions](#)*

---

### Description

Setter method for backgroundSignal slot in a [parameterOptions](#)

### Usage

```
backgroundSignal(object)<-value
```

### Arguments

object	object is an <a href="#">parameterOptions</a> object.
value	value is the value to be assigned to the backgroundSignal slot in <a href="#">parameterOptions</a> . backgroundSignal should be a positive value. Default value is 0.

### Details

Default value: 0. When computing [computeOccupancy](#), a ChIP-seq background signal is used to scale Occupancy by considering both a backgroundSignal and a [maxSignal](#). The backgroundSignal is also used to normalise occupancies to maxOccupancy. The backgroundSignal usually comes from experimental data and is provided by user. As a general rule, if ChIP-seq data is available and will be used in [computeChIPProfile](#), [profileAccuracyEstimate](#) or [plotOccupancyProfile](#), it is advised to use the backgroundSignal from this data. We strongly encourage to set values when building a [parameterOptions](#) object.

### Value

Returns a [parameterOptions](#) object with a new value assigned to the backgroundSignal slot.

### Author(s)

Patrick C.N. Martin <[pcnmartin@gmail.com](mailto:pcnmartin@gmail.com)>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

### Examples

```
# Building occupancyProfileParameters object
OPP <- parameterOptions()
# Setting new value for backgroundSignal
backgroundSignal(OPP) <- 0.2
# Viewing whole object with new updated value
OPP
#Viewing single value in object
backgroundSignal(OPP)
```

---

```
backgroundSignal<-methods
  ~~ Methods for Function backgroundSignal<- ~~
```

---

**Description**

~~ Methods for function backgroundSignal<- ~~

**Methods:**

```
backgroundSignal(object)<-value
```

---

```
boundMolecules      Accessor methods for boundMolecules slot in parameterOptions
                      object.
```

---

**Description**

Extract or Access boundMolecules slot in [parameterOptions](#) object.

**Usage**

```
boundMolecules(object)
```

**Arguments**

object            object is a [parameterOptions](#) object.

**Details**

Default value: 1000

When computing occupancy ([computeOccupancy](#)), a value for the number of bound Molecules to DNA is needed. This value can be updated and set in a [parameterOptions](#) object. If the number of molecules is unknown, it is possible to infer this value with [computeOptimal](#). We strongly encourage to set values when building a [parameterOptions](#) object.

**Value**

Returns boundMolecules slot in [parameterOptions](#) object.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Building parameterOptions object
OPP <- parameterOptions()
#Checking single value by slot accessor
boundMolecules(OPP)
```

---

boundMolecules-methods

~~ *Methods for Function boundMolecules* ~~

---

**Description**

~~ Methods for function boundMolecules ~~

**Methods:**

signature(object = "parameterOptions")

---

boundMolecules<-      *Setter method for the boundMolecules slot in a [parameterOptions](#) object.*

---

**Description**

Setter method for the boundMolecules slot in a [parameterOptions](#) object.

**Usage**

```
boundMolecules(object)<-value
```

**Arguments**

object	object is a <a href="#">parameterOptions</a> object.
value	value is a positive integer or vector of positive integers describing the number of molecules bound to DNA. Default value is 1000.

**Details**

Default value: 1000 When computing occupancy ([computeOccupancy](#)), a value for the number of bound Molecules to DNA is needed. This value can be updated and set in a [parameterOptions](#) object. If the number of molecules is unknown, it is possible to infer this value with [computeOptimal](#). We strongly encourage to set values when building a [parameterOptions](#) object.

**Value**

Returns a [parameterOptions](#) object with an updated value for boundMolecules.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
# Building parameterOptions object
OPP <- parameterOptions()
# Setting new boundMolecules value in OPP
boundMolecules(OPP) <- 5000
#Checking value in whole object
OPP
#Checking single value by slot accessor
boundMolecules(OPP)
```

---

boundMolecules<-methods

~~ *Methods for Function* boundMolecules<- ~~

---

**Description**

~~ Methods for function boundMolecules<- ~~

**Methods:**

signature(object = "parameterOptions", value = "vector")

---

BPFrequency

*Accessor method for BPFrequency slot in a [genomicProfiles](#) object.*

---

**Description**

Extract or Access BPFrequency slot in a [genomicProfiles](#) object.

**Usage**

BPFrequency(object)

**Arguments**

object                    object is a [genomicProfiles](#)

## Details

Default value is `c(0.25,0.25,0.25,0.25)` When generating a Position Weight Matrix from a Position Frequency Matrix, the probability of occurrence of each base pair (Base Pair Frequency) is necessary (as originally described by Gary Stormo). It is possible to set custom values for BPFrequency with a vector of length 4 containing the probability of occurrence of each base pair (A,C,G,T) in order. If Base pair frequency is unknown, BPFrequency will compute base pair frequency from a DNA sequence. The nature of this sequence can be a BSgenome or a DNASTringSet. In order to decrease run time, it is advised to use DNASTringSet

## Value

Returns BPFrequency slot in `genomicProfiles` object.

## Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

## Examples

```
data(ChIPanalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata", package="ChIPanalyser"), "BEAF-32.pfm")
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM, PFMFormat="JASPAR")
#Extracting BPFrequency slot
BPFrequency(GPP)
```

---

BPFrequency-methods     *~~ Methods for Function BPFrequency ~~*

---

## Description

*~~ Methods for function BPFrequency ~~*

## Methods:

`signature(object = "genomicProfilesInternal")`

---

BPFfrequency<-                      *Setter method for BPFfrequency slot in a [genomicProfiles](#) object.*

---

### Description

Setter method for BPFfrequency slot in a [genomicProfiles](#) object. If base pair frequency is unknown, BPFfrequency will compute base pair frequency from a DNA sequence.

### Usage

```
BPFfrequency(object)<-value
```

### Arguments

object	object is a <a href="#">genomicProfiles</a> object.
value	value can three different objects: A vector of length 4 containing the probability of occurrence of each base pair (A,C,G,T) in order. Default value is <code>c(0.25,0.25,0.25,0.25)</code> . A BSgenome of the organism of interest. The base pair frequency will automatically be computed and updated in <a href="#">genomicProfiles</a> . A <a href="#">DNAStringSet</a> of the organisme of interest. The base pair frequency will automatically be computed and updated in <a href="#">genomicProfiles</a> (Prefered method).

### Details

Default value is `c(0.25,0.25,0.25,0.25)` When generating a Postion Weight Matrix from a Position Frequency Matrix, the probability of occurrence of each base pair (Base Pair Frequency) is necessary (as originally described by Gary Stormo). It is possible to set custom values for BPFfrequency with a vector of length 4 containing the probability of occurrence of each base pair (A,C,G,T) in order. If Base pair frequency is unknown, BPFfrequency will compute base pair frequency from a DNA sequence when building a [genomicProfiles](#) object. The nature of this sequence can be aBSgenome object or a [DNAStringSet](#). In order to decrease run time, it is advised to use [DNAStringSet](#).

### Value

Returns a [genomicProfiles](#) object with an updated value for BPFfrequency.

### Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```

data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM,PFMFormat = "JASPAR", BPFrequency=DNASequenceSet)
# Updating BPFrequency
## !! Note!! BPFrequency is used to compute PWM from PFM
## IF updated after building GPP, then it will not influence PWM
## Advised to build with BPFrequency directly
BPFrequency(GPP) <- DNASequenceSet
BPFrequency(GPP) <- c(0.25,0.25,0.25,0.25)

```

---

BPFrequency<-methods    ~- Methods for Function BPFrequency<- ~-

---

**Description**

~- Methods for function BPFrequency<- ~-

**Methods:**

```

signature(object = "genomicProfilesInternal", value = "DNAStrngSet")
signature(object = "genomicProfilesInternal", value = "vector")

```

---

ChIPAnalyserData            *ChIPAnalyserData*

---

**Description**

ChIPAnalyserData is derived from real biological data. The source organism is *Drosophila melanogaster*. The data can be described as genomic data as it contains DNA sequences, loci, genetic information, DNA accessibility data and ChIP-seq data.

**Usage**

```
data(ChIPAnalyserData)
```

**Format**

1. Accessis [GRanges](#) containing DNA Accesibility data for the sequences described above.
2. csis [GRanges](#) containing Chromatin State data for the sequences described above.
3. topis [GRanges](#) containing a locus of interest. In this case *eve strip Locus* on chromosome 2R in *Drosophila melanogaster*
4. chipis a [GRanges](#) containing ChIP score of the eve strip locus in *Drosophila melanogaster*.
5. geneRefis a [GRanges](#) containing UCSC gene reference information

**Value**

Returns a set of Rdata objects as described above.

**Source**

Transcription Factor PFM: Berkeley Drosophila Transcription Network Project (bdtncp.lbl.gov)

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
data(ChIPanalyserData)
```

---

chipMean

*Accessor method for chipMean slot in a parameterOptions object.*

---

**Description**

Accessor method for [chipMean](#) slot in a [parameterOptions](#) object.

**Usage**

```
chipMean(object)
```

**Arguments**

object            object is a [parameterOptions](#)

**Details**

Default vlaue : 150 When computing ChIP-seq like profiles ([computeChIPProfile](#), the occupancy values given by computeOccupancy are transformed into ChIP-seq like profiles. The average size of a ChIP-seq peak was described by Kaplan (Kaplan et al. , 2011). It is advised to use the average width of ChIP peaks from actual ChIP-seq data. We strongly encourage to set values when building a [parameterOptions](#) object.



**Value**

Returns chipMean slot from a [parameterOptions](#) object.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Kaplan T., Li X.-Y., Sabo P.J., Thomas S., Stamatoyannopoulos J.A., Biggin M.D., Eisen M.B. Quantitative models of the mechanisms that control genome-wide patterns of transcription factor binding during early *Drosophila* development, *PLoS Genet.*, 2011, vol. 7 pg. e1001290

**Examples**

```
# Building parameterOptions object
OPP <- parameterOptions()
#Accessing chipMean slot in OPP
chipMean(OPP)
```

---

chipMean-methods      *~~ Methods for Function chipMean ~~*

---

**Description**

*~~ Methods for function chipMean ~~*

**Methods:**

chipMean(object)

---

chipMean<-      *Access methods for chipMean slot in [parameterOptions](#) object.*

---

**Description**

Access methods for chipMean slot in [parameterOptions](#) object.

**Usage**

```
chipMean(object)<-value
```

**Arguments**

object      object is a [parameterOptions](#) object.

value      value is a positive numeric value that will be assigned to the chipMean slot. chipMean describes the average size of a ChIP-seq peak in base pairs.

**Details**

Default value : 150 When computing ChIP-seq like profiles (`computeChIPProfile`), the occupancy values given by `computeOccupancy` are transformed into ChIP-seq like profiles. The average size of a ChIP-seq peak was described by Kaplan (Kaplan et al. , 2011). It is advised to use the average width of ChIP peaks from actual ChIP-seq data. We strongly encourage to set values when building a `parameterOptions` object.

**Value**

Returns a `parameterOptions` object with an updated value for `chipMean` slot.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Kaplan T., Li X.-Y., Sabo P.J., Thomas S., Stamatoyannopoulos J.A., Biggin M.D., Eisen M.B. Quantitative models of the mechanisms that control genome-wide patterns of transcription factor binding during early *Drosophila* development, *PLoS Genet.*, 2011, vol. 7 pg. e1001290

**Examples**

```
# Building parameterOptions object
OPP <- parameterOptions()
# Setting new value for slot
chipMean(OPP) <- 250
```

---

chipMean<-methods      *~~ Methods for Function chipMean<- ~~*

---

**Description**

*~~ Methods for function chipMean<- ~~*

**Methods:**

`chipMean(object)<-value`

---

ChIPScore-class	Class "ChIPScore"
-----------------	-------------------

---

### Description

ChIPScore is the result of the [processingChIP](#) function. This object contains the extracted ChIP Score from ChIP data, the loci of interest and optional parameters associated to ChIPAnalyser. The loci of interest will either be user provided or the top n regions as defined by the reduce argument in processingChIP. This object has the sole purpose of aiding the storage and parsing of data and parameters.

### Objects from the Class

Object of this class are created internally and will be parsed to other objects as is.

### Slots

**scores:** Object of class "list" List of extracted ChIP scores  
**loci:** Object of class "loci" GRanges containing loci of interest  
**ploidy:** Object of class "numeric" Ploidy level of the organism  
**boundMolecules:** Object of class "vector" Number of Bound molecules to the DNA  
**backgroundSignal:** Object of class "numeric" ChIP background signal (average ChIP score)  
**maxSignal:** Object of class "numeric" max ChIP signal  
**lociWidth:** Object of class "numeric" Width of loci if reduce is used and no loci are provided  
**chipMean:** Object of class "numeric" Average ChIP peak width  
**chipSd:** Object of class "numeric" Standard Deviation of ChIP peak width  
**chipSmooth:** Object of class "vector" Smoothing window width for ChIP score  
**stepSize:** Object of class "numeric" Defining resolution size of ChIP like profiles (10bp = signal will be only considered every 10bp)  
**removeBackground:** Object of class "numeric" Signal Threshold to be removed. Default removes all negative scores  
**noiseFilter:** Object of class "character" Type of noise filter to be used on ChIP data.  
**PWMThreshold:** Object of class "numeric" Threshold of PWM scores that will be selected  
**strandRule:** Object of class "character" Rule to compute strand score (max, mean or sum)  
**whichstrand:** Object of class "character" Which strand should be used to compute PWM scores.  
**lambdaPWM:** Object of class "vector" Lambda value - Scaling factor to the PWM  
**naturalLog:** Object of class "logical" PFM to PWM conversion log transform ( natural log or log2)  
**noOfSites:** Object of class "nos" Number of Sites in the PWM that should be used to compute PWM scores.  
**PWMpseudocount:** Object of class "numeric" PWM pseudocount value for PFM to PWM conversion.  
**paramTag:** Object of class "character" Internal Tag - Code progression

**Extends**

Class "[parameterOptions](#)", directly.

**Methods**

**.loci**<- signature(object = "ChIPScore", value = "loci"): ...  
**.scores**<- signature(object = "ChIPScore", value = "list"): ...  
**initialize** signature(.Object = "ChIPScore"): ...  
**loci** signature(object = "ChIPScore"): ...  
**scores** signature(object = "ChIPScore"): ...  
**show** signature(object = "ChIPScore"): ...

**Author(s)**

Patrick C.N. Martin

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**See Also**

[processingChIP](#)

**Examples**

```
showClass("ChIPScore")
```

---

chipSd

*Accessor method for chipSd slot in a [parameterOptions](#) object.*

---

**Description**

Access or Extract chipSd slot in a [parameterOptions](#) object.

**Usage**

```
chipSd(object)
```

**Arguments**

object            object is a [parameterOptions](#)

## Details

When computing ChIP-seq like profiles (`computeChIPProfile`, the occupancy values given by `computeOccupancy` are transformed into ChIP-seq like profiles. The average size of a ChIP-seq peak was described by Kaplan (Kaplan et al. , 2011). The average peak size is subject to variation. This variation is accounted for with `chipSd`. It is advised to use the standard deviation of ChIP peak width from actual ChIP-seq data. We strongly encourage to set values when building a `parameterOptions` object.

## Value

Returns a `parameterOptions` object with an updated value for `chipSd`.

## Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Kaplan T.,Li X.-Y.,Sabo P.J.,Thomas S.,Stamatoyannopoulos J.A., Biggin M.D., Eisen M.B. Quantitative models of the mechanisms that control genome-wide patterns of transcription factor binding during early *Drosophila* development, *PLoS Genet.*,2011, vol. 7 pg. e1001290

## Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
# Accessing chipSd slot
chipSd(OPP)
```

---

chipSd-methods

~~ *Methods for Function chipSd* ~~

---

## Description

~~ Methods for function `chipSd` ~~

## Methods:

`chipSd(object)`

---

chipSd<- *Setter methods for chipSd slot in a [parameterOptions](#) object.*

---

### Description

Setter methods for chipSd slot in a [parameterOptions](#) object.

### Usage

```
chipSd(object)<-value
```

### Arguments

object	object is <a href="#">parameterOptions</a> object.
value	value is a positive numeric value that will be assigned to chipSd slot. Default value is 150.

### Details

When computing CHIP-seq like profiles ([computeChIPProfile](#), the occupancy values given by [computeOccupancy](#) are transformed into CHIP-seq like profiles. The average size of a CHIP-seq peak was described by Kaplan (Kaplan et al. , 2011). The average peak size is subject to variation. This variation is accounted for with `chipSd`. It is advised to use the standard deviation of CHIP peak width from actual CHIP-seq data. We strongly encourage to set values when building a [parameterOptions](#) object.

### Value

Returns a [parameterOptions](#) object with an updated value for `chipSd`.

### Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Kaplan T.,Li X.-Y.,Sabo P.J.,Thomas S.,Stamatoyannopoulos J.A., Biggin M.D., Eisen M.B. Quantitative models of the mechanisms that control genome-wide patterns of transcription factor binding during early Drosophila development, *PLoS Genet.*,2011, vol. 7 pg. e1001290

### Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
# Setting new value for chipSd slot
chipSd(OPP) <- 250
```

---

chipSd<-methods      *~~ Methods for Function chipSd<- ~~*

---

**Description**

~~ Methods for function chipSd<- ~~

**Methods:**

chipSd(object)<-value

---

chipSmooth      *Accessor methods for chipSmooth slot in a [parameterOptions](#) object.*

---

**Description**

Access or Extract chipSmooth slot in a [parameterOptions](#) object.

