

# Package ‘DOQTL’

October 27, 2015

**Version** 1.6.0

**Date** 2012-12-07

**Title** Genotyping and QTL Mapping in DO Mice

**Author** Daniel Gatti, Karl Broman, Andrey Shabalin

**Maintainer** Daniel Gatti <Dan.Gatti@jax.org>

**Depends** R (>= 3.0.0), BSgenome.Mmusculus.UCSC.mm10, GenomicRanges, VariantAnnotation

**Imports** annotate, annotationTools, biomaRt, Biobase, BiocGenerics, corpcor, doParallel, foreach, fpc, hwriter, IRanges, iterators, mclust, QTLRel, regress, rhdf5, Rsamtools, RUnit, XML

**Suggests** MUGAExampleData, doMPI

**Description** DOQTL is a quantitative trait locus (QTL) mapping pipeline designed for Diversity Outbred mice and other multi-parent outbred populations. The package reads in data from genotyping arrays and perform haplotype reconstruction using a hidden Markov model (HMM). The haplotype probabilities from the HMM are then used to perform linkage mapping. When founder sequences are available, DOQTL can use the haplotype reconstructions to impute the founder sequences onto DO genomes and perform association mapping.

**biocViews** GeneticVariability, SNP, Genetics, HiddenMarkovModel

**License** GPL-3

**LazyData** true

**ByteCompile** yes

**URL** <http://do.jax.org>

**NeedsCompilation** yes

## R topics documented:

add.missing.F1s . . . . .	3
add.slash . . . . .	4
addLog . . . . .	4
addLogVector . . . . .	5
assoc.map . . . . .	6
assoc.plot . . . . .	8
batch.normalize . . . . .	9
bayesint . . . . .	10
calc.genoprob . . . . .	11

calc.genoprob.alleles . . . . .	13
calc.genoprob.intensity . . . . .	14
categorize.variants . . . . .	15
cc.trans.probs . . . . .	16
cluster.strains . . . . .	17
coef.doqtl . . . . .	18
coefplot . . . . .	18
colSumsLog . . . . .	19
condense.model.probs . . . . .	20
convert.allele.calls . . . . .	21
convert.genes.to.GRanges . . . . .	22
convert.genotypes . . . . .	22
convert.variants.to.GRanges . . . . .	23
convert.variants.to.numeric . . . . .	23
create.genotype.states . . . . .	24
create.html.page . . . . .	25
create.Rdata.files . . . . .	26
do.colors . . . . .	26
do.states . . . . .	27
do.trans.probs . . . . .	27
do2sanger . . . . .	28
estimate.cluster.params . . . . .	30
example.genes . . . . .	31
example.pheno . . . . .	32
example.qtl . . . . .	33
example.snps . . . . .	34
extract.raw.data . . . . .	34
fast.qtlrel . . . . .	35
fill.in.snps . . . . .	36
filter.geno.probs . . . . .	37
filter.samples . . . . .	38
find.overlapping.genes . . . . .	38
gene.plot . . . . .	39
generic.trans.probs . . . . .	40
genome.summary.plots . . . . .	41
get.chr.lengths . . . . .	42
get.do.states . . . . .	43
get.gene.name . . . . .	43
get.machine.precision . . . . .	44
get.max.geno . . . . .	44
get.mgi.features . . . . .	45
get.num.auto . . . . .	46
get.pattern.variants . . . . .	47
get.strains . . . . .	48
get.trans.probs . . . . .	48
get.variants . . . . .	49
html.report . . . . .	50
intensity.plots . . . . .	51
kinship.probs . . . . .	52
muga.snps.to.keep . . . . .	53
plot.doqtl . . . . .	54
plot.genoprobs . . . . .	55

<i>add.missing.F1s</i>	3
pxg.plot . . . . .	56
qtl.heatmap . . . . .	57
qtl.LRS . . . . .	58
qtl.qtlrel . . . . .	59
qtl.simulate . . . . .	60
rankZ . . . . .	61
read.vcf . . . . .	62
scanone . . . . .	63
scanone.eqtl . . . . .	65
scanone.perm . . . . .	66
sdp.plot . . . . .	67
sex.predict . . . . .	68
snp.plot . . . . .	68
summarize.genotype.transitions . . . . .	70
variant.plot . . . . .	71
write.founder.genomes . . . . .	72
<b>Index</b>	<b>74</b>

---

<code>add.missing.F1s</code>	<i>Add Missing F1 Samples</i>
------------------------------	-------------------------------

---

### Description

Given a set of CC or DO founders, impute the genotypes or intensities of missing F1s.

### Usage

```
add.missing.F1s(founders, snps)
```

### Arguments

<code>founders</code>	List, required: Contains either an element called 'geno' or two elements called 'x' and 'y'.
<code>snps</code>	Data.frame, required: Data.frame containing the SNPs. SNP ID, chr, Mb and cM locations in column 1 - 4, respectively.

### Value

List with the founders data structure updated to include missing FALSE1 samples.

### Author(s)

Daniel Gatti

### Examples

```
## Not run:
load(url("ftp://ftp.jax.org/MUGA/muga_snps.Rdata"))
founders = add.missing.FALSE1s(founders, snps = muga_snps)

## End(Not run)
```

add.slash                      *Add a forward slash to a character string.*

---

**Description**

If the argument does not end with a forward slash, add one.

**Usage**

```
add.slash(path)
```

**Arguments**

path                      Character string containing a file path.

**Value**

Returns a character string with a forward slash added to the end, if the argument did not end with a forward slash already.

**Author(s)**

Daniel Gatti

**Examples**

```
add.slash("/dir")
```

---

addLog                      *Add two log values.*

---

**Description**

When two numbers that are on the log scale must be summed, transforming them back to the non-log scale and taking the sum is slow. This function takes the exp() of the values only when necessary.

**Usage**

```
addLog(x, y)
```

**Arguments**

x                      Numeric containing a value on the log scale.

y                      Numeric containing a value on the log scale.

**Details**

This function checks to see if the difference between the maximum value and the other value is less than the machine precision. If it is, then the exp() is taken for those values that differ by less than the machine precision, they are summed and returned to a log scale. If the maximum value is differs from the other values by greater than the machine precision, then return the maximum value.

**Value**

Numeric value containing the sum of the arguments on a log scale.

**Author(s)**

Daniel Gatti

**See Also**

[addLogVector](#)

**Examples**

```
addLog(log(10), log(1))
```

---

addLogVector

*Add a vector of log values.*

---

**Description**

When a summation must be taken on a vector of numbers that are on the log scale, transforming them back to the non-log scale and taking the sum is slow. This function takes the `exp()` of the values only when necessary.

**Usage**

```
addLogVector(x)
```

**Arguments**

`x` Numeric vector containing values on a log scale to be summed on the untransformed scale.

**Details**

This function checks to see if the difference between the maximum values and the remaining values is less than the machine precision. If it is, then the `exp()` is taken for those values that differ by less than the machine precision, they are summed and returned to a log scale. If the maximum value is differs from the other values by greater than the machine precision, then return the maximum value.

**Value**

Numeric value containing the sum of the values on a log scale.

**Author(s)**

Daniel Gatti

**See Also**

[addLog](#)

**Examples**

```
addLogVector(log(1:10))
```

---

```
assoc.map
```

---

*Perform association mapping on DO mice.*

---

**Description**

Given the phenotypes and genotype probabilities, impute the Sanger SNPs onto DO genomes and perform association mapping.