**Usage**

chipSmooth(object)

**Arguments**

object      object is a [parameterOptions](#) object.

**Details**

When computing ChIP-seq like ([computeChIPProfile](#)) profile from occupancy data (see [computeOccupancy](#)), the profiles are smoothed using a window of a given size. The default value is set at 250 base pairs. If chipSmooth is set to 0 then the profile will not be smoothed. We strongly encourage to set values when building a [parameterOptions](#) object.

**Value**

Returns the chipSmooth slot in an [parameterOptions](#) object.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
# Building parameterOptions object
OPP <- parameterOptions()
# Accessing chipSd slot
chipSmooth(OPP)
```

---

chipSmooth-methods      *~~ Methods for Function chipSmooth ~~*

---

**Description**

*~~ Methods for function chipSmooth ~~*

**Methods:**

signature(object = "parameterOptions")

---

chipSmooth<-                      *Setter method for chipSmooth slot in [parameterOptions](#) object.*

---

**Description**

Setter method for chipSmooth slot in [parameterOptions](#) object.

**Usage**

```
chipSmooth(object) <- value
```

**Arguments**

object                      object is a [parameterOptions](#) object.  
value                        value is the positive numeric value to be assigned to the chipSmooth slot in [parameterOptions](#) Default value is 250 base pairs.

**Details**

When computing ChIP-seq like ([computeChIPProfile](#)) profile from occupancy data (see [computeOccupancy](#)), the profiles are smoothed using a window of a given size. The default value is set at 250 base pairs. If chipSmooth is set to 0 then the profile will not be smoothed. We strongly encourage to set values when building a [parameterOptions](#) object.

**Value**

Returns a [parameterOptions](#) object with an updated value for chipSmooth slot.

**Author(s)**

Patrick C.N Martin <pm16057@essex.ac.uk>



## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

## Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
# Setting new value for chipSd slot
chipSmooth(OPP) <- 250
```

---

chipSmooth<-methods     *~~ Methods for Function chipSmooth<- ~~*

---

## Description

~~ Methods for function chipSmooth<- ~~

### Methods:

signature(object = "parameterOptions", value = "vector")

---

computeChIPProfile     *Computing ChIP-seq like profiles from Occupancy data.*

---

## Description

computeChIPProfile compute ChIP-seq like profile from occupancy data. Occupancy data is computed using [computeOccupancy](#).

## Usage

```
computeChIPProfile(genomicProfiles, loci, parameterOptions = NULL,
  norm = TRUE, method = c("moving_kernel", "truncated_kernel", "exact"),
  peakSignificantThreshold= NULL, cores=1, verbose = TRUE)
```

## Arguments

genomicProfiles

genomicProfiles is the result of [computeOccupancy](#). This object should be a [genomicProfiles](#) object.

loci

loci is either a [GRanges](#) or [ChIPScore](#) object. ChIPScore-class will be the result of [processingChIP](#). This object represents the set of Loci you are interested in analysing. If you have followed the full ChIPanalyser pipe line, you would have used the processingChIP function that would return a ChIPScore-class object containing your loci of interest. GRanges are also supported if you are only using part of the pipeline.

**parameterOptions**

parameterOptions is a [parameterOptions](#) object. This object is used to store the numerous parameters offered by ChIPAnalyser. This argument is optional as all arguments are also parse in both ChIPScore-class and genomicProfiles objects. If you wanted to make some last minute changes, [parameterOptions](#) is the way to go. We recommend that you set your desired options before hand.

**norm**

norm is a logical value. If TRUE, the ChIP-seq like profile will be normalised towards maximum Occupancy. If FALSE, the profile will be left as is.

**method**

method is a character string of one of the following: c("moving\_kernel", "truncated\_kernel", "exact"). If set to moving\_kernel, the peaks will be approximated using Rcpp (Default). If set to truncated\_kernel, the peaks will be approximated however this method does not require Rcpp. If set to exact, the peaks will not be approximated.

**peakSignificantThreshold**

peakSignificantThreshold is a threshold at which peaks will be selected. IMPORTANT: if you select "moving\_kernel" as described in method then this threshold is a numeric value describing the peak tail height cutoff value (Default = 0.001). In the case of "truncated\_kernel" and "exact", the threshold represents a distance in base pair from the peak summit at which the peak should be cut (Default = 1250). The default is set to NULL in this function. This just means that either the value is provided by user with the appropriate method. If not, the default will be selected depending on the method selected.

**cores**

cores is the number of cores that will be used to compute CHIP profiles.

**verbose**

verbose is a logical value. If TRUE, progress messages will be displayed in console. If FALSE, no progress messages will be displayed in console.

**Details**

computeChIPProfile converts Transcription Factor occupancy to a profile resembling the one of a ChIP-seq profile. Internally a few parameters are required to build a ChIP like profile. These parameters are either defined and stored in a [ChIPScore](#) object (Parameters are updated based on your ChIP data), a [genomicProfiles](#) (user defined at the start of the analysis) or a [parameterOptions](#) (if you want to update values as you go along)

**Value**

Returns a [genomicProfiles](#) object containing all ChIP-seq like profile for every combination of [lambdaPWM](#) and [boundMolecules](#) provided by the user.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```

#Extracting Data
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM, PFMFormat="JASPAR",BPFrequency=DNASequenceSet)

# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet)

#Compute PWM Scores
PWMScores <- computePWMScore(genomicProfiles = GenomeWide,
  DNASequenceSet = DNASequenceSet, loci = top, chromatinState = Access)
#Compute Occupancy
Occupancy <- computeOccupancy(genomicProfiles = PWMScores)

#Compute ChIP profiles
chipProfile <- computeChIPProfile(genomicProfiles=Occupancy,loci=top)
chipProfile

```

---

```
computeGenomeWideScores
```

*Computing Genome Wide scores*

---

**Description**

computeGenomeWideScores compute the max and min PWM score over the entire genome.

**Usage**

```
computeGenomeWideScores(genomicProfiles, DNASequenceSet, chromatinState = NULL, parameterOptions =
```

**Arguments**

genomicProfiles

genomicProfiles is a genomicProfiles object containing the PFM, PWM of interest.

DNASequenceSet DNASequenceSet is a BSgenome or DNAStringSet containing the sequence of the organism of interest.

**chromatinState** chromatinState is a GRanges object containing the chromatin States. This can either represent regions of accessible DNA or Chromatin state affinities.

**parameterOptions** parameterOptions is a parameterOptions object containing parameters that you wish to change. The genomicProfiles object will be updated using the values assigned to parameterOptions

**cores** cores is the number of cores that will be used (Numeric value - Default = 1 )

**verbose** verbose is a logical value that will determine if internal progress message will be printed.

### Details

computeGenomeWideScores function computes PWM scores over the entire genome (or accessible Genome if chromatin State are provided ). Genome wide scores are used to determine the maximum and minimum PWM score as well as the average exponential score. These scores will in turn be used to determine which score are above the PWM threshold. The average exponential score is an integrable part of the equation used to compute Occupancy. Using default settings, ChIPanalyser will only compute occupancy on the top 70% of PWM scores. This threshold can be changed. See [PWMThreshold](#)

### Value

Returns a genomicProfiles object with updated values for max score, min score and average-ExpPWMScore.

### Author(s)

Patrick C.N Martin <pm16057@essex.ac.uk>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

### Examples

```
if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR", BPFrequency=DNASequenceSet)

# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet)
```

---

computeOccupancy	<i>Compute Occupancy values from PWM Scores based on model.</i>
------------------	---

---

### Description

computeOccupancy will compute the Occupancy from PWM Scores. As described in detail in the vignette, ChIPAnalyser uses PWM Scores, DNA Accessibility data, the number of bound molecules and a scaling factor of Transcription Factor specificity. This function will compute occupancy using the values assigned to each variable.

### Usage

```
computeOccupancy(genomicProfiles, parameterOptions = NULL,
                 norm = TRUE, verbose = TRUE)
```

### Arguments

genomicProfiles	genomicProfiles is a <a href="#">genomicProfiles</a> object resulting from <a href="#">computePWMScore</a> . IT is important to use this resulting object as the occupancy will only be computed for sites above a threshold.
parameterOptions	parameterOptions is a <a href="#">parameterOptions</a> object containing the adequate values assigned to each Parameter. If not Supplied (parameterOptions = NULL), a new object will be created internally using default values.
norm	norm a logical value which determines if the occupancy should be normalised or not.
verbose	verbose a logical value which determines if progress messages are printed or not.

### Details

computeOccupancy will compute the Occupancy from PWM Scores. As described in detail in the vignette, ChIPAnalyser uses PWM Scores, DNA Accessibility data, the number of bound molecules and a scaling factor of Transcription Factor specificity. This function will compute occupancy using the values assigned to each variable. It should also be noted that the [parameterOptions](#) object contains a set of parameters used to compute Occupancy (not only restricted to this ). These parameters are often dependant on real ChIP-Seq data and will influence the goodness of fit between the predicted model and real ChIP-seq data. We strongly advise that the values assigned to each parameter should be customized in order to increase the model agreement with real world biological data.

### Value

computeOccupancy will return a [genomicProfiles](#). The main difference will reside in the [profiles](#) slot. This slot is generally a list or [GRangesList](#). Within these list type structures are enclosed [GRanges](#) containing the positions of site above threshold, PWM Scores and Occupancy for each site. The series of GRanges will depend on the number of loci that are tested and the number of element in the list will depend on the various combinations of lambdaPWM and boundMolecules.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)

#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR",BPFrequency=DNASequenceSet)
OPP <- parameterOptions()
# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet)

#Compute PWM Scores
PWMScores <- computePWMScore(genomicProfiles = GenomeWide,
  DNASequenceSet = DNASequenceSet,
  loci = top,
  chromatinState = Access)
#Compute Occupancy
Occupancy <- computeOccupancy(genomicProfiles = PWMScores,
  parameterOptions = OPP)
Occupancy
```

---

computeOptimal

*compute Optimal Parameters*

---

**Description**

ChIPAnalyser contains a set of functions some of which require two parameters known as [lambdaPWM](#) and as [boundMolecules](#). These two paramters are not always known. computeOptimal will compute these values by maximising the correlation and minimising the Mean Squared Error between a predicted ChIP-seq-like profile and a real ChIP-seq profile for a given loci.

**Usage**

```
computeOptimal(genomicProfiles, DNASquenceSet, ChIPScore, chromatinState = NULL,
  parameterOptions = NULL, optimalMethod = "all", rank=FALSE, returnAll=TRUE,
  peakMethod="moving_kernel", cores=1)
```

**Arguments**

genomicProfiles

genomicProfiles is a [genomicProfiles](#) object containing at least a Position Frequency Matrix or a Position Weight Matrix. It is strongly advised to customize this object to increase goodness of fit of the model when compared to real ChIP-seq data.

DNASquenceSet

DNASquenceSet is a [DNAStringSet](#) or a [BSgenome](#) of the full sequence of the organism of interest.

ChIPScore

ChIPScore is a named list containing ChIP-seq enrichments for each Loci of interest. This Profile should be normalised to a base pair level. In other words, there should be an enrichment score for each base pair of a given Locus.

chromatinState

chromatinState is a [GRanges](#) object containing either accesible sites or DNA affinity scores.

parameterOptions

[parameterOptions](#) is a [parameterOptions](#) object. If this object is not provided (parameterOptions = NULL), a new object will be created internally. However, it is strongly advised to tailor this object to maximise the goodness of fit of the model when compared to ChIP-seq data.

optimalMethod

optimalMethod is a character string which determines which method for optimal parameter selection should be selected. optimalMethod can be one of the following: pearson, spearman, kendall, ks, fscore, geometric, MSE, or all. Default is set to all.

rank

rank is a logical value indicating if optimal parameters should be based on rank (parameter combination occurring the most over all regions) or average score (best performing combination of paramters on average over all regions selected). DEFAULT = FALSE

returnAll

returnAll is a logical value indicating if all internal objects should be returned. DEFAULT = TRUE. Internal objects are the following: Occupancy Scores, ChIP like profiles, goodness of fit metrics and optimal paramters. If set to FALSE, computeOptimal will only return the optimal parameters.

peakMethod

peakMethod is a character string of one of the following: c("moving\_kernel", "truncated\_kernel", "exact"). If set to moving\_kernel, the peaks will be approximated using Rcpp (Default). If set to truncated\_kernel, the peaks will be approximated however this method does not require Rcpp. If set to exact, the peaks will not be approximated.

cores

cores is the number cores that will be used to compute optimal set of parameters.

**Details**

In order to backward infer the values of [lambdaPWM](#) and [boundMolecules](#), it is possible to use the computeOptimal to find these parameters. It should be noted that this functions requires a ChIP-seq data input. ChIPScore (ChIP-seq data). This should be the output of the [processingChIP](#) function.

**Value**

computeOptimal returns a list respectively described as the optimal set of Parameters ([lambda - lambdaPWM](#) and [boundMolecules](#)), the optimal matrix (a matrix containing accuracy estimates dependant on the parameter chosen), and finally the chosen parameter. If the parameter that was chosen was "all", then each element of this list will contain the optimal set of parameters, optimal matrices for all of the aforementioned parameters (see [optimalMethod](#)).

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata", package="ChIPAnalyser"), "BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
chip<-processingChIP(chip,top)
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR",BPFfrequency=DNASequenceSet)
OPP <- parameterOptions()
#Computing Optimal set of Parameters
optimalParam <- computeOptimal(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet,
  ChIPScore = chip,
  chromatinState = Access,
  parameterOptions = OPP,
  parameter = "all",
  peakMethod="moving_kernel")
```

---

computePWMScore

*Compute PWM Scores of sites above threshold.*

---

**Description**

computePWMScore will compute and extract all sites that exhibit a PWM Score higher than a threshold. This threshold (see [PWMThreshold](#)) will determine the percentage of total sites that should NOT be considered.



**Usage**

```
computePWMScore(genomicProfiles, DNASequenceSet,
  loci = NULL, chromatinState = NULL, parameterOptions=NULL, cores=1, verbose = TRUE)
```

**Arguments**

**DNASequenceSet** DNASequenceSet is a [DNAStrngSet](#) or a BSgenome containing the full sequence of the organism of interest.

**genomicProfiles** genomicProfiles is a [genomicProfiles](#) object resulting from the [computeGenomeWideScores](#) function.

**loci** loci is a [GRanges](#) object containing the Loci of interest or a [ChIPScore](#) object result of [processingChIP](#) function.

**parameterOptions** parameterOptions is a [parameterOptions](#) object containing parameters that you wish to parse/change when computing PWMScores.

**chromatinState** chromatinState is a [GRanges](#) object sites of accessible DNA or DNA affinity scores.

**cores** cores is the number of cores used to compute PWM Scores.

**verbose** verbose is a logical value indicating if progress messages should be printed or not.

**Details**

After determining genome wide scores, it is possible to only compute and extract high affinity sites (in the sense that they have a high PWM Score). If a [PWMThreshold](#) is not set by user, the default value is set at 0.7. This means that 70 % of sites will NOT be selected. Only the top 30 % will be computed and extracted. If one is interested in all PWM Scores at a genome wide scale ( or accessible DNA ), this is possible by setting [PWMThreshold](#) to zero.

**Value**

[computePWMScore](#) will return a [genomicProfiles](#) object. The [profiles](#) slot will have been updated. This slot will now contain a [GRangesList](#) with each element being a [GRanges](#). This [GRanges](#) will contain position of each sites (start, end and strand) and the PWMScore associated to that site.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata", package="ChIPAnalyser"), "BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome
```

```
if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
chip<-processingChIP(chip,top)
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR",BPFrequency=DNASequenceSet)

# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GPP)

#Compute PWM Scores
PWMScores <- computePWMScore(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GenomeWide,
  loci = chip, chromatinState = Access)
PWMScores
```

---

DNASequenceLength      *Accessor method for DNASequenceLength slot in a [genomicProfiles](#)*

---

## Description

Accessor method for DNASequenceLength slot in a [genomicProfiles](#)

## Usage

```
DNASequenceLength(object)
```

## Arguments

object                    object is a [genomicProfiles](#)

## Details

The model on which is based ChIPanalyser requires the length of the DNA sequence used to compute scores. In this circumstance, this DNA Length is the total length of the DNA of the organism of interest or the the Accessible DNA at a genome wide scale.

## Value

Returns DNASequenceLength slot in a [genomicProfiles](#) object.

## Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)

#Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR",BPFrequency=DNASequenceSet)
# Computing Genome Wide
GenomceWide <- computeGenomeWideScores(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GPP)

DNASequenceLength(GenomceWide)
```

---

DNASequenceLength-methods

~~ *Methods for Function* DNASequenceLength ~~

---

**Description**

~~ Methods for function DNASequenceLength ~~

**Methods:**

signature(object = "genomicProfilesInternal")

---

drop

*Accessor Method for the drop slot in a [genomicProfiles](#) object.*

---

**Description**

Accessor Method for the drop slot in a [genomicProfiles](#) object.

**Usage**

drop(object)

**Arguments**

object                    object is a `genomicProfiles` object.

**Details**

During certain computations, it is possible that the Loci of interest do not show any overlap with accessible DNA. If this were to be the case, a warning message will appear in the console but these inaccessible Loci will be stored in this slot. It is also for these reasons that it is imperative for Loci of interest to be named (in this case, a named `GRanges`).

**Value**

Returns a character string with loci containing no accessible DNA.

**Author(s)**

Patrick C.N. Martin <p.martin@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Loading data
data(ChIPAnalyserData)
#Loading PFM files
PFM <- file.path(system.file("extdata", package="ChIPAnalyser"), "BEAF-32.pfm")
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR")

# Loci with no acces - a warning message will be issued
#if loci do not contain accessible DNA
# Otherwise this slot will remain empty

drop(GPP)
```

---

drop-methods

~~ *Methods for Function drop* ~~

---

**Description**

~~ Methods for function drop ~~

**Methods:**

signature(object = "genomicProfilesInternal")

---

evolve *Running the ChIPanalyser implementation of a Genetic algorithm.*

---

### Description

evolve pushes a starting population to evolve in a genetic algorithm.

### Usage

```
evolve(population, DNASequenceSet, ChIPScore,
       genomicProfiles, parameters=NULL, generations=100, mutationProbability=0.3,
       offsprings=5, chromatinState=NULL,
       method="geometric", lambda=TRUE,
       checkpoint=TRUE,
       filename=NULL, cores=1)
```

### Arguments

population	numeric value describing the number of individuals in the starting population. Alternatively - a starting population list as returned by generateStartingPopulation. NOTE: if numeric - the parameter argument is also required.
DNASequenceSet	DNAStrngSet object containing DNA sequences of interest (Extracted from BSgenome)
ChIPScore	ChIPScore object as returned by the processingChIPfunction
genomicProfiles	genomicProfiles object containing minimal information (such as the PWM)
parameters	vector or list containing each parameter that should be added to the chromosome. See generateStartingPopulation
generations	numeric describing the number of generation before the Genetic algorithm should halt.
mutationProbability	numeric describing the rate of mutations for each surviving individual
offsprings	numeric describing the number of individuals surviving to the next generation
chromatinState	GRanges object containing chromatin state information. Each state should be labeled in a meta data column named "name". It is advised to use numeric values for each state name.
method	character string describing the scoring metric that should be used. ChIPanalyser offers twelve different metrics: correlation coefficients (Pearson, Spearman and Kendall), Mean Squared Error (MSE), Kolmogorov–Smirnov Distance, precision, recall, accuracy, F-score, Matthew’s correlation coefficient (MCC) and Area Under Curve Receiver Operator Characteristic (AUC ROC or just AUC)
lambda	logical describing if lambda value should be pre-computed. Setting to TRUE increases the speed of the algorithm.
checkpoint	logical describing if population parameters at each generations should be saved.
filename	character string that will serve as a prefix to the saved intermediate files.
cores	numeric describing the number of cores used to run the GA.

**Details**

ChIPanalyser offers a way of finding optimal solution by using a genetic algorithm. Instead of running the standard analysis, TF binding affinities to chromatin states can be extracted via this more complex method. It should be noted that this method is better suited for the analysis of chromatin states. While the algorithm still works with simple DNA Accessibility, it would potentially take more time for accuracy minor gains.

**Value**

Returns a named list with three elements.

- database saves the data frame containing all scores for each individual since generation 1
- population saves the last population with chromosome values
- fittest saves the fittest individual for a given generation

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**Examples**

```
library(ChIPanalyser)
data(ChIPanalyserData)
# See GA vignette for usage
```

---

```
generateStartingPopulation
```

*Generate Starting population for ChIPanalyser Genetic algorithm*

---

**Description**

generateStartingPopulation generates a starting population with random traits for each individual

**Usage**

```
generateStartingPopulation(population,parameters,names=NULL)
```

**Arguments**

population	numeric value describing the number of individuals in the starting population.
parameters	vector or list containing each parameter that should be added to the chromosome.
names	character describing names that should be added to each individual.

**Details**

generateStartingPopulation generates a starting population to be used in the genetic algorithm implemented in ChIPanalyzer. There are two main ways a starting population can be generated:

1. by name Using names of each parameter that should be parse to each "chromosome". The possible parameters are N, lambda, PWMThreshold, CS ( DNAAffinity or DNAAccessibility also works). CS values should also contain a numeric value associated to each chromatin state you wish to parse. e.g CS1 ... CS14 This will generate a value by sampling from a set of predefined value for each parameters.
2. by value range Using a named list (names for each parameters). Each element of the list should contain three numeric values : length of range, min value, max value. (Internally - values are parse to runif)

**Value**

Returns a list of individuals with a random traits

**Author(s)**

Patrick C.N. Martin

**Examples**

```
## by name
param <- c("N", "lambda", "PWMThreshold", "CS1", "CS2", "CS3")

pop <- generateStartingPopulation(20, param, names = NULL)

# by range
paramValue <- list(c(10, 1, 1000), c(10, 0, 5), c(10, 0, 0.9), c(10, 0, 1), c(10, 0, 1), c(10, 0, 1))

pop <- generateStartingPopulation(20, paramValue, names= param)
```

---

genomicProfiles

*Genomic Profile object*

---

**Description**

genomicProfiles is an S4 object serving two purposes: (i) storing internal computed data and (ii) storing parameter options. This object is parsed through the different steps of the pipeline to facilitate that parsing and changing of parameters.

**Usage**

```
genomicProfiles(..., parameterOptions = NULL, genomicProfiles = NULL, ChIPScore = NULL)
```

## Arguments

...	Any of the user available slots in genomicProfiles.
parameterOptions	If some parameters were already previously computed or stored in a parameterOptions, parsing this object will use those values instead of the default ones.
genomicProfiles	If some parameters were already previously computed or stored in a genomicProfiles, parsing this object will use those values instead of the default ones.
ChIPScore	If some parameters were already previously computed or stored in a ChIPScore, parsing this object will use those values instead of the default ones.

## Details

The genomicProfiles object serves the purpose of storing, and parsing parameters and computed data between the different steps of the pipeline. When creating a genomicProfiles object it is possible to use previously computed values by simply parsing the object to the constructor function.

## Value

Returns a genomicProfiles object with updated slots for all parameters parsed.

## Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

## See Also

[genomicProfiles](#)

[parameterOptions](#)

## Examples

```
PFM <- file.path(system.file("extdata", package="ChIPAnalyser"), "BEAF-32.pfm")
genomicProfiles()
genomicProfiles(PFM=PFM, PFMFormat="JASPAR")
```



---

genomicProfiles-class *Class "genomicProfiles"*

---

## Description

genomicProfiles is an S4 object serving two purposes: (i) storing internal computed data and (ii) storing parameter options. This object is parsed through the different steps of the pipeline to facilitate that parsing and changing of parameters.

## Objects from the Class

Objects can be created by calls of the form `genomicProfiles(ploidy, boundMolecules, backgroundSignal, maxSignal, lociWidth, chipMean, chipSd, chipSmooth, stepSize, noiseFilter, removeBackground, lambdaPWM, PWMpseudocount, naturalLog, noOfSites, PWMThreshold, strandRule, whichstrand, PFM, PWM, PFMFormat, BPFfrequency, minPWMScore, maxPWMScore, profiles, DNASequenceLength, averageExpPWMScore)`.

## Slots

**PWM:** Object of class "matrix": A Position Weight Matrix (either supplied or internally computed if PFM is provided)

**PFM:** Object of class "matrix": A Position Frequency Matrix (may also be a path to file containing PFM)

**PFMFormat:** Object of class "character": A character string of one of the following: raw, trans-fac, JASPAR or sequences

**BPFfrequency:** Object of class "vector": Base Pair Frequency in the genome (if a DNA sequence is provided (as a `DNAStrngSet` or `BSgenome`), will be automatically computed internally). Default: `c(0.25,0.25,0.25,0.25)`

**minPWMScore:** Object of class "vector": Lowest PWM score across the genome (computed and updated internally)

**maxPWMScore:** Object of class "vector": Highest PWM score across the genome (computed and updated internally)

**profiles:** Object of class "GRList": Contains GRanges with sites above threshold and associated metrics (PWMscore and Occupancy) - Computed Internally

**DNASequenceLength:** Object of class "vector": Length of the Genome (or accessible genome) - computed internally

**averageExpPWMScore:** Object of class "vector": Average exponential PWM score across the genome (or accessible genome) - computed internally

**ZeroBackground:** Object of class "vector": Internal background value (computed internally)

**drop:** Object of class "vector": Stores Loci that do contain accessible DNA if it were to be the case (computed and updated internally)

**tags:** Object of class "character" ~Internal Tags~

**ploidy:** Object of class "numeric": A numeric Value describing the ploidy of the organism. Default: 2

**boundMolecules:** Object of class "vector": A vector (or single value) containing the number of bound Molecules (bound Transcription Factors): Default: 1000

backgroundSignal: Object of class "numeric": A numeric value describing the ChIP-seq background Signal (average signal from real ChIP seq data). Default: 0

maxSignal: Object of class "numeric": A numeric value describing the highest ChIP-seq signal (from real ChIP-seq data). Default: 1

lociWidth: Object of class "numeric" ~

chipMean: Object of class "numeric": A numeric value describing the mean width of a ChIP- seq peak. Default:150

chipSd: Object of class "numeric": A numeric value describing the standard deviation of ChIP-seq peaks. Default: 150

chipSmooth: Object of class "vector": A numeric value describing the width of the window used to smooth Occupancy profiles into ChIP profiles. Default:250

stepSize: Object of class "numeric": A numeric value describing the step Size (in base pairs) between each ChIP-seq score. Default:10 (Scored every 10 base pairs)

removeBackground: Object of class "numeric": A numeric value describing the value at which score should be removed. Default:0 (If negative scores then remove)

noiseFilter: Object of class "character" ~Describes the noiseFilter method that will be applied to ChIP data (Zero, mean, median, sigmoid)~

PWMThreshold: Object of class "numeric": Threshold at which PWM Score should be selected (only sites above threshold will be selected - between 0 and 1)

strandRule: Object of class "character": "mean", "max" or "sum" will determine how strand should be handle for computing PWM Scores. Default : "max"

whichstrand: Object of class "character": "+", "-" or "+-" on which strand should PWM Score be computed. Default: "+-"

lambdaPWM: Object of class "vector" A vector (or single value) containing values for lambdaPWM Default:1

naturalLog: Object of class "logical": A logical value describing if natural Log will be used to compute the PWM (if FALSE then log2 will be used). Default: TRUE

noOfSites: Object of class "nos" A Positive integer describing number of sites (in base pair) should be used from the PFM to compute PWM. Default =0 (Full width of binding site will be used when set to 0)

PWMPseudocount: Object of class "numeric": A numeric value describing a PWMPseudocount for PWM computation. Default:1

paramTag: Object of class "character" ~Internal~

### Extends

Class "[genomicProfilesInternal](#)", directly. Class "[parameterOptions](#)", directly.

### Methods

**initialize** signature(.Object = "genomicProfiles"): ...

**show** signature(object = "genomicProfiles"): ...

### Author(s)

Partick C. N. Martin <p.martin@essex.ac.uk>

## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

## See Also

[genomicProfiles parameterOptions](#)

## Examples

```
showClass("genomicProfiles")
```

---

```
genomicProfilesInternal-class  
  Class "genomicProfilesInternal"
```

---

## Description

Non exported class. Represents the stripped down version of genomicProfiles.

## Objects from the Class

Created Internally.

## Slots

PWM: Object of class "matrix" ~~  
PFM: Object of class "matrix" ~~  
PFMFormat: Object of class "character" ~~  
BPFrequency: Object of class "vector" ~~  
minPWMScore: Object of class "vector" ~~  
maxPWMScore: Object of class "vector" ~~  
profiles: Object of class "GRList" ~~  
DNASequenceLength: Object of class "vector" ~~  
averageExpPWMScore: Object of class "vector" ~~  
ZeroBackground: Object of class "vector" ~~  
drop: Object of class "vector" ~~  
tags: Object of class "character" ~~

**Methods**

```

.averageExpPWMScore<- signature(object = "genomicProfilesInternal", value = "numeric"):
  ...
.DNASequenceLength<- signature(object = "genomicProfilesInternal", value = "vector"):
  ...
.drop<- signature(object = "genomicProfilesInternal", value = "vector"): ...
.generatePWM signature(object = "genomicProfilesInternal"): ...
.maxPWMScore<- signature(object = "genomicProfilesInternal", value = "vector"): ...
.minPWMScore<- signature(object = "genomicProfilesInternal", value = "vector"): ...
.profiles<- signature(object = "genomicProfilesInternal", value = "GRLList"): ...
.tags signature(object = "genomicProfilesInternal"): ...
.tags<- signature(object = "genomicProfilesInternal", value = "character"): ...
averageExpPWMScore signature(object = "genomicProfilesInternal"): ...
BPFfrequency signature(object = "genomicProfilesInternal"): ...
BPFfrequency<- signature(object = "genomicProfilesInternal", value = "DNAStringSet"):
  ...
BPFfrequency<- signature(object = "genomicProfilesInternal", value = "vector"): ...
DNASequenceLength signature(object = "genomicProfilesInternal"): ...
drop signature(object = "genomicProfilesInternal"): ...
maxPWMScore signature(object = "genomicProfilesInternal"): ...
minPWMScore signature(object = "genomicProfilesInternal"): ...
PFMFormat signature(object = "genomicProfilesInternal"): ...
PFMFormat<- signature(object = "genomicProfilesInternal", value = "character"): ...
PositionFrequencyMatrix signature(object = "genomicProfilesInternal"): ...
PositionFrequencyMatrix<- signature(object = "genomicProfilesInternal", value = "character"):
  ...
PositionFrequencyMatrix<- signature(object = "genomicProfilesInternal", value = "matrix"):
  ...
PositionWeightMatrix signature(object = "genomicProfilesInternal"): ...
PositionWeightMatrix<- signature(object = "genomicProfilesInternal", value = "matrix"):
  ...
profiles signature(object = "genomicProfilesInternal"): ...

```

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**See Also**

[genomicProfiles](#)  
[parameterOptions](#)

**Examples**

```
showClass("genomicProfilesInternal")
```

---

`getHighestFitnessSolutions`  
*Get Highest Fitness Solutions*

---

**Description**

`getHighestFitnessSolutions` extract best solution from a ChIPanalyser GA/evolve Run.

**Usage**

```
getHighestFitnessSolutions(population,child=2,method="geometric")
```

**Arguments**

<code>population</code>	Population list as output by the evolve function.
<code>child</code>	numeric describing the number of solution to be extracted from Population list.
<code>method</code>	character string describing which scoring method should be used and selected from "geometric","ks","MSE","pearson","spearman","kendall", "recall","precesion","fscore","MCC" or "AUC".

**Details**

This function only serves as a way of extracting data from the poppulation list. Ultimately - it is just a wrapper for some indexing.

**Value**

Return the index of the top "child" solutions.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**Examples**

```
library(ChIPAnalyser)
data(ChIPAnalyserData)
# See GA vignette for usage
```

---

getTestingData      *Extract testing data from ChIPscore object*

---

**Description**

getTestingData extracts selected regions from ChIPscore object to be used as testing set.

**Usage**

```
getTestingData(ChIPscore, loci = 1)
```

**Arguments**

ChIPscore	ChIPscore object as returned by processingChIP
loci	numeric describing index of loci to be used as testing data.

**Value**

Returns ChIPscore object with the selected testing loci.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**Examples**

```
library(ChIPAnalyser)
data(ChIPAnalyserData)
# See GA vignette for usage
test <- processingChIP(chip, top)
test <- getTestingData(test, 1:2)
```

---

getTrainingData      *Extract training data from ChIPscore object*

---

**Description**

getTrainingData extracts selected regions from ChIPscore object to be used as training set.

**Usage**

```
getTrainingData(ChIPscore, loci = 1)
```

**Arguments**

ChIPscore	ChIPscore object as returned by processingChIP
loci	numeric describing index of loci to be used as training data.