**Usage**

```
assoc.map(pheno, pheno.col = 1, probs, K, addcovar, snps, chr, start, end,
model = c("additive", "dominance", "full"), scan = c("one", "two"),
output = c("lod", "p-value", "bic"),
snp.file = "ftp://ftp.jax.org/SNPtools/variants/cc.snps.NCBI38.txt.gz")
assoc.map.perms(pheno, pheno.col = 1, probs, addcovar, snps,
model = c("additive", "dominance", "full"),
scan = c("one", "two"), output = c("lod", "p-value", "bic"),
snp.file = "ftp://ftp.jax.org/SNPtools/variants/cc.snps.NCBI38.txt.gz",
nperm = 1000)
```

**Arguments**

pheno	Data.frame containing the phenotype data. Sample IDs must be in rownames. One of the columns should be called sex and contain M or FALSE to indicate the sex of each sample.
pheno.col	Either a numeric vector containing column IDs to map in pheno or a vector of column names in pheno.
probs	A 3 dimensional array of genotype probabilities as provided from <a href="#">condense.model.probs</a> . Samples, founder and markers in dims 1:3. All dimensions must have dimnames.
K	Numeric matrix containing kinship values for the samples in pheno and probs. Sample IDs must be in rownames and colnames.
addcovar	Numeric matrix of additive covariates to use in mapping. Sample IDs must be in rownames.
snps	Data.frame containing marker IDs, chromosomes, Mb and cM locations in columns 1:4.
chr	Character containing the chromosome on which to map.
start	Numeric value containing the proximal position for mapping. May be in Mb or base pairs (see Details).
end	Numeric value containing the distal position for mapping. May be in Mb or base pairs (see Details).
model	Character string that is one of "additive", "dominance" or "full". Indicates the type of model to fit. Note that the probs must match the type of model being fit. See <a href="#">condense.model.probs</a> to output different probs.

scan	Character string that is either "one", or "two" indicating whether a single scan or a pairwise scan should be performed across the interval.
output	Character string that is either "lod", "p-value" or "bic" indicating the mapping statistic to return.
snp.file	Character string containing the full path to the SNP file to use. Currently points to a location on the <a href="#">Jackson Laboratory FALSETP server</a>
nperm	Integer indicating the number of permutations to perform. Default = 1000.

### Details

FALSEor each interval between two markers, we take the average founder haplotype contribution for each sample. Then, using the proportion of each founder (8 in the case of DO mice), we impute the Sanger SNPs in this interval onto each DO sample.

The start and ending locations are assumed to be in Mb if they are below 200 and in bp if over 200.

### Value

Data.frame containing the locations, SNPs and mapping statistic for the requested samples in the requested interval.

### Author(s)

Daniel Gatti

### References

Combined sequence-based and genetic mapping analysis of complex traits in outbred rats. Rat Genome Sequencing and Mapping Consortium, Baud A, Hermsen R, Guryev V, Stridh P, Graham D, McBride MW, FALSEoroud T, Calderari S, Diez M, Ockinger J, Beyeen AD, Gillett A, Abdelmagid N, Guerreiro-Cacais AO, Jagodic M, Tuncel J, Norin U, Beattie E, Huynh N, Miller WH, Koller DL, Alam I, FALSEalak S, Osborne-Pellegrin M, Martinez-Membrives E, Canete T, Blazquez G, Vicens-Costa E, Mont-Cardona C, Diaz-Moran S, Tobena A, Hummel O, Zelenika D, Saar K, Patone G, Bauerfeind A, Bihoreau MT, Heinig M, Lee YA, Rintisch C, Schulz H, Wheeler DA, Worley KC, Muzny DM, Gibbs RA, Lathrop M, Lansu N, Toonen P, Ruzius FALSEP, de Bruijn E, Hauser H, Adams DJ, Keane T, Atanur SS, Aitman TJ, FALSElicek P, Malinauskas T, Jones EY, Ekman D, Lopez-Aumatell R, Dominiczak AFALSE, Johannesson M, Holmdahl R, Olsson T, Gauguier D, Hubner N, FALSEernandez-Teruel A, Cuppen E, Mott R, FALSElint J. *Nat Genet.* 2013 Jul;45(7):767-75. doi: 10.1038/ng.2644. Epub 2013 May 26. PMID: 23708188 Using progenitor strain information to identify quantitative trait nucleotides in outbred mice. Yalcin B, FALSElint J, Mott R. *Genetics.* 2005 Oct;171(2):673-81. Epub 2005 Aug 5. PMID: 16085706 Mouse genomic variation and its effect on phenotypes and gene regulation. Keane TM, Goodstadt L, Danecek P, White MA, Wong K, Yalcin B, Heger A, Agam A, Slater G, Goodson M, FALSEurlotte NA, Eskin E, Nellaker C, Whitley H, Cleak J, Janowitz D, Hernandez-Pliego P, Edwards A, Belgard TG, Oliver PL, McIntyre RE, Bhomra A, Nicod J, Gan X, Yuan W, van der Weyden L, Steward CA, Bala S, Stalker J, Mott R, Durbin R, Jackson IJ, Czechanski A, Guerra-Assuncao JA, Donahue LR, Reinholdt LG, Payseur BA, Ponting CP, Birney E, FALSElint J and Adams DJ *Nature* 2011;477;7364;289-94 PUBMED: 21921910 Sequence-based characterization of structural variation in the mouse genome. Yalcin B, Wong K, Agam A, Goodson M, Keane TM, Gan X, Nellaker C, Goodstadt L, Nicod J, Bhomra A, Hernandez-Pliego P, Whitley H, Cleak J, Dutton R, Janowitz D, Mott R, Adams DJ and FALSElint J *Nature* 2011;477;7364;326-9 PUBMED: 21921916

**See Also**

[assoc.plot](#)

**Examples**

```
## Not run: assoc.map(pheno = pheno, pheno.col = 1, probs = probs, K = K, addcovar = addcovar,
                    snps = snps, chr = 1, start = 40, end = 45)
## End(Not run)
```

---

assoc.plot

*Plot association mapping results.*

---

**Description**

After performing association mapping using [assoc.map](#), plot the mapping statistic along with genes in the QTL interval.

**Usage**

```
assoc.plot(results,
           mgi.file = "ftp://ftp.jax.org/SNPtools/genes/MGI.20130703.sorted.txt.gz",
           highlight, highlight.col = "red", thr, show.sdps = FALSE, ...)
```

**Arguments**

results	Data.frame containing output from <a href="#">assoc.map</a> .
mgi.file	Character string containing the full path to a Tabix indexed file of gene locations. Default points to a version of the MGI genome feature file.
highlight	Character vector containing gene symbols to highlight in the plot.
highlight.col	Vector of colors to use when highlighting genes.
thr	Numeric value above which data points should be colored red and SNPs with these points returned.
show.sdps	Logical value (default = FALSE) that is TRUE if the strain distribution pattern (SDP) for the SNPs should be shown. When used with thr, only plots the founder SDPs for SNPs above thr.
...	Additional arguments passed to plot.

**Details**

Given the output from [assoc.map](#), plot the LOD or difference in BIC values across the QTL interval in the top panel. Plot the genes in the interval in the lower panel. Make sure to use Sanger SNP and MGI feature files that are on the same genome build.

**Value**

A plot with the mapping statistic in the top panel and genes in the lower panel. If thr is not missing, then filter the SNPs in the results argument and return only those with a mapping statistic greater than thr.



**Author(s)**

Daniel Gatti

**See Also**[assoc.map](#)**Examples**

```
## Not run:
results = assoc.map(pheno = pheno, pheno.col = 1, probs = probs, K = K, addcovar = addcovar,
snps = snps, chr = 1, start = 40, end = 45)
assoc.plot(results, thr = 3, show.sdps = TRUE)

## End(Not run)
```

---

`batch.normalize`*Batch normalize the X & Y intensity data.*

---

**Description**

This function batch normalizes the X & Y intensity data by subtracting batch medians from the X & Y intensities.