**Value**

Returns ChIPscore object with the selected training loci.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**Examples**

```
library(ChIPAnalyser)
data(ChIPAnalyserData)
# See GA vignette for usage
test <- processingChIP(chip, top)
test <- getTrainingData(test, 1:2)
```

---

GRList-class

Class "GRList"

---

**Description**

Virtual Class to handle multiple data types for one slot ([profiles](#))

**Objects from the Class**

A virtual Class: No objects may be created from it.

**Methods**

[GRList-class](#) The purpose of this virtual class is to store data of two different formats in one slot: GRangesList and Lists

**Author(s)**

Patrick C. N. Martin <p.martin@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
showClass("GRList")
```

---

```
initialize-methods    ~~ Methods for Function initialize ~~
```

---

### Description

~~ Methods for function initialize ~~

#### Methods:

signature(.Object = "ChIPScore") Initialize ChIPScore

signature(.Object = "genomicProfiles") Initialize genomicProfiles

signature(.Object = "parameterOptions") Initialize parameterOptions

---

```
lambdaPWM            Accessor Method for the lambdaPWM slot in a parameterOptions ob-  
ject
```

---

### Description

Accessor Method for the lambdaPWM slot in a [parameterOptions](#) object

### Usage

```
lambdaPWM(object)
```

### Arguments

object            object is [parameterOptions](#) object

### Details

The model underlying ChIPanalyser internally infers two paramters: number of bound molecules and lambda. Lambda represents a scaling factor for the Position weight matrix (PWM). This can be described as how well does a TF discriminate between high affinity and very high affinity sites.

### Value

Returns the value assigned to the lambdaPWM slot in a [parameterOptions](#) object.

### Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.



**Examples**

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions(lambdaPWM=1)
#Setting new Value for lambdaPWM
lambdaPWM(GPP)
```

---

lambdaPWM-methods      *~~ Methods for Function lambdaPWM ~~*

---

**Description**

~~ Methods for function lambdaPWM ~~

**Methods:**

lambdaPWM(object)

---

lambdaPWM<-                      *Setter Method for the lambdaPWM slot in a [parameterOptions](#) object*

---

**Description**

Setter Method for the lambdaPWM slot in a [parameterOptions](#) object

**Usage**

```
lambdaPWM(object)<-value
```

**Arguments**

object	object is <a href="#">parameterOptions</a> object
value	value is the numeric value to be assigned to the lambdaPWM slot. Default set at 1.

**Details**

The model underlying ChIPAnalyser internally infers two paramters: number of bound molecules and lambda. Lambda represents a scaling factor for the Position weight matrix (PWM). This can be described as how well does a TF discriminate between high affinity and very high affinity sites.

**Value**

Returns the value assigned to the lambdaPWM slot in a [parameterOptions](#) object.

**Author(s)**

Patrick C. N. Martin <p.martin@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions(lambdaPWM=1)
#Setting new Value for lambdaPWM
lambdaPWM(GPP) <- 2
```

---

lambdaPWM<-methods      *~~ Methods for Function lambdaPWM<- ~~*

---

**Description**

Setter method for the lambdaPWM slot in the parameterOptions

**Methods:**

lambdaPWM(object)<-value

---

loci      *Accessor Method for the loci slot in a ChIPScore object*

---

**Description**

Setter Method for the loci slot in a ChIPScore object

**Usage**

loci(object)

**Arguments**

object      object is ChIPScore object

**Details**

When using the [processingChIP](#), this functions will return a name GRanges with the loci of interest. These loci will either result from user input or extracted from the ChIP profiles (see [processingChIP](#) and [lociWidth](#)). This functions enables you to extract those loci from the [ChIPScore](#) object.

**Value**

Returns the value assigned to the loci slot in a [ChIPScore](#) object.

**Author(s)**

Patrick C. N. Martin <p.martin@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Loading data
data(ChIPanalyserData)

chip<-processingChIP(chip,top)
loci(chip)
```

---

loci-class

*Class "loci"*

---

**Description**

Setter for Loci of interest parsed to or extracted from the ChIPScore object

**Objects from the Class**

A virtual Class: No objects may be created from it.

**Methods**

**.loci<-** signature(object = "ChIPScore", value = "loci"): ...

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**See Also**

[ChIPScore](#)

**Examples**

```
showClass("loci")
```

---

```
loci-methods      ~~ Methods for Function loci ~~
```

---

**Description**

Accessor method for the loci slot in [ChIPScore](#)

**Methods:**

loci{Object} Loci of interest parsed to or extracted from the ChIPScore object

---

```
lociWidth      Accessor Method for the lociWidth slot in a parameterOptions object
```

---

**Description**

Setter Method for the lociWidth slot in a [parameterOptions](#) object

**Usage**

```
lociWidth(object)
```

**Arguments**

object            object is [parameterOptions](#) object

**Details**

When using the [processingChIP](#) function, the provided ChIP scores will be split into bins of a given size. lociWidth determines the Size of that bin. Default is set at 20 000 bp. This means that the ChIP profiles provided will be split into bins of 20 000 bp over the entire profile provided if no loci of interest is provided.

**Value**

Returns the value assigned to the lociWidth slot in a [parameterOptions](#) object.

**Author(s)**

Patrick C. N. Martin <p.martin@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions(lociWidth=20000)
#Accessing new Value for lociWidth
lociWidth(GPP)
```

---

lociWidth-methods      *~~ Methods for Function lociWidth ~~*

---

**Description**

Accessor method for the loci slot in [ChIPScore](#)

**Methods:**

`lociWidth(object)` Setting width of regions when using the reduce argument and NOT providing your own loci when using the `processingChIP` function.

---

`lociWidth<-`                      *Setter Method for the lociWidth slot in a [parameterOptions](#) object*

---

**Description**

Setter Method for the lociWidth slot in a [parameterOptions](#) object

**Usage**

```
lociWidth(object)<-value
```

**Arguments**

object	object is <a href="#">parameterOptions</a> object
value	value is the numeric value to be assigned to the lociWidth slot. Default set at 1.

**Details**

When using the [processingChIP](#) function, the provided ChIP scores will be split into bins of a given size. `lociWidth` determines the Size of that bin. Default is set at 20 000 bp. This mean that the ChIP profiles provided will be split into bins of 20 000 bp over the entire profile provided if no loci of interest is provided.

**Value**

Returns the value assigned to the lociWidth slot in a [parameterOptions](#) object.

**Author(s)**

Patrick C. N. Martin <p.martin@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions(lociWidth=20000)
#Setting new Value for lociWidth
lociWidth(GPP) <- 30000
```

---

lociWidth<-methods      *~~ Methods for Function lociWidth<- ~~*

---

**Description**

Setter method for the loci slot in [ChIPScore](#)

**Methods:**

lociWidth(Object)<-value

---

maxPWMScore      *Accessor function for maxPWMScore slot in a [genomicProfiles](#) object.*

---

**Description**

Accessor function for maxPWMScore slot in a [genomicProfiles](#) object.

**Usage**

```
maxPWMScore(object)
```

**Arguments**

object      object is a [genomicProfiles](#) object.

**Details**

maxPWMScore is a numerical value that can be described as the highest PWM score computed at a genome wide scale. This value is computed and updated in the [genomicProfiles](#) object after using the [computeGenomeWideScores](#).

**Value**

Returns the value of assigned to the maxPWMScore slot in a [genomicProfiles](#) object.

**Author(s)**

Patrick C. N. Martin <p.martin@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Loading data
#Data extraction
data(ChIPanalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPanalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR")

# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GPP)
maxPWMScore(GenomeWide)
## If used before computeGenomeWidePWMScore, will return NULL
```

---

maxPWMScore-methods    *~~ Methods for Function maxPWMScore ~~*

---

**Description**

Accessor method for maxPWMScore

**Methods:**

maxPWMScore(object)

---

maxSignal	<i>Accessor method for the maxSignal slot in a <a href="#">parameterOptions</a> object.</i>
-----------	---

---

### Description

Accessor method for the maxSignal slot in a [parameterOptions](#) object.

### Usage

```
maxSignal(object)
```

### Arguments

object            object is a [parameterOptions](#) object.

### Details

In the context of ChIPanalyser, maxSignal represents the maximum normalised ChIP-Seq signal of a given Transcription factor (or DNA binding protein). Although, A default value of 1 has been assigned to this slot, we strongly recommend to tailor this value accordingly. We strongly encourage to set values when building a [parameterOptions](#) object.

### Value

Returns the value assigned to the maxSignal slot in a [parameterOptions](#) object.

### Author(s)

Patrick C.N. Martin <p.martin@essex.ac.uk>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

### Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for maxSignal
maxSignal(OPP)
```



---

maxSignal-methods      *~~ Methods for Function maxSignal ~~*

---

### Description

Accessor method for maxSignal

### Methods:

maxSignal(object) Maximum ChIP signal extracted from ChIP data (see processingChIP)

---

maxSignal<-                      *Setter method for maxSignal slot in a parameterOptions object.*

---

### Description

Setter method for maxSignal slot in a [parameterOptions](#) object.

### Usage

```
maxSignal(object) <- value
```

### Arguments

object                      object is a [parameterOptions](#) object.  
value                        value is a numerical value to be assigned to the maxSignal slot.

### Details

In the context of ChIPanalyser, maxSignal represents the maximum normalised ChIP-Seq signal of a given Transcription factor (or DNA binding protein). Although, A default value of 1 has been assigned to this slot, we strongly recommend to tailor this value accordingly. We strongly encourage to set values when building a [parameterOptions](#) object.

### Value

Returns a [parameterOptions](#) with an updated value for maxSignal.

### Author(s)

Patrick C.N. Martin <p.martin@essex.ac.uk>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for maxSignal
maxSignal(OPP) <- 1.8
```

---

maxSignal<-methods      *~~ Methods for Function maxSignal<- ~~*

---

**Description**

Setter method for maxSignal

**Methods:**

maxSignal(Object)<-value Maximum ChIP signal extracted from ChIP data (see processingChIP)

---

minPWMScore              *Accessor method the minPWMScore slot in a [genomicProfiles](#) object*

---

**Description**

Accessor method the minPWMScore slot in a [genomicProfiles](#) object

**Usage**

```
minPWMScore(object)
```

**Arguments**

object                  object is a [genomicProfiles](#) object.

**Details**

minPWMScore can be described as the lowest PWM score computed at a genome wide scale. Although it is possible to assign a value to minPWMScore, we strongly advise to use the value computed and assigned internally. This value is computed in the [computeGenomeWideScores](#) function.

**Value**

Returns the value assigned to the minPWMScore slot in a [genomicProfiles](#) object.

**Author(s)**

Patrick C. N. Martin <p.martin@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```

#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR")

# Computing Genome Wide
GenomceWide <- computeGenomeWideScores(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GPP)
minPWMScore(GenomceWide)

## If used before computeGenomeWidePWMScore, will return NULL

```

---

minPWMScore-methods    *~~ Methods for Function minPWMScore ~~*

---

**Description**

Accessor for minPWMScore

**Methods:**

minPWMScore(object) Minimum PWM score computed during the computeGenomeWideScores step.

---

naturalLog                      *Accessor method the naturalLog slot in a [parameterOptions](#) object.*

---

**Description**

Accessor method the naturalLog slot in a [parameterOptions](#) object.

**Usage**

```
naturalLog(object)
```

**Arguments**

object                      object is [parameterOptions](#) object.

## Details

During the computation of a Position Weight Matrix, the Position Probability Matrix (derived from a Position Frequency Matrix) is log transformed. This parameter provides which "log transform" will be used. If TRUE, the Natural Log will be used (ln). If FALSE, log2 will be used. We strongly encourage to set values when building a `parameterOptions` object.

## Value

Returns the value assigned to the `naturalLog` slot in a `parameterOptions` object.

## Author(s)

Patrick C.N. Martin <p.martin@essex.ac.uk>

## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

## Examples

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions(naturalLog=TRUE)
#Setting new Value for naturalLog
naturalLog(GPP)
```

---

naturalLog-methods      *~~ Methods for Function naturalLog ~~*

---

## Description

Accessor method for the `naturalLog` slot in a `parameterOptions` object.

## Methods:

`naturalLog(object)`

---

naturalLog<-                      *Setter method for the naturalLog slot in a [parameterOptions](#) object.*

---

### Description

Setter method for the naturalLog slot in a [parameterOptions](#) object.

### Usage

```
naturalLog(object)<- value
```

### Arguments

object	object is a <a href="#">parameterOptions</a> object.
value	value is a logical value that will determine if the natural log or log2 should be used for the computation of the Position Weight Matrix.

### Details

During the computation of a Position Weight Matrix, the Position Probability Matrix (derived from a Position Frequency Matrix) is log transformed. This parameter provides which "log transform" will be used. If TRUE, the Natural Log will be used (ln). If FALSE, log2 will be used. We strongly encourage to set values when building a [parameterOptions](#) object.

### Value

Returns [parameterOptions](#) object with an updated value for the naturalLog slot.

### Author(s)

Patrick C.N. Martin <p.martin@essex.ac.uk>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

### Examples

```
# Loading data
data(ChIPanalyserData)

#Building data objects
OPP <- parameterOptions(naturalLog=TRUE)
#Setting new Value for naturalLog
naturalLog(OPP) <- FALSE
```

---

naturalLog<-methods     *~~ Methods for Function naturalLog<- ~~*

---

### Description

Setter method for the naturalLog slot in a [parameterOptions](#) object.

### Methods:

naturalLog(object)<-value

---

noiseFilter                     *Accessor Method for the noiseFilter slot in a [parameterOptions](#) object*

---

### Description

Accessor Method for the noiseFilter slot in a [parameterOptions](#) object

### Usage

noiseFilter(object)

### Arguments

object                     object is [parameterOptions](#) object

### Details

Noise filtering method that should be used on ChIP-seq data. Four methods are available: Zero, Mean, Median and Sigmoid. Zero removes all ChIP-seq scores below zero, mean under the mean score, median under median score and sigmoid assigns a weight to each score based on a logistic regression curve. Mid point is set at 95 95 quantile of ChIP-seq scores. Below midpoint will receive a score between 0 and 1 , everything above will receive a score between 1 and 2

### Value

Returns the value assigned to the noiseFilter slot in a [parameterOptions](#) object.

### Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions(noiseFilter="sigmoid")
#Setting new Value for noiseFilter
noiseFilter(GPP)
```

---

noiseFilter-methods    *~~ Methods for Function noiseFilter ~~*

---

**Description**

Accessor method for noiseFilter

**Methods:**

noiseFilter(object) Noise Filter that will be applied to ChIP scores

---

noiseFilter<-                    *Setter Method for the noiseFilter slot in a [parameterOptions](#) object*

---

**Description**

Setter Method for the noiseFilter slot in a [parameterOptions](#) object

**Usage**

```
noiseFilter(object) <- value
```

**Arguments**

object	object is <a href="#">parameterOptions</a> object
value	value is the value to be assigned to the noiseFilter slot (zero - mean - median -sigmoid)

**Details**

Noise filtering method that should be used on ChIP-seq data. Four methods are available: Zero, Mean, Median and Sigmoid. Zero removes all ChIP-seq scores bellow zero, mean under the mean score, median under median score and sigmoid assigns a weight to each score based on a logistic regression curve. Mid point is set at 95 95 quantile of ChIP-seq scores. Below midpoint will receive a score between 0 and 1 , everything above will receive a score between 1 and 2

**Value**

Returns the value assigned to the noiseFilter slot in a [parameterOptions](#) object.

**Author(s)**

Patrick C. N. Martin <p.martin@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions(noiseFilter="sigmoid")
#Setting new Value for noiseFilter
noiseFilter(GPP) <-"zero"
```

---

```
noiseFilter<-methods    ~~ Methods for Function noiseFilter<- ~~
```

---

**Description**

Setter method for noiseFilter

**Methods:**

noiseFilter(object)<-value Noise Filter that will be applied to CHIP scores

---

```
noOfSites              Accessor Method for the noOfSites slot in a parameterOptions ob-
                       ject
```

---

**Description**

Accessor Method for the noOfSites slot in a [parameterOptions](#) object

**Usage**

```
noOfSites(object)
```

**Arguments**

object                object is [parameterOptions](#) object

**Details**

While computing Position Weight Matricies (PWM) from Position Frequency Matricies (PFM), it is possible to restrict the number of sites that will be used to compute the PWM. The default is set at "all". In this case, all sites will be used to compute the PWM.



**Value**

Returns the value assigned to the noOfSites slot in a [parameterOptions](#) object.

**Author(s)**

Patrick C. N. Martin <p.martin@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions(noOfSites="all")
#Setting new Value for naturalLog
noOfSites(GPP)
```

---

noOfSites-methods      *~~ Methods for Function noOfSites ~~*

---

**Description**

*~~ Methods for function noOfSites ~~*

**Methods:**

signature(object = "parameterOptions")

---

noOfSites<-      *Setter Method for the noOfSites slot in a [parameterOptions](#) object.*

---

**Description**

Setter Method for the noOfSites slot in a [parameterOptions](#) object.