**Usage**

```
batch.normalize(path = ".", snps)
quantilenorm(x1, y1, x2, y2)
```

**Arguments**

path	Character, the full path to the input files, which must be either "x.txt" and "y.txt" or "x.filt.txt" and "y.filt.txt".
snps	Data.frame, with three columns containing SNP ID, chromosome and Mb location in that order. May be obtained from <a href="ftp://ftp.jax.org/MUGA">ftp://ftp.jax.org/MUGA</a> .
x1	Numeric matrix containing X intensities for batch 1 containing samples in rows and markers in columns. Number of samples should be larger than x2.
y1	Numeric matrix containing Y intensities for batch 1 containing samples in rows and markers in columns. Number of samples should be larger than y2.
x2	Numeric matrix containing X intensities for batch 2 containing samples in rows and markers in columns.
y2	Numeric matrix containing Y intensities for batch 2 containing samples in rows and markers in columns.

**Details**

quantile.norm adjusts the intensities of samples in batch 2 to those of batch 1. The number of samples in batch 1 should be greater than the number of samples in batch 2. At each SNP, we form quantiles of the X1 (or Y1) intensity distribution, discarding the upper and lower 0.01

**Value**

`FALSE` or `batch.normalize`: returns value is returned. The batch normalized intensities are written to "x.filt.batch.norm.txt" and "y.filt.batch.norm.txt".

`FALSE` or `quantilenorm`: returns normalized X and Y values for batch 2.

**Note**

`FALSE` future releases may include more sophisticated normalization algorithms.

**Author(s)**

Daniel Gatti

**See Also**

[extract.raw.data](#), [filter.samples](#)

**Examples**

```
## Not run:
  load(url("ftp://ftp.jax.org/MUGA/muga_snps.Rdata"))
  batch.normalize(path = "/demo/MUGA/", snps = muga_snps)

## End(Not run)
```

---

bayesint

*FALSE* find a Bayesian Credible Interval around a QTL.

---

**Description**

This function normalizes the area under the QTL curve on the given chromosome and finds a region that is 95

**Usage**

```
bayesint(qtl, chr, prob = 0.95, expandtomarkers = FALSE)
```

**Arguments**

<code>qtl</code>	data.frame: four columns with SNP ID, Chr, position, and LOD in each column.
<code>chr</code>	character: the chromosome on which the QTL lies.
<code>prob</code>	numeric: must be between 0 and 1
<code>expandtomarkers</code>	boolean: if TRUE, expand the QTL interval to the nearest flanking markers. Default = FALSE

**Value**

Data frame with the SNP ID, Chr, position and LOD for the left and right side of the interval and the maximum QTL.

**Author(s)**

Daniel Gatti

**References**

Saunak, S. (2001) A Statistical FALSE framework for Quantitative Trait Mapping. *Genetics*, **159** (1), 371–387.

**See Also**

[scanone](#), [scanone.perm](#)

**Examples**

```
## Not run: bayesint(qtl, 1)
```

---

 calc.genoprob

---

*Calculate the founder genotype probabilities at each SNP.*


---

**Description**

This function performs genome reconstruction using either allele calls or allele intensities. We recommend using allele intensities where available because they often produce better genotype reconstructions.

**Usage**

```
calc.genoprob(data, chr = "all", output.dir = ".", plot = TRUE,
  array = c("megamuga", "muga", "other"),
  samplotype = c("DO", "CC", "DOF1", "other"), method = c("intensity", "allele"),
  founders, transprobs, snps)
```

**Arguments**

**data** A list with named elements containing the information needed to reconstruct genomes.

When `method = intensity`: `x`: Numeric matrix, `num.samples` x `num.snps`, with `X` intensities for all samples. Sample IDs and SNP IDs must be in `rownames` and `colnames`. `y`: Numeric matrix, `num.samples` x `num.snps`, with `Y` intensities for all samples. Sample IDs and SNP IDs must be in `rownames` and `colnames`. `sex`: Character vector, containing "M" or F indicating sex. Sample IDs must be in `names`. `gen`: Character matrix containing the generation of DO outbreeding for each sample. For the DO, this should be "DO" followed by a number with no space between them. For CC mice, this should be CC. Sample IDs must be in `names`.

When `method = allele`: `geno`: Character matrix, `num.samples` x `num.snps`, with allele calls (A,C,G,T,H or N) for all samples. Sample IDs and SNP IDs must be in `rownames` and `colnames`. `sex`: Character vector, containing "M" or F indicating sex. Sample IDs must be in `names`. `gen`: Character matrix containing the generation of DO outbreeding for each sample. For the DO, this should be "DO" followed by a number with no space between them. For CC mice, this should be CC. Sample IDs must be in `names`.

chr	Character vector containing chromosomes to run. Must match the chromosome IDs in the snps table. "all" (default) will run all chromosomes.
output.dir	Character string containing the full path where output should be written. The directory must exist already.
plot	Boolean that is true if the user would like to plot a sample chromosome as the model progresses. Default = TRUE.
array	Character string indicating the array type. Must be one of "megamuga", "muga" or "other". Default equals "megamuga".
samplotype	Character string containing the type of samples being run. Must be one of "DO", "CC", "DOF1", or "other". Default equals "DO".
method	Character string containing method of genome reconstruction. Must be one of "intensity" or "allele". Default equals "intensity".
founders	List containing founder information for non-DO or CC crosses. <i>Not required for DO.</i> When method = intensity: x: Numeric matrix, num.samples x num.snps, with X intensities for all founders and F1s (if available). Sample IDs and SNP IDs must be in rownames and colnames. y: Numeric matrix, num.samples x num.snps, with Y intensities for all founders and F1s (if available). Sample IDs and SNP IDs must be in rownames and colnames. sex: Character vector, containing "M" or "F" indicating sex. Sample IDs must be in names. code: Character vector containing two letter genotype codes for each founder sample. Sample IDs must be in names. When method = allele: geno: Character matrix, num.samples x num.snps, with allele calls for all founders and F1s (if available). Sample IDs and SNP IDs must be in rownames and colnames. sex: Character vector, containing "M" or "F" indicating sex. Sample IDs must be in names. code: Character vector containing two letter genotype codes for each founder sample. Sample IDs must be in names. When samplotype = DOF1 x: Numeric matrix, num.samples x num.snps, with X intensities for all founders and F1s (if available). Sample IDs and SNP IDs must be in rownames and colnames. y: Numeric matrix, num.samples x num.snps, with Y intensities for all founders and F1s (if available). Sample IDs and SNP IDs must be in rownames and colnames. sex: Character vector, containing "M" or "F" indicating sex. Sample IDs must be in names. code: Character vector containing two letter genotype codes for each founder sample. This should be "II" for an inbred mutant strain. Sample IDs must be in names. direction: Character string that is either "DOxMUT" if a female DO was crossed with a male mutant mouse or "MUTxDO" if a female mutant mouse was crossed with a male DO. This affects the genotyping of the X chromosome.
transprobs	Function to call to estimate the transition probabilities between markers for non-DO samples. <i>Not required for DO.</i>
snps	Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively. <i>Not required for DO.</i>

**Value**

No value is returned. The output files are written to output.dir.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run:
  calc.genoprob(cross, chr.to.run = 1:19, output.dir = "do.data", plot = FALSE,
init.means = NULL, init.covars = NULL)

## End(Not run)
```

---

calc.genoprob.alleles *Calculate the founder genotype probabilities at each SNP using allele calls.*

---

**Description**

This function performs genome reconstruction using allele calls. We recommend using allele intensities where available because they often produce better genotype reconstructions.

**Usage**