**Usage**

```
noOfSites(object) <- value
```

**Arguments**

object      object is a [parameterOptions](#) object.  
value      value is a positive integer that will be assigned to the noOfSites slot.

## Details

While computing Position Weight Matrices (PWM) from Position Frequency Matrices (PFM), it is possible to restrict the number of sites that will be used to compute the PWM. The default is set at "all". In this case, all sites will be used to compute the PWM.

## Value

Returns a `parameterOptions` object with an updated value for the `noOfSites` slot.

## Author(s)

Patrick C.N. Martin <p.martin@essex.ac.uk>

## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

## Examples

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions(noOfSites=0)
#Setting new Value for naturalLog
noOfSites(GPP) <- 8
```

---

noOfSites<-methods      *~~ Methods for Function noOfSites<- ~~*

---

## Description

Setter method for `noOfSites`

### Methods:

```
noOfSites(object)<-"all"
```

```
noOfSites(object)<-value
```

---

nos-class	<i>Class "nos"</i>
-----------	--------------------

---

**Description**

Virtual class to handle Number of Sites

**Objects from the Class**

A virtual Class: No objects may be created from it.

**Methods**

No methods defined with class "nos" in the signature.

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
showClass("nos")
```

---

parameterOptions	<i>parameter Options object</i>
------------------	---------------------------------

---

**Description**

parameterOptions is an object used to store and parse the various parameters needed throughout this analysis pipeline.

**Usage**

```
parameterOptions(ploidy = 2, boundMolecules = 1000, backgroundSignal = 0, maxSignal = 1, lociWidth =
```

**Arguments**

ploidy	<b>ploidy</b> : A numeric Value describing the ploidy of the organism. Default: 2
boundMolecules	<b>boundMolecules</b> : A vector (or single value) containing the number of bound Molecules (bound Transcription Factors): Default: 1000
backgroundSignal	<b>backgroundSignal</b> : A numeric value describing the CHIP-seq background Signal (average signal from real CHIP seq data). Default: 0
maxSignal	<b>maxSignal</b> : A numeric value describing the highest CHIP-seq signal (from real CHIP-seq data). Default: 1
lociWidth	<b>lociWidth</b> : A numeric value describing the width of the bins used to split CHIP profiles parsed to <code>processingCHIP</code> . Default = 20000
chipMean	<b>chipMean</b> : A numeric value describing the mean width of a CHIP- seq peak: Default:200
chipSd	<b>chipSd</b> : A numeric value describing the standard deviation of CHIP-seq peaks. Default: 200
chipSmooth	<b>chipSmooth</b> : A numeric value describing the width of the window used to smooth Occupancy profiles into CHIP profiles. Default:250
stepSize	<b>stepSize</b> : A numeric value describing the step Size (in base pairs) between each CHIP-seq score. Default:10 (Scored every 10 base pairs)
removeBackground	<b>removeBackground</b> : A numeric value describing the value at which score should be removed. Default:0 (If negative scores then remove)
noiseFilter	<b>noiseFilter</b> : A character string of one of the following: Zero, Mean, Median, or Sigmoid. Noise filter that will be applied to the CHIP Score during the <code>processingCHIP</code> step.
naturalLog	<b>naturalLog</b> : A logical value describing if natural Log will be used to compute the PWM (if FALSE then log2 will be used). Default: TRUE
noOfSites	<b>noOfSites</b> : A Positive integer descibing number of sites (in base pair) should be used from the PFM to compute PWM. Default =0 (Full width of binding site will be used when set to 0)
PWMThreshold	<b>PWMThreshold</b> : Threshold at which PWM Score should be selected (only sites above threshold will be selected - between 0 and 1)
strandRule	<b>strandRule</b> : 'mean', 'max' or 'sum' will dertermine how strand should be handle for computing PWM Scores. Default : 'max'
whichstrand	<b>whichstrand</b> : '+', '-' or '+-' on which strand should PWM Score be computed. Default: '+-'
PWMPseudocount	<b>PWMPseudocount</b> : A numeric value describing a PWMPseudocount for PWM computation. Default:1
lambdaPWM	A vector (or single value) contaning values for the ScalingFactorPWM (Also known as lambda).Default:1

**Details**

ChIPanalyser requires a lot of parameters. `parameterOptions` was created with the intent of storing and parsing these numerous arguments to the different functions. All parameters in this object are optional although strongly recommend. Some parameters are extracted and updated from function along the pipeline e.g. `maxSignal` and `backgroundSignal` are extracted during the `processingCHIP` step. These paramters will be automatically parsed. If you do not which to use them ( or any other parameter) simply parse a new `parameterOptions` object with your desired paramters.

**Value**

Returns a parameterOptions with updated values.

**Author(s)**

Patrick C. N. Martin <p.martin@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**See Also**

[genomicProfiles](#)

**Examples**

```
#
parameterOptions(ploidy = 2, boundMolecules = 1000, backgroundSignal = 0,
  maxSignal = 1, lociWidth = 20000, chipMean = 200, chipSd = 200,
  chipSmooth = 250, stepSize = 10, removeBackground = 0, noiseFilter = "zero",
  naturalLog = TRUE, noOfSites = "all", PWMThreshold = 0.7,
  strandRule = "max", whichstrand = "+-", PWMpseudocount = 1,
  lambdaPWM = 1)
```

---

parameterOptions-class

*Class "parameterOptions"*

---

**Description**

parameterOptions is an object used to store and parse the various parameters needed throughout this analysis pipeline.

**Objects from the Class**

Objects can be created by calls of the form parameterOptions(ploidy, boundMolecules, backgroundSignal, maxSignal, lociWidth, chipMean, chipSd, chipSmooth, stepSize, noiseFilter, removeBackground, lambdaPWM, PWMpseudocount, naturalLog, noOfSites, PWMThreshold, strandRule, whichstrand).

**Slots**

**ploidy:** Object of class "numeric": A numeric Value describing the ploidy of the organism. Default: 2

**boundMolecules:** Object of class "vector": A vector (or single value) containing the number of bound Molecules (bound Transcription Factors): Default: 1000

**backgroundSignal:** Object of class "numeric": A numeric value describing the ChIP-seq background Signal (average signal from real ChIP seq data). Default: 0

**maxSignal:** Object of class "numeric": A numeric value describing the highest ChIP-seq signal (from real ChIP-seq data). Default: 1

**lociWidth:** Object of class "numeric": A numeric value describing bin size when splitting ChIP seq scores). Default: 20 000

**chipMean:** Object of class "numeric": A numeric value describing the mean width of a ChIP- seq peak. Default:150

**chipSd:** Object of class "numeric": A numeric value describing the standard deviation of ChIP- seq peaks. Default: 150

**chipSmooth:** Object of class "vector": A numeric value describing the width of the window used to smooth Occupancy profiles into ChIP profiles. Default:250

**stepSize:** Object of class "numeric": A numeric value describing the step Size (in base pairs) between each ChIP-seq score. Default:10 (Scored every 10 base pairs)

**removeBackground:** Object of class "numeric": A numeric value describing the value at which score should be removed. Default:0 (If negative scores then remove)

**noiseFilter:** Object of class "character" Describes noiseFilter method applied to ChIP scores

**PWMThreshold:** Object of class "numeric": Threshold at which PWM Score should be selected (only sites above threshold will be selected - between 0 and 1)

**strandRule:** Object of class "character": "mean", "max" or "sum" will determine how strand should be handle for computing PWM Scores. Default : "max"

**whichstrand:** Object of class "character": "+", "-" or "+-" on which strand should PWM Score be computed. Default: "+-"

**lambdaPWM:** Object of class "vector" A vector (or single value) containing values for lambdaPWM Default:1

**naturalLog:** Object of class "logical": A logical value describing if natural Log will be used to compute the PWM (if FALSE then log2 will be used). Default: TRUE

**noOfSites:** Object of class "nos" A Positive integer describing number of sites (in base pair) should be used from the PFM to compute PWM. Default =0 (Full width of binding site will be used when set to 0)

**PWMPseudocount:** Object of class "numeric": A numeric value describing a PWMPseudocount for PWM computation. Default:1

**paramTag:** Object of class "character" ~Internal~

## Methods

**.paramTag** signature(object = "parameterOptions"): ...

**.paramTag<-** signature(object = "parameterOptions", value = "character"): ...

**.ZeroBackground** signature(object = "parameterOptions"): ...

**.ZeroBackground<-** signature(object = "parameterOptions", value = "vector"): ...

**backgroundSignal** signature(object = "parameterOptions"): ...

**backgroundSignal<-** signature(object = "parameterOptions", value = "numeric"): ...

**boundMolecules** signature(object = "parameterOptions"): ...

**boundMolecules<-** signature(object = "parameterOptions", value = "vector"): ...

**chipMean** signature(object = "parameterOptions"): ...

**chipMean<-** signature(object = "parameterOptions", value = "numeric"): ...

**chipSd** signature(object = "parameterOptions"): ...

**chipSd**<- signature(object = "parameterOptions", value = "numeric"): ...  
**chipSmooth** signature(object = "parameterOptions"): ...  
**chipSmooth**<- signature(object = "parameterOptions", value = "vector"): ...  
**initialize** signature(.Object = "parameterOptions"): ...  
**lambdaPWM** signature(object = "parameterOptions"): ...  
**lambdaPWM**<- signature(object = "parameterOptions", value = "vector"): ...  
**lociWidth** signature(object = "parameterOptions"): ...  
**lociWidth**<- signature(object = "parameterOptions", value = "numeric"): ...  
**maxSignal** signature(object = "parameterOptions"): ...  
**maxSignal**<- signature(object = "parameterOptions", value = "numeric"): ...  
**naturalLog** signature(object = "parameterOptions"): ...  
**naturalLog**<- signature(object = "parameterOptions", value = "logical"): ...  
**noiseFilter** signature(object = "parameterOptions"): ...  
**noiseFilter**<- signature(object = "parameterOptions", value = "character"): ...  
**noOfSites** signature(object = "parameterOptions"): ...  
**noOfSites**<- signature(object = "parameterOptions", value = "character"): ...  
**noOfSites**<- signature(object = "parameterOptions", value = "numeric"): ...  
**ploidy** signature(object = "parameterOptions"): ...  
**ploidy**<- signature(object = "parameterOptions", value = "numeric"): ...  
**PWMpseudocount** signature(object = "parameterOptions"): ...  
**PWMpseudocount**<- signature(object = "parameterOptions", value = "numeric"): ...  
**PWMThreshold** signature(object = "parameterOptions"): ...  
**PWMThreshold**<- signature(object = "parameterOptions", value = "numeric"): ...  
**removeBackground** signature(object = "parameterOptions"): ...  
**removeBackground**<- signature(object = "parameterOptions", value = "vector"): ...  
**show** signature(object = "parameterOptions"): ...  
**stepSize** signature(object = "parameterOptions"): ...  
**stepSize**<- signature(object = "parameterOptions", value = "numeric"): ...  
**strandRule** signature(object = "parameterOptions"): ...  
**strandRule**<- signature(object = "parameterOptions", value = "character"): ...  
**whichstrand** signature(object = "parameterOptions"): ...  
**whichstrand**<- signature(object = "parameterOptions", value = "character"): ...

### Author(s)

Partick C. N. Martin <p.martin@essex.ac.uk>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**See Also**

[genomicProfiles](#)

**Examples**

```
showClass("parameterOptions")
```

---

PFMFormat

*Accessor method for the PFMFormat slot in a [genomicProfiles](#) object*

---

**Description**

Accessor method for the PFMFormat slot in a [genomicProfiles](#) object

**Usage**

```
PFMFormat(object)
```

**Arguments**

object            object is a [genomicProfiles](#) object

**Details**

If loading a [PositionFrequencyMatrix](#) from a file, the format of the file should be specified. Default is raw. Please keep in mind that this argument is used when parsing the [PositionFrequencyMatrix](#) file. IF this argument is changed after building the [genomicProfiles](#) with a PositionFrequency-Matrix file, this will not influence the parsing of the file. PFMFormat can be one of the following: "raw", "transfac", "JASPAR" or "sequences"

**Value**

Returns the value assigned to the PFMFormat slot a [genomicProfiles](#)

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.



**Examples**

```
# Loading data
data(ChIPAnalyserData)
#Loading PFM files
PFM <- file.path(system.file("extdata", package="ChIPAnalyser"), "BEAF-32.pfm")
#Building data objects
#### THIS IS THE PREFERRED METHOD FOR SETTING PFMFormat
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR")
#Setting New value for PFMFormat
PFMFormat(GPP)
```

---

PFMFormat-methods      *~~ Methods for Function PFMFormat ~~*

---

**Description**

Accesor method for the PFMFormat slot in a [genomicProfiles](#) object

**Methods:**

PFMFormat(object)

---

PFMFormat<-                      *Setter method for the PFMFormat slot in a [genomicProfiles](#) object*

---

**Description**

Setter method for the PFMFormat slot in a [genomicProfiles](#) object

**Usage**

PFMFormat(object) <- value

**Arguments**

object	object is a <a href="#">genomicProfiles</a> object
value	value is character string of one of the following: "raw","transfac","JASPAR" or "sequences". If loading a <a href="#">PositionFrequencyMatrix</a> from a file, the format of the file should specified. Default is JASPAR.

**Details**

If loading a [PositionFrequencyMatrix](#) from a file, the format of the file should be specified. Default is JASPAR. Please keep in mind that this argument is used when parsing the [PositionFrequencyMatrix](#) file. IF this argument is changed after building the [genomicProfiles](#) with a [PositionFrequencyMatrix](#) file, this will not influence the parsing of the file.

**Value**

Returns a [genomicProfiles](#) object with an updated value for the PFMFormat slot.

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
# Loading data
data(ChIPanalyserData)
#Loading PFM files
PFM <- file.path(system.file("extdata",package="ChIPanalyser"),"BEAF-32.pfm")
#Building data objects
#### THIS IS THE PREFERRED METHOD FOR SETTING PFMFormat
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR")
#Setting New value for PFMFormat
PFMFormat(GPP) <- "JASPAR"
```

---

PFMFormat<-methods      *~~ Methods for Function PFMFormat<- ~~*

---

**Description**

Setter method for the PFMFormat slot in a [genomicProfiles](#) object

**Methods:**

PFMFormat(object)<-value

---

ploidy                      *Accessor method for the ploidy slot in a [parameterOptions](#) object*

---

**Description**

Accessor method for the ploidy slot in a [parameterOptions](#) object

**Usage**

ploidy(object)

**Arguments**

object                      object is a [parameterOptions](#) object

**Details**

Default value for ploidy is set a 2. It should be mentioned that ChIPanalyser is based on a model that also considers the ploidy of the organism of interest however this only considers simple poly-ploidy (or haploidy). The model does not consider hybrids such as wheat.

**Value**

Returns the value assigned to the ploidy slot in a [parameterOptions](#) object

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for maxSignal
ploidy(OPP)
```

---

ploidy-methods                      *~~ Methods for Function ploidy ~~*

---

**Description**

Accessor method for the ploidy slot in a [parameterOptions](#) object

**Methods:**

ploidy(object)

---

ploidy<-                              *Setter Method for the ploidy slot in an [parameterOptions](#) object*

---

**Description**

Setter Method for the ploidy slot in an [parameterOptions](#) object

**Usage**

```
ploidy(object)<- value
```

**Arguments**

object                      object is a [parameterOptions](#) object  
value                        value is a positive integer that describes the ploidy of the organism of interest.

**Details**

Default value for ploidy is set a 2. It should be mentioned that ChIPanalyser is based on a model that also considers the ploidy of the organism however this only considers simple polyploidy (or haploidy). The model does not consider hybrids such as wheat.

**Value**

Returns a `parameterOptions` object with an updated value for the ploidy slot.