```
calc.genoprob.alleles(data, chr, founders, snps, output.dir = ".",
trans.prob.fxn = do.trans.probs, plot = FALSE)
```

**Arguments**

data	A list with named elements containing the information needed to reconstruct genomes. geno: Character matrix, num.samples x num.snps, with allele calls (A,C,G,T,H or N) for all samples. Sample IDs and SNP IDs must be in rownames and colnames. sex: Character vector, containing "M" or F indicating sex. Sample IDs must be in names. gen: Character matrix containing the generation of DO outbreeding for each sample. For the DO, this should be "DO" followed by a number with no space between them. For CC mice, this should be CC. Sample IDs must be in names.
chr	Character vector containing chromosomes to run. Must match the chromosome IDs in the snps table. "all" (default) will run all chromosomes.
founders	List containing founder information for non-DO or CC crosses. When method = allele: geno: Character matrix, num.samples x num.snps, with allele calls (A,C,G,T,H or N) for all founders and FALSEs (if available). Sample IDs and SNP IDs must be in rownames and colnames. sex: Character vector, containing "M" or F indicating sex. Sample IDs must be in names. code: Character vector containing two letter genotype codes for each founder sample. Sample IDs must be in names.
snps	Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively. <i>Not required for DO.</i>
output.dir	Character string containing the full path where output should be written. The directory must exist already.
plot	Boolean that is true if the user would like to plot a sample chromosome as the model progresses. Default = TRUE.
trans.prob.fxn	Function to use when computing the transition probabilities between markers.

**Value**

No value is returned. The output files are written to output.dir.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run:
  calc.genoprob.alleles(data, chr = 1:19, founders = founders,
snp = snps, output.dir = "do.data")

## End(Not run)
```

---

calc.genoprob.intensity

*Calculate the founder genotype probabilities at each SNP.*

---

**Description**

This function performs genome reconstruction using allele intensities. We recommend using allele intensities where available because they often produce better genotype reconstructions.

**Usage**

```
calc.genoprob.intensity(data, chr, founders, snps, output.dir = ".", trans.prob.fxn,
plot = FALSE)
```

**Arguments**

data	A list with named elements containing the information needed to reconstruct genomes. When method = intensity: x: Numeric matrix, num.samples x num.snps, with X intensities for all samples. Sample IDs and SNP IDs must be in rownames and colnames. y: Numeric matrix, num.samples x num.snps, with Y intensities for all samples. Sample IDs and SNP IDs must be in rownames and colnames. sex: Character vector, containing "M" or "F" indicating sex. Sample IDs must be in names. gen: Character matrix containing the generation of DO outbreeding for each sample. For the DO, this should be "DO" followed by a number with no space between them. For CC mice, this should be CC. Sample IDs must be in names.
chr	Character vector containing chromosomes to run. Must match the chromosome IDs in the snps table. "all" (default) will run all chromosomes.
founders	List containing founder information for non-DO or CC crosses. <i>Not required for DO.</i> When method = intensity: x: Numeric matrix, num.samples x num.snps, with X intensities for all founders and F1s (if available). Sample IDs and SNP IDs must be in rownames and colnames. y: Numeric matrix, num.samples x num.snps, with Y intensities for all founders and F1s (if available). Sample IDs and SNP

	IDs must be in rownames and colnames. sex: Character vector, containing "M" or "F" indicating sex. Sample IDs must be in names. code: Character vector containing two letter genotype codes for each founder sample. Sample IDs must be in names.
snps	Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively. <i>Not required for DO.</i>
output.dir	Character string containing the full path where output should be written. The directory must exist already.
trans.prob.fxn	FALSEunction to call to estimate the transition probabilities between markers for non-DO samples. <i>Not required for DO.</i>
plot	Boolean that is true if the user would like to plot a sample chromosome as the model progresses. Default = TRUE.

**Value**

No value is returned. The output files are written to output.dir.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run:
  calc.genoprob.intensity(data, chr, founders, snps, output.dir = ".", trans.prob.fxn,
plot = FALSE)

## End(Not run)
```

---

categorize.variants     *categorize.variants*

---

**Description**

This function intersects the given variants with the genes in that region and classifies them according to "intergenic", "3UTR", "exon", "intron" or "5UTR".

**Usage**

```
categorize.variants(variants,
  mgi.file = "http://cgd.jax.org/tools/SNPtools/MGI/MGI.20130305.sorted.txt.gz")
```

**Arguments**

variants	data.frame, Variants as returned by <code>get.variants{get.variants}</code> .
mgi.file	Character, full path to the MGI feature file. On the JAX campus, this defaults to "http://cgd.jax.org/tools/SNPtools/MGI/MGI.20130305.sorted.txt.gz".

**Value**

FALSEor SNPs and Indels: data.frame: with eight columns: ID, CHR, POS, REFALSE, ALT, symbol, id, type. The first four columns are simply copied over from the SNP file. The symbol column contains the Gene Symbol. The id column contains a gene ID (MGI, Ensembl, NCBI or VEGA). The type column contains "intergenic", "3UTR", "exon", "intron" or "5UTR", depending on the location of the variant in a gene. FALSEor SVs: data.frame: with eight columns: ID, CHR, POS, REFALSE, ALT, symbol, id, type. The first four columns are simply copied over from the SNP file. The symbol column contains the Gene Symbol. The id column contains a gene ID (MGI, Ensembl, NCBI or VEGA). The type column contains "intergenic", "3UTR", "exon", "intron" or "5UTR", depending on the location of the variant in a gene.

**Author(s)**

Daniel Gatti

**See Also**

[get.variants{get.variants}](#)

**Examples**

```
data(example.snps)
snp.type = categorize.variants(variants = example.snps[1:50,])
```

---

cc.trans.probs

*Transition probabilities for CC mice.*

---

**Description**

This function returns the transition probabilities for fully inbred Collaborative Cross mice.

**Usage**

```
cc.trans.probs(states, snps, chr = c(1:19, "X"), sex = c("M", "F"))
```

**Arguments**

states	Character vector containing the two letter codes (i.e. AA, BB, CC, etc.) for the homozygous states.
snps	Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively. <i>Not required for DO.</i>
chr	Character containing the chromosome.
sex	Character that is either M or FALSE, indicating the sex to use. Only relevant on X chromosome.

**Details**

This function calculates the transition probabilities for fully inbred CC mice between two markers. It uses the equations for eight way RILs by sib mating.



**Value**

A matrix of transition probabilities between genotype states.

**Note**

This function has not been fully tested.

**Author(s)**

Daniel Gatti

**References**

The genomes of recombinant inbred lines. Broman KW. Genetics. 2005 Feb;169(2):1133-46. Epub 2004 Nov 15. Erratum in: Genetics. 2006 Aug;173(4):2419. PMID: 15545647

**Examples**

```
## Not run: cc.trans.probs(states = states, snps = snps, chr = 1)
```

---

cluster.strains	<i>cluster.strains</i>
-----------------	------------------------

---

**Description**

Given a set of numeric SNPs, cluster the strains based on allele sharing. This function hierarchically clusters the strains based on the proportion of SNPs that share alleles between each strain. Note: numeric snps, not character snps, are the input to this function.

**Usage**

```
cluster.strains(variants)
```

**Arguments**

variants            data.frame, numeric variants as returned by [convert.variants.to.numeric](#).

**Value**

data.frame: of numeric variants with the strains clustered.

**Author(s)**

Daniel Gatti

**See Also**

[convert.variants.to.numeric](#), [variant.plot](#)

**Examples**

```
data(example.snps)
variants = convert.variants.to.numeric(variants = example.snps[1:100,])
variants = cluster.strains(variants)
```

coef.doqtl *Return the coefficients of a DOQTL object.*

---

### Description

Return the coefficients of a DOQTL object.

### Usage

```
## S3 method for class 'doqtl'  
coef(object, ...)
```

### Arguments

object            A DOQTL object as returned by [scanone](#).  
...                Additional arguments.

### Value

List containing matrices with QTL mapping model coefficients.

### Author(s)

Daniel Gatti

### See Also

[scanone](#)

### Examples

```
head(coef(example.qtl[[1]]))
```

---

coefplot *Plot the QTL model coefficients*

---

### Description

Given a DOQTL object, plot the founder allele coefficients on one chromosome. The coefficients are centered around zero before plotting.

### Usage

```
coefplot(doqtl, chr = 1, stat.name = "LOD", conf.int = TRUE, legend = TRUE,  
colors = "D0", sex, ...)
```

**Arguments**

doqtl	A DOQTL object as produced by <a href="#">scanone</a> . A list containing two elements: lod and coef.
chr	Character containing the chromosome to plot.
stat.name	Character string containing the name of the mapping statistic.
conf.int	Boolean that is TRUE if the QTL support interval should be shaded in the plot. Default = TRUE.
legend	Boolean that is TRUE if the color legend for the DO founders should be drawn. Default = TRUE.
colors	Either "DO", in which case DO colors are supplied or a data.frame with three columns containing the founder letter code, founder strain name and founder color in columns 1:3.
sex	Character that is either FALSE or M, indicating the sex to use. Only used on X chromosome.
...	Additional arguments to be passed to plot.

**Value**

No value is returned. A plot with the founder coefficients in the top panel and the LOD score in the bottom panel is drawn.

**Author(s)**

Daniel Gatti

**See Also**

[scanone](#), [plot.doqtl](#)

**Examples**

```
## Not run:
coefplot(qt1, chr = 1)

## End(Not run)
```

---

colSumsLog

*Sum columns of log transformed data.*


---

**Description**

Given a matrix of log transformed values, sum the rows or columns on the untransformed scale.

**Usage**

```
colSumsLog(logmat)
```

**Arguments**

logmat	Numeric matrix of natural log transformed values.
--------	---

**Details**

See [addLog](#).

**Value**

Numeric vector with values summed on an untransformed scale.

**Author(s)**

Daniel Gatti

**See Also**

[addLog](#)

**Examples**

```
colSumsLog(matrix(log(runif(100)), nrow = 10, ncol = 10))
```

---

condense.model.probs    *Condense 36 state genotypes down to founder genotypes.*

---

**Description**

Additive condenses the heterozygous genotype calls down to the founder allele contributions. Dominance will eventually provide additive and dominance values (method currently uncertain). FALSEull simply gathers all of the genotype probabilities together.

**Usage**

```
condense.model.probs(path = ".", write, model = c("additive", "dominance", "full"))
get.additive(files, samples)
get.dominance(files, samples)
get.full(files, samples)
```

**Arguments**

path	Character containing the path to the *.Rdata files that contain the genotype probabilities.
write	Character that is the filename to write the results to.
model	Character string that is one of "additive", "dominance" or "full". See details.
files	Vector of files to read.
samples	Character vector of sample IDs that match the files argument.

**Details**

get.additive, get.dominance and get.full are helper functions.

**Value**

Three dimensional array of haplotype or genotype probabilities. Num. samples by num. founders (or genotypes) by num. SNPs. Writes out to a \*.Rdata file.

**Author(s)**

Daniel Gatti

**See Also**

[calc.genoprob](#)

**Examples**

```
## Not run: condense.model.probs(write = "model.probs.Rdata")
```

---

convert.allele.calls *Convert allele calls to numeric values.*

---

**Description**

Converts allele calls in A,C,G,T,H,N format into numbers with 0: homozygous A, 1: heterozygous, 2: homozygous B, 3: no call.

**Usage**

```
convert.allele.calls(geno)
```

**Arguments**

geno                    Character matrix containing A, C, G, T, H or N.

**Value**

Numeric matrix containing 0, 1, 2 or 3.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run: convert.allele.calls(geno)
```

```
convert.genes.to.GRanges
```

*Convert MGI genes to GRanges.*

---

**Description**

Given the output of `get.mgi.features`, convert the results to a GRanges object.

**Usage**

```
convert.genes.to.GRanges(mgi)
```

**Arguments**

`mgi` Date.frame as returned by `get.mgi.features`.

**Value**

GRanges object containing the genes in the MGI argument.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run: convert.genes.to.GRanges(mgi)
```

---

```
convert.genotypes
```

*Convert the genotype data from A,C,G,T format to A, H, B, N.*

---

**Description**

Convert the genotype data from A,C,G,T format to A, H, B, N.

**Usage**

```
convert.genotypes(geno)
```

**Arguments**

`geno` Character matrix containing A, C, G, T allele calls.

**Value**

Character matrix containing A, H, B, N.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run: convert.genotypes(geno)
```

---

```
convert.variants.to.GRanges  
  convert.variants.to.GRanges
```

---

**Description**

Given a data.frame of SNPs, convert the SNP locations to a GRanges object.

**Usage**

```
convert.variants.to.GRanges(variants)
```

**Arguments**

variants            Data frame with four header columns and SNPs in the remaining columns.

**Details**

This function creates a GRanges object from the CHR and POS columns of the SNP data.frame.

**Value**

GRanges object containing the SNP locations.

**Author(s)**

Daniel Gatti

**Examples**

```
data(example.snps)  
gr = convert.variants.to.GRanges(example.snps[1:100,])
```

---

```
convert.variants.to.numeric  
  convert.variants.to.numeric
```

---

**Description**

Given a matrix or data frame with character ACGT SNP values, convert them to numeric values with the major allele coded as 0 and the minor allele as 1.

**Usage**

```
convert.variants.to.numeric(variants)
```

**Arguments**

variants            Data frame with four header columns. The SNPs must be in ACGT format in columns 5 through ncol(snp).

**Details**

This function is used before calling plot.alleles() to convert the SNPs into a numeric form suitable for plotting.

**Value**

Data.frame of the same dimensions as the snps argument, but with the alleles converted to 0 or 1.

**Author(s)**

Daniel Gatti

**Examples**

```
data(example.snps)
numeric.snps = convert.variants.to.numeric(example.snps[1:100,])
```

---

```
create.genotype.states
```

*Create genotype states.*

---

**Description**

Given a set of founders, create all of the possible unphased genotype states between them.

**Usage**

```
create.genotype.states(founders)
```

**Arguments**

founders            Character vector of letter codes indicating the founders.

**Details**

Given a set of founder IDs, create all possible unphased genotypes and sort them.

**Value**

Character vector of unphased genotypes that can be created from the given founders.

**Author(s)**

Daniel Gatti

**Examples**

```
create.genotype.states(founders = LETTERS[1:8])
```



---

create.html.page      *Create an HTML QTL report*

---

## Description

Given a DOQTL object, create an HTML page that reports the QTL and creates QTL plots. Permutations for assessing significance thresholds can be supplied.

## Usage

```
create.html.page(path, qtl, pheno.name, perms, assoc)
```

## Arguments

path	Character string containing the path to which to write the report.
qtl	DOQTL object containing a list with two elements: lod and coef.
pheno.name	Character string containing the phenotype name.
perms	Numeric vector containing the permutation LOD scores for this phenotype.
assoc	Boolean, if TRUE, look for corresponding *.Rdata files containing the names of the qtl in the current working directory and plot the association plots. If FALSE (default), do not plot association analysis.

## Details

The function creates an HTML page with a QTL plot, a table of significant QTL and coefficient plots for the significant loci.