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for maxSignal
ploidy(OPP) <- 2
```

---

ploidy<-methods      *~~ Methods for Function ploidy<- ~~*

---

**Description**

Setter Method for the ploidy slot in an `parameterOptions` object

**Methods:**

ploidy(object)<-value

---

plotOccupancyProfile    *Plot Occupancy Profiles*

---

**Description**

plotOccupancyProfile plots the predicted profiles. If provided, this functions will also plot ChIP-seq profiles, PWMScores (or Occupancy), chromatin States, Goodness of Fit estimates and gene information.

**Usage**

```
plotOccupancyProfile(predictedProfile, ChIPScore = NULL, chromatinState = NULL,
  occupancy = NULL, goodnessOfFit = NULL, PWM=FALSE,
  geneRef = NULL, addLegend = TRUE, ...)
```

**Arguments**

predictedProfile	predictedProfile is a either <a href="#">GRanges</a> containing the predicted profiles for one loci, all loci selected for one paramter, or all loci selected for all parameter combinations selected. (see <a href="#">searchSites</a> )
ChIPScore	ChIPScore is a ChIPscore object containing ChIPscore (or a list of numeric values representing ChIP scores (Experimental ChIP))
chromatinState	chromatinState is a <a href="#">GRanges</a> containing accesible DNA sites or chromatin States.
occupancy	occupancy is a <a href="#">GRanges</a> or a genomicProfiles object contaning PWM scores and Occupancy ( see computeOccupancy)
goodnessOfFit	goodnessOfFit results of the <a href="#">profileAccuracyEstimate</a> function.
PWM	PWM is a logical value that in the case occupancy is provided which of occupancy scores of PWM scores hould be plotted. Default set at FALSE
geneRef	geneRef is a <a href="#">GRanges</a> containing gene information on exons,introns, UTR's, enhancers or any other genetic element to be plotted.
addLegend	addLegend is a logical value defining if the legend should be added. The legend will add all elements provided. See details.
...	Any other graphical Parameter of the following : cex, cex.lab, cex.main, densityCS , densityGR , ylab, xlab, main, colPred, colChIP, colOccup, colCS, colGR, n_axis_ticks. See details.

**Details**

Once the predicted ChIP-seq like profiles have been computed, it is possible to plot these profiles.

This functions allows to control graphical parameters. In short:

\* col = color values - exact number of colors or colors that will be used in a colorRampPalette.

\* cex = font sizes - for text, axis labels and main

\* Density = fill density for chromatin state and/or geneRef blocks

Pred = predictedProfile ChIP = ChIP score (Experimental ChIP data) CS = Chromatin States GR = Gene reference Occup = Occupnacy locations

**Value**

Returns a profile plot with "Occupancy" on the y axis and DNA position on the the X- axis.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```

#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR", BPFrequency=DNASequenceSet)

# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GPP)

#Compute PWM Scores
PWMScores <- computePWMScore(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GenomeWide,
  loci = top, chromatinState = Access)
#Compute Occupancy
Occupancy <- computeOccupancy(genomicProfiles = PWMScores)

#Compute ChIP profiles
chipProfile <- computeChipProfile(loci = top,
  genomicProfiles = Occupancy)

#Plotting Profile
plotOccupancyProfile(predictedProfile=chipProfile,
  ChIPScore = chip,
  chromatinState = Access,
  occupancy = Occupancy,
  geneRef =geneRef)

plotOccupancyProfile(predictedProfile=chipProfile,
  ChIPScore = chip,
  chromatinState = Access,
  occupancy = Occupancy,
  geneRef = geneRef,
  colCS = c("red","blue"),
  densityGR = 60)

```

## Description

plotOptimalHeatMaps will plot heat maps of optimal Parameters and highlight the optimal combination of [lambdaPWM](#) and [boundMolecules](#)

## Usage

```
plotOptimalHeatMaps(optimalParam, contour=TRUE, col=NULL, main=NULL, layout=TRUE, overlay=FALSE)
```

## Arguments

optimalParam	optimalParam is a list containing containing optimal matrices (or only one if only one parameter was selected). These matrices are the result of the computeOptimal function
contour	parameter is logical. Should contour lines be plotted?
col	col vector of colours to be used for each heat map. If none are specified, rainbow colours will be used. NOTE: colour vector will be recycled if not enough colours are provided.
main	main title.
layout	layout is either TRUE or FALSE specifying if standard layout should be used or not. If TRUE, each heat map will be plotted on an individual page with a heat map scale of the right side.
overlay	overlay is either TRUE or FALSE specifying if an overlay plot should be produced. The overlay plot takes the top 10 percent of best performing parameters per scoring metric and overlays them in a single plot. The resulting plots shows the optimal set of parameters for all metrics combined.

## Details

Once the optimal set of Parameters ( [lambdaPWM](#) and [boundMolecules](#) ), it is possible to plot the results in the form of a heat map. Each heat map will be plotted in a separate page if layout = TRUE, If layout= FALSE, it is up to the user to define how they wish to layout their heat maps.

## Value

Returns a heat map of optimal combinations of [lambdaPWM](#) and [boundMolecules](#). The x axis represents the different value assigned to lambda ( [lambdaPWM](#) ) and the y axis represents the different values to boundMolecules ( [boundMolecules](#) ).

## Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```

#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)

#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR",BPFrequency=DNASequenceSet)

#Computing Optimal set of Parameters
optimalParam <- computeOptimal(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet,
  ChIPScore = chip,
  chromatinState = Access,
  parameterOptions = OPP,
  parameter = "all",
  peakMethod="moving_kernel")
plotOptimalHeatMaps(optimalParam)

```

---

PositionFrequencyMatrix

*Accessor method for the PFM slot in a genomicProfiles object*

---

**Description**

Accessor method for the PFM slot in a genomicProfiles object

**Usage**

```
PositionFrequencyMatrix(object)
```

**Arguments**

object                    object is a [genomicProfiles](#) object

**Details**

After creating a [genomicProfiles](#) object, it is possible to access the Position Frequency Matrix slot. However this slot will be empty if the [genomicProfiles](#) object was built using directly a Position Weight Matrix. See [genomicProfiles](#)



**Value**

Returns the Position Frequency Matrix (PFM slot) used to compute the [PositionWeightMatrix](#) in a [genomicProfiles](#) object

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
#Loading data
data(ChIPanalyserData)
#Loading PFM files
PFM <- file.path(system.file("extdata", package="ChIPanalyser"), "BEAF-32.pfm")
#Building genomicProfiles object
GPP<-genomicProfiles(PFM=PFM,PFMFormat="JASPAR")
# Accessing Slot
PositionFrequencyMatrix(GPP)
```

---

PositionFrequencyMatrix-methods

~~ *Methods for Function* PositionFrequencyMatrix ~~

---

**Description**

Accessor method for the PFM slot in a [genomicProfiles](#) object

**Methods:**

PositionFrequencyMatrix(object)

---

PositionFrequencyMatrix<-

*Setter method for the PFM slot in a [genomicProfiles](#) object*

---

**Description**

Setter method for the PFM slot in a [genomicProfiles](#) object

**Usage**

PositionFrequencyMatrix(object)<- value

**Arguments**

object	object is a <a href="#">genomicProfiles</a> object
value	value can be of two forms. Either a matrix in the form of a Position Frequency Matrix or a path/to/file character string.

**Details**

The Position Frequency Matrix is one of the fundamental object that needs to be supplied to a [genomicProfiles](#). If after building a [genomicProfiles](#), only the Position Frequency Matrix needs to be modified then it is possible to manually update the value of this matrix using the function above. There are two options for the type of data that may be supplied to the PFM slot: a matrix in the form of a Position Frequency Matrix (matrix with four rows - one for each base pair (ACTG) and a number of columns equal to the number of sites in the binding site), or it is possible (also recommended) to provide a path to the file containing the Position Frequency Matrix. This Position Frequency Matrix file may come in multiple form such as RAW, Transfac or JASPAR. WARNING: if a [genomicProfiles](#) object has already been created and only the PFM is supplied/updated, then the Positon Weight Matrix will automatically updated as well.

**Value**

Returns a [genomicProfiles](#) with an updated PFM slot (as described above this will lead to an updated PositionWeightMatrix).

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
#Loading data
data(ChIPanalyserData)
#Loading PFM files
PFM <- file.path(system.file("extdata", package="ChIPanalyser"), "BEAF-32.pfm")
#Building genomicProfiles object
# NOT ADVISED!!!! PLEASE PARSE PFM AND PFMFormat together
GPP<-genomicProfiles(PFMFormat = "JASPAR")
#Setting PFM
PositionFrequencyMatrix(GPP) <- PFM
```

---

PositionFrequencyMatrix<-methods

~~ *Methods for Function* PositionFrequencyMatrix<- ~~

---

**Description**

Setter method for the PFM slot in a [genomicProfiles](#) object

**Methods:**

```
PositionFrequencyMatrix(object)<-"path/to/file/"  
PositionFrequencyMatrix(object)<-value
```

---

PositionWeightMatrix *Accessor Method for the PWM slot in a [genomicProfiles](#) object*

---

**Description**

Accessor Method for the PWM slot in a [genomicProfiles](#) object

**Usage**

```
PositionWeightMatrix(object)
```

**Arguments**

object                    object is a [genomicProfiles](#)

**Details**

After creating a [genomicProfiles](#) object, it is possible to access the Position Weight Matrix stored in this slot. This slot should always contain something. This slot is either supplied by user or directly computed from a Position Frequency Matrix when supplied.

**Value**

Returns a matrix in the form of a Position Weight Matrix

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
#Loading data  
data(ChIPanalyserData)  
#Loading PFM files  
PFM <- file.path(system.file("extdata", package="ChIPAnalyser"), "BEAF-32.pfm")  
#Building genomicProfiles object  
GPP<-genomicProfiles(PFM=PFM,PFMFormat="JASPAR")  
# Accessing Slot  
PositionWeightMatrix(GPP)
```

---

 PositionWeightMatrix-methods

 ~~ Methods for Function PositionWeightMatrix ~~
 

---

### Description

Accessor Method for the PWM slot in a [genomicProfiles](#) object

### Methods:

PositionWeightMatrix(object)

---

PositionWeightMatrix&lt;-

*Setter Method for the PositionWeightMatrix slot in a [genomicProfiles](#) object*

---

### Description

Setter Method for the PositionWeightMatrix slot in a [genomicProfiles](#) object

### Usage

PositionWeightMatrix(object) <- value

### Arguments

object	object is a <a href="#">genomicProfiles</a> object
value	value is a matrix in the form of a Position Weight Matrix.

### Details

If a Position Weight Matrix is readily available, it is possible to directly assign this matrix to the PWM slot. However, this is only possible if a [genomicProfiles](#) object has already been created. In that case, we advise to first create a [genomicProfiles](#) object. It should be noted that this Position Weight Matrix will be automatically computed from a Position Frequency Matrix. If no Position Frequency Matrix are available, then a Position Weight Matrix can be directly assigned to this slot.

### Value

Returns a [genomicProfiles](#) object with an updated value for the PWM slot

### Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
#Building genomicProfiles object
GPP <- genomicProfiles()
#Setting PWM to PositionWeightMatrix slot
PWM <- matrix(runif(32,-10,20), ncol=8)
rownames(PWM) <- c("A","C","T","G")
PositionWeightMatrix(GPP) <- PWM
```

---

PositionWeightMatrix<-methods

~~ Methods for Function PositionWeightMatrix<- ~~

---

**Description**

Setter Method for the PositionWeightMatrix slot in a [genomicProfiles](#) object

**Methods:**

PositionWeightMatrix(object)<-value

---

processingChIP

*Pre-processing ChIP-seq data*

---

**Description**

processingChIP will process and extract ChIP scores at a set of loci of interest.

**Usage**

```
processingChIP(profile, loci=NULL, reduce=NULL,
               peaks=NULL, chromatinState=NULL, parameterOptions=NULL,
               cores=1)
```

**Arguments**

profile	profile is a path to a UCSC format file, a GRanges or data frame. The input data should contain 4 columns: chromosome, start, end and score.
loci	loci is <a href="#">GRanges</a> describing the loci at which ChIP scores should be extracted. If NULL, a set of Loci will be extracted from profile. The data provided will then be split into bins of width equal to lociWidth (Default 20kbp) Default=NULL
reduce	reduce is a the top regions to select based on the mean ChIP score. If peaks are provided, regions overlapping with known peaks will be selected based on highest ChIP score. If NULL, all regions will be considered. Default=NULL
parameterOptions	parameterOptions is an <a href="#">parameterOptions</a> object containing chip Parameters to be parsed for ChIP score extraction. If NULL, <a href="#">parameterOptions</a> will be built internally with default ChIP extraction parameters (see <a href="#">chipSmooth</a> , <a href="#">chipSd</a> and <a href="#">chipMean</a> ) Default=NULL

peaks	peaks is a path to UCSC format file or a GRanges object containing location of ChIP peaks. Default=NULL
chromatinState	chromatinState is a GRanges containing Accessible DNA or chromatin States. If provided, regions will be selected only if they contain accessible DNA. Default=NULL
cores	cores is the number of cores used to extract ChIP scores. Default = 1

### Details

When using `computeOptimal`, it is required to supply real ChIP data in order to have a point of comparison. The correlation and MSE Scores are computed based on how well the model fits biological data. `processingChIP` will extract this data from ChIP data at loci of interest. When using the `reduce` option, this function will only select the top regions based on peak height or mean ChIP score. `processingChIP` will also extract `maxSignal` and `backgroundSignal` from ChIP data and parse it to a `parameterOptions` object.

### Value

Returns a `ChIPScore` object containing extracted (and normalised) ChIP scores, the loci of interest and newly extracted Parameters (e.g. `maxSignal`)

### Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

### Examples

```
#Data extraction
data(ChIPAnalyserData)

## Extracting ChIP scores at loci of interest

ChIP<-processingChIP(profile=chip, loci=top)
```

---

profileAccuracyEstimate

*Estimating Accuracy of predicted Profiles*

---

### Description

`profileAccuracyEstimate` will compare the predicted ChIP-seq-like profile to real ChIP-seq data and return a set of metrics describing how accurate the predicted model is compared to real data.

**Usage**

```
profileAccuracyEstimate(genomicProfiles,ChIPScore,
  parameterOptions=NULL,method="all",cores=1)
```

**Arguments**

`genomicProfiles`  
`genomicProfiles` is the result of [computeChIPProfile](#)

`ChIPScore`  
`ChIPScore` is the result of processing ChIP. Extracted/Normalised experimental ChIP scores.

`parameterOptions`  
`parameterOptions` is a [parameterOptions](#) object for parameter specification.

`method`  
`method` is the method that will be used to assess model quality against ChIP-seq data. Method can be one of the following: pearson, spearman, kendall, ks, geometric.fscore, MSE, or all.Fscore contains f-score, precision, recall, MCC, Accuracy and AUC ROC.

`cores`  
`cores` is the number of cores used to extract ChIP scores. Default = 1

**Details**

In order to assess the quality of the model against experimental ChIP-seq data, ChIPanalyzer offers a wide range of method to choose from. These methods are also used when computing optimal parameters.

**Value**

Returns list of goodness of fit metrics for each loci and each parameter selected.

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
#Data extraction
data(ChIPanalyzerData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPanalyzer"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
```

```

DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR", BPFrequency=DNASequenceSet)

# Computing Genome Wide
GenomeWide <- computeGenomeWideScore(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet)

#Compute PWM Scores
PWMScores <- computePWMScore(genomicProfiles = GenomeWide,
  DNASequenceSet = DNASequenceSet, loci = top, chromatinState = Access)
#Compute Occupancy
Occupancy <- computeOccupancy(genomicProfiles = PWMScores)

#Compute ChIP profiles
chipProfile <- computeChIPProfile(genomicProfiles=Occupancy,loci=top)
#Estimating accuracy estimate
AccuracyEstimate <- profileAccuracyEstimate(genomicProfiles = chipProfile,
  ChIPScore = chip,
  occupancyProfileParameters = OPP)

```

---

profiles-methods

~~ *Methods for Function profiles* ~~

---

### Description

Accessor method for profiles in a [genomicProfiles](#) object

### Methods:

profiles(object) Computed PWM scores, Occupancy or ChIP-seq like profiles for loci of interest and paramter combination of interest.

---

PWMpseudocount

*Accessor Method for a PWMpseudocount slot in a [parameterOptions](#)*

---

### Description

Accessor Method for a PWMpseudocount slot in a [parameterOptions](#)

### Usage

PWMpseudocount(object)

### Arguments

object            object is a [parameterOptions](#) object.



## Details

In the context of Position Weight Matrices, the pseudocount is used to avoid 0 probabilities during the transformation of Position Frequency Matrix to a Position Probability Matrix and finally to a Position Weight Matrix. It is essentially a sample correction that is added in the case of small sample size. The effect of the base pair to which a pseudocount was assigned will not influence the model nor will create mathematical issues such as infinities or zero division. Default is set at 1.

## Value

Returns the value assigned to a PWMpseudocount slot in a [parameterOptions](#) object

## Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

## Examples

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions(PWMpseudocount=0)
#Accessing slot value
PWMpseudocount(GPP)
```

---

PWMpseudocount-methods

~~ *Methods for Function PWMpseudocount* ~~

---

## Description

Accessor Method for a PWMpseudocount slot in a [parameterOptions](#)

## Methods:

PWMpseudocount(object)

---

PWMpseudocount<-      *Setter Method for the pseudocount slot in a [parameterOptions](#) object*

---

## Description

Setter Method for the pseudocount slot in a [parameterOptions](#) object

## Usage

```
PWMpseudocount(object) <- value
```

## Arguments

object	object is a <a href="#">parameterOptions</a> object
value	value is a numeric value that will be assigned to the pseudocount slot. Default is set at 1

## Details

In the context of Position Weight Matricies, the pseudocount is used to avoid 0 probabilities during the transformation of Position Frequency Matrix to a Position Probability Matrix and finally to a Position Weight Matrix. It is essentially a sample correction that is added in the case of small sample size. The effect of the base pair to which a pseudocount was assigned will not influence the model nor will create mathematical issues such as infinities or zero division.

## Value

Returns a [parameterOptions](#) object with an updated value for the pseudocount slot.

## Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

## Examples

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions( PWMpseudocount=0)
#Setting Value for new PWMpseudocount
PWMpseudocount(GPP) <- 1
```

---

 PWMpseudocount<-methods

 ~~ *Methods for Function PWMpseudocount<-* ~~
 

---

**Description**

Setter Method for the pseudocount slot in a [parameterOptions](#) object

**Methods:**

PWMpseudocount(object)<-value

---

PWMThreshold

*Accessor method for the PWMThreshold slot in a [parameterOptions](#) object*

---

**Description**

Accessor method for the PWMThreshold slot in a [parameterOptions](#) object

**Usage**

PWMThreshold(object)

**Arguments**

object            object is a [parameterOptions](#) object

**Details**

The computePWMScore function requires a so-called PWM Threshold. This threshold represents the Threshold at which PWM Score should be selected. The PWMThreshold is a positive numeric value (between 0 and 1. If set at 0, all sites will be selected. If set at 0.7 (Default value), then 70 % of PWM Score (and by extension binding sites) will be IGNORED. The top 30 % will be selected.

**Value**

Returns the value assigned to the PWMThreshold slot in a [parameterOptions](#) object

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions(PWMThreshold=0.7)
#Accessing Value for PWMThreshold
PWMThreshold(GPP)
```

---

PWMThreshold-methods    *~~ Methods for Function PWMThreshold ~~*

---

**Description**

Accessor method for the PWMThreshold slot in a [parameterOptions](#) object

**Methods:**

PWMThreshold(object)

---

PWMThreshold<-                    *Setter Method for the PWMThreshold slot in a [parameterOptions](#) object*

---

**Description**

Setter Method for the PWMThreshold slot in a [parameterOptions](#) object

**Usage**

```
PWMThreshold(object) <- value
```

**Arguments**

object	object is a <a href="#">parameterOptions</a> object
value	value is a numeric value (between 0 and 1) to be assigned to the PWMThreshold slot in <a href="#">parameterOptions</a> object. Default is set at 0.7

**Details**

The computePWMScore function requires a so-called PWM Threshold. This threshold represents the Threshold at which PWM Score should be selected. The PWMThreshold is a positive numeric value (between 0 and 1). If set at 0, all sites will be selected. If set at 0.7 (Default value), then 70 % of PWM Score (and by extension binding sites) will be IGNORED. The top 30 % will be selected.

**Value**

Returns [parameterOptions](#) object with an updated value for the PWMThreshold slot

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions(PWMThreshold=0.7)
#Setting Value for new PWMThreshold
PWMThreshold(GPP) <- 0.8
```

---

PWMThreshold<-methods ~ Methods for Function PWMThreshold<- ~

---

**Description**

Setter Method for the PWMThreshold slot in a [parameterOptions](#) object

**Methods:**

PWMThreshold(object)<-value

---

removeBackground      *Accessor Method for the removeBackground slot in a*  
[parameterOptions](#) *object*

---

**Description**

Accessor Method for the removeBackground slot in a [parameterOptions](#) object

**Usage**

```
removeBackground(object)
```

**Arguments**

object                  object is a [parameterOptions](#) object

**Details**

A numeric value describing a threshold at which Occupancy signals must be removed (Default is set at 0). The removal of Occupancy signals will occur when computing [computeOccupancy](#) (see [computeOccupancy](#) function)

**Value**

Returns the value assigned to the removeBackground slot in a [parameterOptions](#) object

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
#Building parameterOptions object
OPP <- parameterOptions()
#Accessing Value for removeBackground
removeBackground(OPP)
```

---

removeBackground-methods

~~ *Methods for Function removeBackground* ~~

---

**Description**

Accessor Method for the removeBackground slot in a [parameterOptions](#) object

**Methods:**

removeBackground(object)

---

removeBackground<-     *Setter Method for the removeBackground slot in a parameterOptions object*

---

**Description**

Setter Method for the removeBackground slot in a [parameterOptions](#) object

**Usage**

```
removeBackground(object) <-value
```

**Arguments**

object                object is an [parameterOptions](#) object  
value                 value is positive numerical value to be assigned to the removeBackground slot in a [parameterOptions](#) object. Default is set a 0.

**Details**

A numeric value describing a threshold at which Occupancy signals must be removed (Default is set at 0). The removal of Occupancy signals will occur when computing `computeOccupancy` (see `computeOccupancy` function)

**Value**

Returns an `parameterOptions` object with an updated value for the `removeBackground` slot

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
#Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for removeBackground
removeBackground(OPP) <- 0.1
```

---

removeBackground<-methods

~~ *Methods for Function* removeBackground<- ~~

---

**Description**

Setter Method for the `removeBackground` slot in a `parameterOptions` object

**Methods:**

`removeBackground(object)<-value`

---

scores *Accessor Method for the scores slot in a [ChIPScore](#) object*

---

**Description**

Setter Method for the scores slot in a [ChIPScore](#) object

**Usage**

```
scores(object)
```

**Arguments**

object            object is [ChIPScore](#) object

**Details**

When using the [processingChIP](#), this functions will return a name list of normalised ChIP scores at loci of interest. This functions enables you to extract those scores from the [ChIPScore](#) object.

**Value**

Returns the value assigned to the scores slot in a [ChIPScore](#) object.

**Author(s)**

Patrick C. N. Martin <p.martin@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

**Examples**

```
# Loading data
data(ChIPanalyserData)

chip<-processingChIP(chip,top)
str(scores(chip))
```

---

scores-methods            *~~ Methods for Function scores ~~*

---

**Description**

Accessor method for scores slot in a [ChIPScore](#) object.

**Methods:**

scores(object)    Extracted and normalised ChIP scores at loci of interest.



---

searchSites	<i>Searching function for Sites above threshold and predicted ChIP-seq Profiles</i>
-------------	---

---

### Description

searchSites is function enabling quick extraction and search for parameter combinations and/or loci in any [genomicProfiles](#) object from computeOccupancy onwards.

### Usage

```
searchSites(Sites,lambdaPWM="all",BoundMolecules="all", Locus="all")
```

### Arguments

Sites	Sites is either a <a href="#">genomicProfiles</a> or the result of computeOptimal
lambdaPWM	lambdaPWM is a numeric vector describing the ScalingFactors that should be searched within Sites.
BoundMolecules	BoundMolecules is a numeric vector describing the BoundMolecules that should be searched within Sites.
Locus	Locus is a character vector describing the Loci that should be searched within Sites.

### Details

When testing numerous combinations of lambdaPWM and boundMolecules on top of many loci, it can become challenging to navigate the large data output searchSites will make searching in this slot a lot easier. If all arguments are left at their default value of "all", then all Parameters will be searched thus returning the full list of Sites above threshold. If a value for lambdaPWM is user provided then only this lambdaPWM will be selected (all boundMolecules and loci will also be selected). searchSites also works on the result of computeOptimal.

### Value

Returns object of same time as parsed to this function with only the parameters and/or loci selected.

### Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```

#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR", BPFfrequency=DNASequenceSet)

# Computing Genome Wide
GenomeWide <- computeGenomeWideScore(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet)

#Compute PWM Scores
PWMScores <- computePWMScore(genomicProfiles = GenomeWide,
  DNASequenceSet = DNASequenceSet, loci = top, chromatinState = Access)
#Compute Occupancy
Occupancy <- computeOccupancy(genomicProfiles = PWMScores)
searchSites(Occupancy,ScalingFactor=c(1,4), BoundMolecules = c(1,100),
  Locus="eve")

#Compute ChIP profiles
chipProfile <- computeChIPProfile(genomicProfiles=Occupancy,loci=top)
searchSites(chipProfile,ScalingFactor=c(1,4), BoundMolecules = c(1,100),
  Locus="eve")

optimalParam <- computeOptimal(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet,
  ChIPScore = chip,
  chromatinState = Access,
  parameterOptions = OPP,
  parameter = "all",
  peakMethod="moving_kernel")

searchSites(optimalParam,ScalingFactor=c(1,4), BoundMolecules = c(1,100),
  Locus="eve")

```

---

setChromatinStates      *setChromatinStates*

---

**Description**

setChromatinStates sets chromatin state affinity values to a GRanges object.

**Usage**

```
setChromatinStates(population,chromatinStates)
```

**Arguments**

`population` Population list containing all individuals and associated parameter. Must contain chromatin state affinity values. See `generateStartingPopulation`.

`chromatinStates` GRanges object containing chromatin state locations.

**Details**

Chromatin states can be loaded into R as a GRanges object. Each range represents the extent of a certain chromatin state and the chromatin state type should be assigned to a meta data column called "name". The affinity values names should be set accordingly.

**Value**

Returns a GRanges object with affinity scores for each chromatin state range. Affinity scores are placed in the DNAAffinity meta data column.

**Author(s)**

Patrick C.N. Martin

**Examples**

```
library(ChIPAnalyser)
# Input data
data(ChIPAnalyserData)

pop <- 10
params <- c("N","lambda","PWMThreshold", paste0("CS",seq(1:11)))
start_pop <- generateStartingPopulation(pop, params)

cs <- setChromatinStates(start_pop,cs)
```

---

show-methods

~~ *Methods for Function show* ~~

---

**Description**

Show methods for various objects

**Methods:**

```
signature(object = "ChIPScore")
signature(object = "genomicProfiles")
signature(object = "parameterOptions")
```

singleRun

*singleRun***Description**

singleRun runs ChIPanalyser after optimal paramters have been found by the evolve function.

**Usage**

```
singleRun(indiv,DNAAffinity,
          genomicProfiles,DNASequenceSet,
          ChIPScore,fitness="all")
```

**Arguments**

indiv	Population list containing the top scoring individual. Note that this should be a list of length 1 containing another list with all parameter values.
DNAAffinity	GRanges object as outputed by the setchromatinStates.
genomicProfiles	genomicProfiles object containing PWM scores and other desired metrics. Note that PWMThreshold, lambda and N will be overwritten using values from indiv.
DNASequenceSet	DNA string set object containing DNA sequence of interest.
ChIPScore	ChIPScore object as outputed by the processingChIP function.
fitness	character string describing which metric should be used to assess fitness and should be one of the following:"geometric","ks","MSE","pearson","spearman","kendall","recall","precesion","fscore","MCC","Accuracy" or "AUC".

**Details**

Once the genetic algorithm has been optimised, the top individual may be run on its own to get predicted ChIP profiles. The use of this function requires a few extract steps in order to predict ChIP profiles.

First, the index of the top individual should be extracted (see getHighestFitnessSolutions). Second, using this index, subset top individual from GA population. Note this should be done using "[" single bracket notation as, a list of length 1 containing another list with all parameter values is required for the next steps. Yes, this is might seem annoying but the functions were design for list structures... Third, setchromatinStates using the top individual list. This will add chromatin affinity values to your chromatinState GRanges. Use this new chromatinState object as your new chromatinState object. Fourth, parse your indiv list object to singleRun.

**Value**

Return a list with three elements. First element contains a genomicProfiles object with occupancy scores. Second element contains a genomicProfiles objecy with ChIP profile scores. Third element contains a goodness of fit metrics.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com

**Examples**

```
library(ChIPanalyser)
data(ChIPanalyserData)
# See GA vignette for usage
```

---

`splitData`*Get Training and Testing data from ChIPscore objects*

---

**Description**

`splitData` splits processed ChIP data into training and testing sets.

**Usage**

```
splitData(ChIPscore, dist = c(80,20), as.proportion = TRUE)
```

**Arguments**

<code>ChIPscore</code>	ChIPscore object as returned by <code>processingChIP</code>
<code>dist</code>	If <code>as.proportion</code> is to <code>TRUE</code> , split the data into desired proportions. Default sets 80% training and 20% testing. If <code>as.proportion</code> is to <code>FALSE</code> , a vector of 4 numeric values describing start and end of training and testing respectively.
<code>as.proportion</code>	Logical describing if values provided to <code>dist</code> should be treated as % of training and testing or if <code>dist</code> should be considered as start and end of loci selected for training and testing respectively.

**Value**

Returns a named list of ChIPScore objects

\* `trainingSet` = ChIPscore containing training set \* `testingSet` = ChIPscore containing testing set.

**Author(s)**

Patrick C.N. Martin <pcnmartin@gmail.com>

**Examples**

```
library(ChIPanalyser)
data(ChIPanalyserData)
# See GA vignette for usage
test <- processingChIP(chip, top)
usingDist <- splitData(test, dist = c(50,50), as.proportion = TRUE )
usingIndex <- splitData(test, dist = c(1,2,3,4), as.proportion = FALSE )
```

---

stepSize	Accessor method of the stepSize slot in <a href="#">parameterOptions</a> object
----------	---

---

### Description

Accessor method of the stepSize slot in [parameterOptions](#) object

### Usage

```
stepSize(object)
```

### Arguments

object            object is a [parameterOptions](#) object.

### Details

It possible to restrict the size of the ChIP-seq-like profile produced by [computeChIPProfile](#). Instead of returning ChIP-seq like score for each base pair, it is possible to skip base pairs and only return the predicted enrichment score for every "n" base pair (n is the value assigned to stepSize). This will reduce the size of the output data (unless step size is very large, this will not affect the accuracy of the model). Default is set at 10 base pairs.

### Value

Returns the value assigned to the stepSize slot in a [parameterOptions](#)

### Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

### Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for maxSignal
stepSize(OPP)
```

---

stepSize-methods      *~~ Methods for Function stepSize ~~*

---

### Description

Accessor method of the stepSize slot in [parameterOptions](#) object

### Methods:

stepSize(object)

---

stepSize<-      *Setter Method for the stepSize slot in a [parameterOptions](#)*

---

### Description

Setter Method for the stepSize slot in a [parameterOptions](#)

### Usage

```
stepSize(object) <- value
```

### Arguments

object      object is a [parameterOptions](#) object  
value      value is a positive numeric value that will be assigned to the stepSize slot in a [parameterOptions](#) object. Default is set at 10 base pairs.

### Details

It possible to restrict the size of the ChIP-seq-like profile produced by [computeChIPProfile](#). Instead of returning ChIP-seq like score for each base pair, it is possible to skip base pairs and only return the predicted enrichment score for every "n" base pair (n is the value assigned to stepSize). This will reduce the size of the output data (unless step size is very large, this will not affect the accuracy of the model). Default is set at 10 base pairs.

### Value

Returns a [parameterOptions](#) object with an updated value for the stepSize slot.

### Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

### References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
# Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for maxSignal
stepSize(OPP) <- 20
```

---

```
stepSize<-methods      ~~ Methods for Function stepSize<- ~~
```

---

**Description**

Setter Method for the stepSize slot in a [parameterOptions](#)

**Methods:**

```
stepSize(object)<-value
```

---

```
strandRule            Accessor Method for the strandRule slot in a parameterOptions object
```

---

**Description**

Accessor Method for the strandRule slot in a [parameterOptions](#) object

**Usage**

```
strandRule(object)
```

**Arguments**

object                object is a [parameterOptions](#) object

**Details**

When computing the PWM Scores and if [whichstrand](#) is set to "+-", strandRule will determine how to handle both strands ( one of three options : "mean", "max", "sum"). If set to "mean", the average PWM Score of both strand will be computed. If set to "max", the highest PWM score between each strand will be selected and finally "sum" will sum both score together. Default set at "max"

**Value**

Returns the value assigned to strandRule slot (one of three options : "mean", "max", "sum") in a [parameterOptions](#) object

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>



## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

## Examples

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions( strandRule="max")
#Accesssing Value for strandRule
strandRule(GPP)
```

---

strandRule-methods      *~~ Methods for Function strandRule ~~*

---

## Description

Accessor Method for the strandRule slot in a [parameterOptions](#) object

### Methods:

strandRule(object)

---

strandRule<-                      *Setter method for the strandRule slot in a [parameterOptions](#) object.*

---

## Description

Setter method for the strandRule slot in a [parameterOptions](#) object.