## Value

Data.frame with the significant QTL from this phenotype.

## Author(s)

Daniel Gatti

## See Also

[html.report](#)

## Examples

```
## Not run: create.html.page(path, qtl, pheno.name, perms)
```

---

```
create.Rdata.files      Convert *.txt files to *.Rdata files.
```

---

**Description**

This is used to convert the \*.txt genotype probability files to \*.Rdata files.

**Usage**

```
create.Rdata.files(prob.files)
```

**Arguments**

prob.files Character vector containing genotype probability file names.

**Value**

No value returned. Write out a \*.Rdata file for each \*.txt file.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run: create.Rdata.files(prob.files)
```

---

```
do.colors      do.colors
```

---

**Description**

The letter codes, strain names and official colors for the DO founders.

**Usage**

```
do.colors
```

**Format**

A data frame with 8 rows and 3 columns.

CC\_Designation a factor with levels A B C D E FALSE G H

Strain a factor with levels 129S1/SvImJ A/J C57BL/6J CAST/EiJ NOD/ShiLtJ NZO/H1LtJ PWK/PhJ  
WSB/EiJ

R\_Color a factor with levels #00A000 #00A0FALSE0 #1010FALSE0 #808080 #9000FALSE4 #FALSE00000  
#FALSE08080 #FALSE0FALSE000

**Details**

This contains the official colors that should be used when plotting data involving the DO founders.

**Source**

Copied from [UNC Systems Genetics Website](#)

**Examples**

```
do.colors
```

---

do.states	<i>do.states</i>
-----------	------------------

---

**Description**

The 36 unphased genotype states for the DO on the autosomes and X chromosome. Also the 8 DO founder letter codes.

**Usage**

```
do.states
```

**Format**

A list frame with 3 elements.

auto Character vector with two letter codes for each of the possible DO genotype states.

X List with two elements, FALSE and M, containing the genotype codes for the X chromosome.

founders Character vector containing the founder letter codes.

**Details**

This contains the letter codes for each of the 36 unphased genotype states in the DO. It also contains the founder letters.

**Examples**

```
do.states
```

---

do.trans.probs	<i>Determine DO transition probabilities</i>
----------------	--

---

**Description**

Determine the genotype state transition probability for DO mice of a specific generation between all of the markers on a given chromosome.

**Usage**

```
do.trans.probs(states, snps, chr = c(1:19, "X"), sex = c("M", "F"), do.gen)
```

**Arguments**

states	Character vector of possible genotype states.
snps	Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively.
chr	Character containing the chromosome for which transition probabilities should be calculated.
sex	Character that is one of FALSE or M, indicating the sex to use on the X chromosome.
do.gen	Number vector indicating the DO outbreeding generations to calculate.

**Details**

This function is used to calculate the transition probabilities between markers for different DO outbreeding generations.

**Value**

List containing one element per unique DO generation supplied in the do.gen argument. Each list element contains a 3 dimensional array of transition probabilities between each pair of markers (num.states by num.states by num.markers - 1).

**Author(s)**

Daniel Gatti and Karl Broman

**References**

Haplotype probabilities in advanced intercross populations. Broman KW. G3 (Bethesda). 2012 FALSEeb;2(2):199-202. doi: 10.1534/g3.111.001818. Epub 2012 FALSEeb 1. PMID: 22384398  
 Genotype probabilities at intermediate generations in the construction of recombinant inbred lines. Broman KW. Genetics. 2012 FALSEeb;190(2):403-12. doi: 10.1534/genetics.111.132647. PMID: 22345609

**Examples**

```
## Not run: do.trans.probs(states, snps, chr = c(1:19, "X"), sex = c("M", F), do.gen)
```

---

do2sanger

*Impute the Sanger SNPs onto DO genomes*

---

**Description**

Given a set of DO genotype probability files and the location of the Tabix indexed Sanger file, impute the Sanger SNPs on to DO genomes.

**Usage**

```
do2sanger(do.files, snps, output.file = "do2sanger.txt", snp.file =
"ftp://ftp.jax.org/SNPtools/variants/cc.snps.NCBI38.txt.gz")
```

## Arguments

do.files	Character vector of *.genotype.probs.Rdata files that contain the posterior probabilities.
snps	Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively.
output.file	Character string to write the results to.
snp.file	Character string with path to a Tabix indexed SNP file. Default is from <a href="#">JAX FALSETP site</a>

## Details

We read in a single genotype probability file, which must have been saved as a \*.Rdata file. The format is a matrix with markers in rows and states in columns and dimnames for both. FALSEor each pair of markers, we take the average genotype probability. Then we take the DO genotype with the highest probability and split it into the two founder haplotypes. We get the Sanger SNPs for each of the two founders, convert them to 1, 1 or 2 and insert them into the DO sample.

## Value

No value is returned. The Sanger SNPs mapped onto the DO genomes are written out to the output.file. The file will contain 0, 1 or 2 as the allele calls.

## Author(s)

Daniel Gatti

## References

Combined sequence-based and genetic mapping analysis of complex traits in outbred rats. Rat Genome Sequencing and Mapping Consortium, Baud A, Hermesen R, Guryev V, Stridh P, Graham D, McBride MW, FALSEoroud T, Calderari S, Diez M, Ockinger J, Beyeen AD, Gillett A, Abdelmagid N, Guerreiro-Cacais AO, Jagodic M, Tuncel J, Norin U, Beattie E, Huynh N, Miller WH, Koller DL, Alam I, FALSEalak S, Osborne-Pellegrin M, Martinez-Membrives E, Canete T, Blazquez G, Vicens-Costa E, Mont-Cardona C, Diaz-Moran S, Tobena A, Hummel O, Zelenika D, Saar K, Patone G, Bauerfeind A, Bihoreau MT, Heinig M, Lee YA, Rintisch C, Schulz H, Wheeler DA, Worley KC, Muzny DM, Gibbs RA, Lathrop M, Lansu N, Toonen P, Ruzius FALSEP, de Bruijn E, Hauser H, Adams DJ, Keane T, Atanur SS, Aitman TJ, FALSElicek P, Malinauskas T, Jones EY, Ekman D, Lopez-Aumatell R, Dominiczak AFALSE, Johannesson M, Holmdahl R, Olsson T, Gauguier D, Hubner N, FALSEernandez-Teruel A, Cuppen E, Mott R, FALSElint J. *Nat Genet.* 2013 Jul;45(7):767-75. doi: 10.1038/ng.2644. Epub 2013 May 26. PMID: 23708188 Using progenitor strain information to identify quantitative trait nucleotides in outbred mice. Yalcin B, FALSElint J, Mott R. *Genetics.* 2005 Oct;171(2):673-81. Epub 2005 Aug 5. PMID: 16085706 Mouse genomic variation and its effect on phenotypes and gene regulation. Keane TM, Goodstadt L, Danecek P, White MA, Wong K, Yalcin B, Heger A, Agam A, Slater G, Goodson M, FALSEurlotte NA, Eskin E, Nellaker C, Whitley H, Cleak J, Janowitz D, Hernandez-Pliego P, Edwards A, Belgard TG, Oliver PL, McIntyre RE, Bhomra A, Nicod J, Gan X, Yuan W, van der Weyden L, Steward CA, Bala S, Stalker J, Mott R, Durbin R, Jackson IJ, Czechanski A, Guerra-Assuncao JA, Donahue LR, Reinholdt LG, Payseur BA, Ponting CP, Birney E, FALSElint J and Adams DJ *Nature* 2011;477;7364;289-94 PUBMED: 21921910 Sequence-based characterization of structural variation in the mouse genome. Yalcin B, Wong K, Agam A, Goodson M, Keane TM, Gan X, Nellaker C, Goodstadt L, Nicod J, Bhomra A, Hernandez-Pliego P, Whitley H, Cleak J, Dutton R, Janowitz D, Mott R, Adams DJ and FALSElint J *Nature* 2011;477;7364;326-9 PUBMED: 21921916

**See Also**

[assoc.map](#)

**Examples**

```
## Not run: do2sanger(do.files, snps, output.file = "do2sanger.txt",
  snp.file = "ftp://ftp.jax.org/SNPtools/variants/cc.snps.NCBI38.txt.gz")
## End(Not run)
```

---

estimate.cluster.params

*Estimate genotype cluster means and variances*

---

**Description**

Given the X and Y intensity data, perform model based clustering and estimate the genotype state cluster means and variances.

**Usage**

```
estimate.cluster.params(founders, data, chr)
keep.homozygotes(founders)
```

**Arguments**

- |          |   |
|----------|---|
| founders | List containing founder information for non-DO or CC crosses. <i>Not required for DO.</i><br><br>When method = intensity: x: Numeric matrix, num.samples x num.snps, with X intensities for all founders and FALSEs (if available). Sample IDs and SNP IDs must be in rownames and colnames. y: Numeric matrix, num.samples x num.snps, with Y intensities for all founders and FALSEs (if available). Sample IDs and SNP IDs must be in rownames and colnames. sex: Character vector, containing "M" or F indicating sex. Sample IDs must be in names. code: Character vector containing two letter genotype codes for each founder sample. Sample IDs must be in names.   |
| data     | A list with named elements containing the information needed to reconstruct genomes.<br><br>When method = intensity: x: Numeric matrix, num.samples x num.snps, with X intensities for all samples. Sample IDs and SNP IDs must be in rownames and colnames. y: Numeric matrix, num.samples x num.snps, with Y intensities for all samples. Sample IDs and SNP IDs must be in rownames and colnames. sex: Character vector, containing "M" or F indicating sex. Sample IDs must be in names. gen: Character matrix containing the generation of DO outbreeding for each sample. FALSE or the DO, this should be "DO" followed by a number with no space between them. FALSE or CC mice, this should be CC. Sample IDs must be in names. |
| chr      | Character containing the current chromosome.  |

**Details**

At each marker, use `mclust` to perform model based clustering on all of the data and get estimates of the means and variances for each cluster. Then assign each of the 36 genotype states to the nearest founder cluster.

`keep.homozygotes` is an internal helper function.

**Value**

List containing two elements:

`r.t.means`        Three dimensional array containing rho and theta genotype cluster means.

`r.t.covars`      Three dimensional array containing rho and theta genotype cluster variances.

**Author(s)**

Daniel Gatti

**See Also**

[hmm.intensity](#)

**Examples**

```
## Not run: estimate.cluster.params(founders, data, chr)
```

---

example.genes

*example.genes*

---

**Description**

A set of genes from the Mouse Genome Informatics that are used in the documentation examples. (<http://www.sanger.ac.uk/resources/mouse/genomes/>) FALSErom Chr 7: 103 - 105 Mb on NCBI Build 38.

**Source**

<http://informatics.jax.org/>

**Examples**

```
data(example.genes)
```

---

example.pheno	<i>Example phenotypes.</i>
---------------	----------------------------

---

**Description**

Example phenotypes.

**Usage**

```
example.pheno
```

**Format**

A data frame with 149 observations on the following 8 variables. Sample IDs in rownames and phenotype names in colnames.

Sample Character vector containing sample IDs.

Sex Character vector containing the sex of each animal.

Gen Character vector containing DO outbreeding generation and litter.

Diet Character vector containing diet, either chow or hf for high fat.

Coat.Color Character vector containing text description of mouse colors.

albino Numeric vector containing 1 if the mouse's coat color was white.

black Numeric vector containing 1 if the mouse's coat color was black.

HDL2 Numeric vector of high density lipoprotein values.

**Details**

Data from Svenson et.al. paper below.

**References**

High-resolution genetic mapping using the Mouse Diversity outbred population. Svenson KL, Gatti DM, Valdar W, Welsh CE, Cheng R, Chesler EJ, Palmer AA, McMillan L, Churchill GA. *Genetics*. 2012 Feb;190(2):437-47 PMID: 223445611

**Examples**

```
head(example.pheno)
```



---

`example.qtl`*Example QTL.*

---

### Description

Example QTL for the albino and HDLD2 traits in [example.pheno](#). Albino is a binary, Mendelian trait that maps to the Tyrosinase locus on Chr 7. HDLD2 (high density lipoprotein) is a complex trait with many loci.

### Usage

```
example.qtl
```

### Format

A data frame with 149 observations on the following 8 variables. Sample IDs in rownames and phenotype names in colnames.

Sample Character vector containing sample IDs.

Sex Character vector containing the sex of each animal.

Gen Character vector containing DO outbreeding generation and litter.

Diet Character vector containing diet, either chow or hf for high fat.

Coat.Color Character vector containing text description of mouse colors.

albino Numeric vector contining 1 if the mouse's coat color was white.

black Numeric vector contining 1 if the mouse's coat color was black.

HDLD2 Numeric vector of high density lipoprotein values.

### Details

Data from Svenson et.al. paper below.

### References

High-resolution genetic mapping using the Mouse Diversity outbred population. Svenson KL, Gatti DM, Valdar W, Welsh CE, Cheng R, Chesler EJ, Palmer AA, McMillan L, Churchill GA. *Genetics*. 2012 FALSEeb;190(2):437-47 PMID: 223445611

### Examples

```
names(example.qtl)
names(example.qtl[[1]])
```

---

example.snps	<i>example.snps</i>
--------------	---------------------

---

### Description

A set of SNPs from the Sanger Mouse Genome project that are used in the documentation examples. (<http://www.sanger.ac.uk/resources/mouse/genomes/>) FALSErom Chr 7: 103 - 105 Mb on NCBI Build 38.

### Source

<http://www.sanger.ac.uk/resources/mouse/genomes/>

### Examples

```
data(example.snps)
```

---

extract.raw.data	<i>Extract intensities, genotypes and call rates from from raw MUGA or MegaMUGA data files</i>
------------------	--

---

### Description

This function accepts a vector of input directories containing the raw MUGA or MegaMUGA raw data files. FALSEor each directory, the function reads the X and Y intensities, call rates and allele calls for all samples. It then combines all samples and writes the data to "x.txt", "y.txt", "geno.txt" and "call.rate.batch.txt" in the user specified output directory.

### Usage

```
extract.raw.data(in.path = ".", prefix, out.path = ".", array = c("megamuga", "muga"))
```

### Arguments

in.path	character vector, the full path to all MUGA directories from which data should be extracted.
prefix	character vector of same length as in.path containing a prefix to add to each sample ID in data sets being processed.
out.path	character, the full path to the directory where the output files should be written.
array	character, default = "megamuga", the type of array, either "muga" or "megamuga".

### Details

This function searches each directory for files with names containing "Sample\_Map.txt" and "\*\_FALSEinalReport.txt". This has been the format that GeneSeek has consistently produced. The call rates are extracted and are written, along with a batch ID, to "call.rate.batch.txt". The X and Y intensities are extracted from the "FALSEinalReport" file and written to "x.txt" and "y.txt" respectively. The allele calls are extracted from the "FALSEinalReport" files and are written to "geno.txt". The prefix argument may be used to add a prefix to the sample IDs in order to distinguish different data sets.

**Value**

No return value. The files are written to the out.path directory.

**Note**

Do not change the names of the output files. They are required for downstream processing.

**Author(s)**

Daniel M. Gatti

**See Also**

[filter.samples](#), [batch.normalize](#)

**Examples**