## Usage

```
strandRule(object) <- value
```

## Arguments

object	object is a <a href="#">parameterOptions</a> object
value	value is a character string and can be one of the following ‘mean’, ‘max’, ‘sum’. This will only apply if <a href="#">whichstrand</a> is ‘+’. Default set at ‘max’

## Details

When computing the PWM Scores and if [whichstrand](#) is set to ‘+’, strandRule will determine how to handle both strands ( one of three options : ‘mean’, ‘max’, ‘sum’). If set to ‘mean’, the average PWM Score of both strand will be computed. If set to ‘max’, the highest PWM score between each strand will be selected and finally ‘sum’ will sum both score together. Default set at ‘max’

**Value**

Returns a [parameterOptions](#) object with an updated value for the strandRule slot

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions(strandRule="max")
#Setting New Value for strandRule
strandRule(GPP) <- "mean"
```

---

strandRule<-methods     *~~ Methods for Function strandRule<- ~~*

---

**Description**

Setter method for the strandRule slot in a [parameterOptions](#) object.

**Methods:**

strandRule(object)<-value

---

whichstrand	<i>Accessor method for the whichstrand slot in a <a href="#">parameterOptions</a> object</i>
-------------	--

---

**Description**

Accessor method for the whichstrand slot in a [parameterOptions](#) object

**Usage**

whichstrand(object)

**Arguments**

object                    object is a [parameterOptions](#) object

**Details**

PWM Score may be computed on either the positive strand ("+"), the negative strand ("-") or on both strands ("+-").

**Value**

Returns on which strand PWM Scores should be computed ( whichstrand in a [parameterOptions](#) object)

**Author(s)**

Patrick C. N. Martin <pm16057@essex.ac.uk>

**References**

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

**Examples**

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions( whichstrand="+-")
#Setting New Value for whichstrand
whichstrand(GPP)
```

---

whichstrand-methods    *~~ Methods for Function whichstrand ~~*

---

**Description**

Accessor method for the whichstrand slot in a [parameterOptions](#) object

**Methods:**

whichstrand(object)

---

whichstrand<-                    *Setter method for the whichstrand slot in a [parameterOptions](#) object*

---

## Description

Setter method for the whichstrand slot in a [parameterOptions](#) object

## Usage

```
whichstrand(object) <- value
```

## Arguments

object	object is a <a href="#">parameterOptions</a> object
value	value is a character string specifying which strand should be used to compute PWM Scores. The three available options are the following: "+", "-", or "+-". Default is "+-"

## Details

PWM Score may be computed on either the positive strand ("+"), the negative strand ("-") or on both strands ("+-").

## Value

Returns a [parameterOptions](#) object with an updated value for the whichstrand slot

## Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

## References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

## Examples

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions( whichstrand="+-")
#Setting New Value for whichstrand
whichstrand(GPP) <- "+"
```

---

*whichstrand<-methods*    *~~ Methods for Function whichstrand<- ~~*

---

**Description**

Setter method for the *whichstrand* slot in a [parameterOptions](#) object

**Methods:**

`whichstrand(object)<-value`

# Index

## \* classes

ChIPScore-class, 19  
genomicProfiles-class, 41  
genomicProfilesInternal-class, 43  
GRList-class, 47  
loci-class, 51  
nos-class, 67  
parameterOptions-class, 69

## \* methods

averageExpPWMScore-methods, 7  
backgroundSignal-methods, 8  
backgroundSignal<-methods, 10  
boundMolecules-methods, 11  
boundMolecules<-methods, 12  
BPFrequency-methods, 13  
BPFrequency<-methods, 15  
chipMean-methods, 17  
chipMean<-methods, 18  
chipSd-methods, 21  
chipSd<-methods, 23  
chipSmooth-methods, 24  
chipSmooth<-methods, 25  
DNASequenceLength-methods, 35  
drop-methods, 36  
initialize-methods, 48  
lambdaPWM-methods, 49  
lambdaPWM<-methods, 50  
loci-methods, 52  
lociWidth-methods, 53  
lociWidth<-methods, 54  
maxPWMScore-methods, 55  
maxSignal-methods, 57  
maxSignal<-methods, 58  
minPWMScore-methods, 59  
naturalLog-methods, 60  
naturalLog<-methods, 62  
noiseFilter-methods, 63  
noiseFilter<-methods, 64  
noOfSites-methods, 65  
noOfSites<-methods, 66  
PFMFormat-methods, 73  
PFMFormat<-methods, 74  
ploidy-methods, 75

ploidy<-methods, 76

PositionFrequencyMatrix-methods,  
81

PositionFrequencyMatrix<-methods,  
82

PositionWeightMatrix-methods, 84

PositionWeightMatrix<-methods, 85

profiles-methods, 88

PWMPseudocount-methods, 89

PWMPseudocount<-methods, 91

PWMThreshold-methods, 92

PWMThreshold<-methods, 93

removeBackground-methods, 94

removeBackground<-methods, 95

scores-methods, 96

show-methods, 99

stepSize-methods, 103

stepSize<-methods, 104

strandRule-methods, 105

strandRule<-methods, 106

whichstrand-methods, 107

whichstrand<-methods, 109

## \* package

ChIPAnalyser-package, 5

.DNASequenceLength<- , genomicProfilesInternal , vector-method  
(genomicProfilesInternal-class),  
43

.ZeroBackground , parameterOptions-method  
(parameterOptions-class), 69

.ZeroBackground<- , parameterOptions , vector-method  
(parameterOptions-class), 69

.averageExpPWMScore<- , genomicProfilesInternal , numeric-method  
(genomicProfilesInternal-class),  
43

.drop<- , genomicProfilesInternal , vector-method  
(genomicProfilesInternal-class),  
43

.generatePWM , genomicProfilesInternal-method  
(genomicProfilesInternal-class),  
43

.maxPWMScore<- , genomicProfilesInternal , vector-method  
(genomicProfilesInternal-class),  
43

- .minPWMScore<-, genomicProfilesInternal, vector-method (genomicProfilesInternal-class), 43
- .paramTag, parameterOptions-method (parameterOptions-class), 69
- .paramTag<-, parameterOptions, character-method (parameterOptions-class), 69
- .profiles<-, genomicProfilesInternal, GRList-method (genomicProfilesInternal-class), 43
- .tags, genomicProfilesInternal-method (genomicProfilesInternal-class), 43
- .tags<-, genomicProfilesInternal, character-method (genomicProfilesInternal-class), 43
- Access (ChIPAnalyserData), 15
- averageExpPWMScore, 6
- averageExpPWMScore, genomicProfilesInternal-method (genomicProfilesInternal-class), 43
- averageExpPWMScore-methods, 7
- backgroundSignal, 7, 68
- backgroundSignal, parameterOptions-method (parameterOptions-class), 69
- backgroundSignal-methods, 8
- backgroundSignal<-, 9
- backgroundSignal<-methods, 10
- backgroundSignal<-, parameterOptions, numeric-method (parameterOptions-class), 69
- boundMolecules, 10, 26, 30–32, 68, 79
- boundMolecules, parameterOptions-method (parameterOptions-class), 69
- boundMolecules-methods, 11
- boundMolecules<-, 11
- boundMolecules<-methods, 12
- boundMolecules<-, parameterOptions, vector-method (parameterOptions-class), 69
- BPFrequency, 12
- BPFrequency, genomicProfilesInternal-method (genomicProfilesInternal-class), 43
- BPFrequency-methods, 13
- BPFrequency<-, 14
- BPFrequency<-methods, 15
- BPFrequency<-, genomicProfilesInternal, DNAStrngSet-method (genomicProfilesInternal-class), 43
- BPFrequency<-, genomicProfilesInternal, vector-method (genomicProfilesInternal-class), 43
- chip (ChIPAnalyserData), 15
- ChIPAnalyser (ChIPAnalyser-package), 5
- ChIPAnalyser-package, 5
- ChIPAnalyserData, 15
- chipMean, 16, 16, 68, 85
- chipMean, parameterOptions-method (parameterOptions-class), 69
- chipMean-methods, 17
- chipMean<-, 17
- chipMean<-methods, 18
- chipMean<-, parameterOptions, numeric-method (parameterOptions-class), 69
- ChIPScore, 25, 26, 33, 50–54, 96
- ChIPScore-class, 19
- chipSd, 20, 68, 85
- chipSd, parameterOptions-method (parameterOptions-class), 69
- chipSd-methods, 21
- chipSd<-, 22
- chipSd<-methods, 23
- chipSd<-, parameterOptions, numeric-method (parameterOptions-class), 69
- chipSmooth, 23, 68, 85
- chipSmooth, parameterOptions-method (parameterOptions-class), 69
- chipSmooth-methods, 24
- chipSmooth<-, 24
- chipSmooth<-methods, 25
- chipSmooth<-, parameterOptions, vector-method (parameterOptions-class), 69
- computeChIPProfile, 8, 9, 16, 18, 21–24, 25, 87, 102, 103
- computeGenomeWideScores, 6, 27, 33, 54, 58
- computeOccupancy, 8–11, 23–25, 29, 93, 95
- computeOptimal, 10, 11, 30, 86
- computePWMScore, 29, 32, 33
- cs (ChIPAnalyserData), 15
- DNASequenceLength, 6, 34
- DNASequenceLength, genomicProfilesInternal-method (genomicProfilesInternal-class), 43
- DNASequenceLength-methods, 35
- DNAStrngSet, 13, 14, 31, 33, 41
- drop, 35
- drop, genomicProfilesInternal-method (genomicProfilesInternal-class), 43
- drop-methods, 36
- generateStartingPopulation, 38

- geneRef (ChIPAnalyserData), 15  
 genomicProfiles, 6, 12–14, 25, 26, 29, 31, 33–36, 39, 40, 43, 45, 54, 55, 58, 69, 72–74, 80–85, 88, 97  
 genomicProfiles-class, 41  
 genomicProfilesInternal, 42  
 genomicProfilesInternal-class, 43  
 getHighestFitnessSolutions, 45  
 getTestingData, 46  
 getTrainingData, 46  
 GRanges, 16, 25, 29, 31, 33, 36, 77, 85, 86  
 GRangesList, 29, 33  
 GRList-class, 47, 47  
  
 initialize, ChIPScore-method  
     (initialize-methods), 48  
 initialize, genomicProfiles-method  
     (initialize-methods), 48  
 initialize, parameterOptions-method  
     (initialize-methods), 48  
 initialize-methods, 48  
  
 lambdaPWM, 26, 30–32, 48, 79  
 lambdaPWM, parameterOptions-method  
     (parameterOptions-class), 69  
 lambdaPWM-methods, 49  
 lambdaPWM<-, 49  
 lambdaPWM<-methods, 50  
 lambdaPWM<-, parameterOptions, vector-method  
     (parameterOptions-class), 69  
 loci, 50  
 loci, ChIPScore-method  
     (ChIPScore-class), 19  
 loci-class, 51  
 loci-methods, 52  
 lociWidth, 52, 68  
 lociWidth, parameterOptions-method  
     (parameterOptions-class), 69  
 lociWidth-methods, 53  
 lociWidth<-, 53  
 lociWidth<-methods, 54  
 lociWidth<-, parameterOptions, numeric-method  
     (parameterOptions-class), 69  
  
 maxPWMScore, 54  
 maxPWMScore, genomicProfilesInternal-method  
     (genomicProfilesInternal-class), 43  
 maxPWMScore-methods, 55  
 maxSignal, 8, 9, 56, 68  
 maxSignal, parameterOptions-method  
     (parameterOptions-class), 69  
 maxSignal-methods, 57  
  
 maxSignal<-, 57  
 maxSignal<-methods, 58  
 maxSignal<-, parameterOptions, numeric-method  
     (parameterOptions-class), 69  
 minPWMScore, 58  
 minPWMScore, genomicProfilesInternal-method  
     (genomicProfilesInternal-class), 43  
 minPWMScore-methods, 59  
  
 naturalLog, 59, 68  
 naturalLog, parameterOptions-method  
     (parameterOptions-class), 69  
 naturalLog-methods, 60  
 naturalLog<-, 61  
 naturalLog<-methods, 62  
 naturalLog<-, parameterOptions, logical-method  
     (parameterOptions-class), 69  
 noiseFilter, 62, 68  
 noiseFilter, parameterOptions-method  
     (parameterOptions-class), 69  
 noiseFilter-methods, 63  
 noiseFilter<-, 63  
 noiseFilter<-methods, 64  
 noiseFilter<-, parameterOptions, character-method  
     (parameterOptions-class), 69  
 noOfSites, 64, 68  
 noOfSites, parameterOptions-method  
     (parameterOptions-class), 69  
 noOfSites-methods, 65  
 noOfSites<-, 65  
 noOfSites<-methods, 66  
 noOfSites<-, parameterOptions, character-method  
     (parameterOptions-class), 69  
 noOfSites<-, parameterOptions, numeric-method  
     (parameterOptions-class), 69  
 nos-class, 67  
  
 parameterOptions, 7–11, 16–18, 20–24, 26, 29, 31, 33, 40, 42, 43, 45, 48, 49, 52, 53, 56, 57, 59–66, 67, 74–76, 85–95, 102–109  
 parameterOptions-class, 69  
 PFMFormat, 72  
 PFMFormat, genomicProfilesInternal-method  
     (genomicProfilesInternal-class), 43  
 PFMFormat-methods, 73  
 PFMFormat<-, 73  
 PFMFormat<-methods, 74  
 PFMFormat<-, genomicProfilesInternal, character-method  
     (genomicProfilesInternal-class), 43



- ploidy, 68, 74  
 ploidy, parameterOptions-method  
     (parameterOptions-class), 69  
 ploidy-methods, 75  
 ploidy<-, 75  
 ploidy<-methods, 76  
 ploidy<- , parameterOptions, numeric-method  
     (parameterOptions-class), 69  
 plotOccupancyProfile, 8, 9, 76  
 plotOptimalHeatMaps, 78  
 PositionFrequencyMatrix, 72, 73, 80  
 PositionFrequencyMatrix, genomicProfilesInternal-method  
     (genomicProfilesInternal-class),  
     43  
 PositionFrequencyMatrix-methods, 81  
 PositionFrequencyMatrix<-, 81  
 PositionFrequencyMatrix<-methods, 82  
 PositionFrequencyMatrix<- , genomicProfilesInternal, character-method  
     (genomicProfilesInternal-class),  
     43  
 PositionFrequencyMatrix<- , genomicProfilesInternal, matrix-method  
     (genomicProfilesInternal-class),  
     43  
 PositionWeightMatrix, 81, 83  
 PositionWeightMatrix, genomicProfilesInternal-method  
     (genomicProfilesInternal-class),  
     43  
 PositionWeightMatrix-methods, 84  
 PositionWeightMatrix<-, 84  
 PositionWeightMatrix<-methods, 85  
 PositionWeightMatrix<- , genomicProfilesInternal, matrix-method  
     (genomicProfilesInternal-class),  
     43  
 processingChIP, 19, 20, 25, 31, 33, 50, 52,  
     53, 68, 85, 96  
 profileAccuracyEstimate, 8, 9, 77, 86  
 profiles, 29, 33, 47  
 profiles (profiles-methods), 88  
 profiles, genomicProfilesInternal-method  
     (genomicProfilesInternal-class),  
     43  
 profiles-methods, 88  
 PWMpseudocount, 68, 88  
 PWMpseudocount, parameterOptions-method  
     (parameterOptions-class), 69  
 PWMpseudocount-methods, 89  
 PWMpseudocount<-, 90  
 PWMpseudocount<-methods, 91  
 PWMpseudocount<- , parameterOptions, numeric-method  
     (parameterOptions-class), 69  
 PWMThreshold, 28, 32, 33, 68, 91  
 PWMThreshold, parameterOptions-method  
     (parameterOptions-class), 69  
 PWMThreshold-methods, 92  
 PWMThreshold<-, 92  
 PWMThreshold<-methods, 93  
 PWMThreshold<- , parameterOptions, numeric-method  
     (parameterOptions-class), 69  
 removeBackground, 68, 93  
 removeBackground, parameterOptions-method  
     (parameterOptions-class), 69  
 removeBackground-methods, 94  
 removeBackground<-, 94  
 removeBackground<-methods, 95  
 removeBackground<- , parameterOptions, vector-method  
     (parameterOptions-class), 69  
 scores, 96  
 scores, ChIPScore-method  
     (character-method)  
     (ChIPScore-class), 19  
 scores-methods, 96  
 searchSites, 77, 97  
 setChromatinStates, 98  
 show, ChIPScore-method (show-methods), 99  
 show, genomicProfiles-method  
     (show-methods), 99  
 show, parameterOptions-method  
     (show-methods), 99  
 show-methods, 99  
 singleRun, 100  
 splitData, 101  
 stepSize, 68, 102  
 stepSize, parameterOptions-method  
     (parameterOptions-class), 69  
 stepSize-methods, 103  
 stepSize<-, 103  
 stepSize<-methods, 104  
 stepSize<- , parameterOptions, numeric-method  
     (parameterOptions-class), 69  
 strandRule, 68, 104  
 strandRule, parameterOptions-method  
     (parameterOptions-class), 69  
 strandRule-methods, 105  
 strandRule<-, 105  
 strandRule<-methods, 106  
 strandRule<- , parameterOptions, character-method  
     (parameterOptions-class), 69  
 top (ChIPAnalyserData), 15  
 whichstrand, 68, 104, 105, 106  
 whichstrand, parameterOptions-method  
     (parameterOptions-class), 69  
 whichstrand-methods, 107

`whichstrand<-`, [108](#)  
`whichstrand<-methods`, [109](#)  
`whichstrand<-`, `parameterOptions`, `character-method`  
(`parameterOptions-class`), [69](#)