```
## Not run:
in.path = c("/tmpdir/DataSet1", "/tmpdir/DataSet2")
extract.raw.data(in.path = in.path, prefix = c("ds1", "ds2"),
  out.path = "/tmpdir/output", array = "muga")

## End(Not run)
```

---

fast.qtlrel

*QTL mapping using QTLRel*


---

**Description**

This extracts teh core of the QTLRel algorithm for additive covariates.

**Usage**

```
fast.qtlrel(pheno, probs, K, addcovar, snps)
```

**Arguments**

pheno	Data.frame containing the phenotype data. Sample IDs in rownames.
probs	Three dimensional numeric array containing the founder haplotype contributions. Num.samples by num.founder by num.markers. Sample IDs, founder letters and SNP IDs must be in dimnames.
K	Numeric matrix containing the kinship between samlpes. Sample IDs must be in rownames and colnames.
addcovar	Numeric matrix containing additive covariates to run in the mapping model. Sample IDs must be in rownames.
snps	Data.frame containing the marker locations. SNP ID, chromosome, Mb anc cM locations in columns 1 through 4, respectively.

**Details**

We extracted code from QTLRel, but removed several options to speed up the pipeline for QTL mapping with additive covariates and a kinship matrix.

**Value**

List containing two elements:

- lod                    List containing two elements: A: Numeric matrix containing the SNP ID, chromosome, Mb, cM, percent variance explained, likelihood ratio statistic, log of the odd ratio, p-value and  $-\log_{10}(\text{p-value})$  for autosomes. X: Numeric matrix containing the SNP ID, chromosome, Mb, cM, percent variance explained, likelihood ratio statistic, log of the odd ratio, p-value and  $-\log_{10}(\text{p-value})$  for X chromosome.
- coef                    List containing two elements: A : Numeric matrix containing QTL model coefficients on autosomes. Markers in rows. X : List containing two elements for the X chromosome: FALSE : Numeric matrix containing QTL model coefficients for females. Markers in rows. M : Numeric matrix containing QTL model coefficients for males. Markers in rows.

**Author(s)**

Daniel Gatti

**References**

Cheng R, Abney M, Palmer AA, Skol AD. QTLRel: an R package for genome-wide association studies in which relatedness is a concern. BMC Genet. 2011 Jul 27;12:66.

**See Also**

[scanone](#), [scanone](#)

**Examples**

```
## Not run: fast.qtlrel(pheno, probs, K, addcovar, snps)
```

---

fill.in.snps

*Interpolate between SNPs at the same cM value.*

---

**Description**

Go through the SNPs and look for stretches where the differences in cM values from one SNP to the next equals 0. Interpolate from the first SNP to the last SNP in such a stretch.

**Usage**

```
fill.in.snps(snps)
```

**Arguments**

snps                    Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively.

**Details**

On each chromosome, look for sets of contiguous markers that have the same cM location. Interpolate evenly spaced markers spanning the markers proximal and distal to this set.

**Value**

Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run: fill.in.snps(snps)
```

---

filter.geno.probs	<i>Remove SNPs where the genotype probabilities are too low for one founder state</i>
-------------------	---

---

**Description**

This function accepts the 8 state founder probabilities and searches for SNPs where the founder probabilities are too low for one founder. It removes these SNPs. The QTL mapping model becomes numerically unstable if these SNPs are not removed.

**Usage**

```
filter.geno.probs(geno)
```

**Arguments**

geno                    Numeric 3D array, with samples in dim[1], states = dim[2] and SNPs in dim[3].

**Value**

Numeric 3D array, with samples in dim[1], states = dim[2] and SNPs in dim[3]. The SNPs with low probabilities for a single founder have been removed.

**Author(s)**

Daniel Gatti

**See Also**

[scanone](#), [scanone.perm](#)

**Examples**

```
## Not run:
  filter.geno.probs(geno)

## End(Not run)
```

---

filter.samples	<i>FALSE</i> ilter X, Y and genotype data by call rate
----------------	--

---

### Description

This function reads in "x.txt", "y.txt", "geno.txt" and "call.rate.batch.txt" from the user specified input directory and removes samples with a call rate less than the supplied threshold (default = 0.9). It then writes the files out to "x.filt.txt", "y.filt.txt", "geno.filt.txt" and "call.rate.batch.filt.txt".

### Usage

```
filter.samples(path = ".", thr = 0.9)
```

### Arguments

path	Character, the full path to the directory where the files reside.
thr	Numeric, call rate threshold below which samples will be removed. Default = 0.9.

### Value

data.frame with the sample IDs and call rates of the removed samples. The intensity and genotype files are written to the input directory.

### Author(s)

Daniel Gatti

### See Also

[extract.raw.data](#), [batch.normalize](#)

### Examples

```
## Not run:
  filter.samples(path = "/tmpdir/output")

## End(Not run)
```

---

find.overlapping.genes	<i>find.overlapping.genes</i>
------------------------	-------------------------------

---

### Description

FALSEind genes that intersect with a given set of variants.

**Usage**

```
find.overlapping.genes(variants,
  mgi.file = "http://cgd.jax.org/tools/SNPtools/MGI/MGI.20130305.sorted.txt.gz",
  type = c("gene", "exon"))
```

**Arguments**

variants	Data.frame with variants. The type attribute must be set to one of "snp", "indel", or "sv".
mgi.file	Character. FALSEull path to the MGI gene file. Defaults to "http://cgd.jax.org/tools/SNPtools/MGI/MGI.20130305.sorted.txt.gz".
type	Character. One of "gene" or "exon". Indicates whether to intersect SNPs with genes (from beginning to end, including introns) or exons only.

**Details**

Gets the gene locations from MGI and variants.

**Value**

Data.frame with gene locations and symbols.

**Author(s)**

Daniel Gatti

**See Also**

[get.mgi.features](#){get.mgi.features}

**Examples**

```
## Not run:
data(example.snps)
x = find.overlapping.genes(variants = example.snps[19478:19506,])

## End(Not run)
```

---

gene.plot

*gene.plot*

---

**Description**

Given a genomic region, plot the genes in the interval. `line.up.genes` and `resolve.collisions` are internal functions.

**Usage**

```
gene.plot(mgi, rect.col = "grey30", text.col = "black", ...)
```

**Arguments**

<code>mgi</code>	Data.frame with MGI gene locations as returned by <code>get.mgi.features{get.mgi.features}</code> .
<code>rect.col</code>	Color vector that is the color to use to plot gene rectangles. May be a single color for all genes or a vector with colors for each gene. Default = "grey30".
<code>text.col</code>	Color vector that is the color to use to plot gene names. May be a single color for all genes or a vector with colors for each gene. Default = "black".
<code>...</code>	Other arguments to be passed into plot.

**Details**

The spacing algorithm attempts to organize the genes in such a way that they do not collide. The `rect.col` and `text.col` arguments are recycled if they are shorter than the number of genes. They can be used to highlight specific genes.

**Value**

Data.frame with gene locations and symbols.

**Author(s)**

Daniel Gatti

**See Also**

`get.mgi.features{get.mgi.features}`

**Examples**

```
## Not run:
data(example.genes)
g = gene.plot(mgi = example.genes)

## End(Not run)
```

---

`generic.trans.probs`    *Generic transition probabilities*

---

**Description**

Generic function to provide starting transition probabilities. Not ready for use in this version.

**Usage**

```
generic.trans.probs(states, snps, chr = c(1:19, "X"), sex = c("M", "F"))
```

**Arguments**

<code>states</code>	Character vector containing the genotype states coded as letters.
<code>snps</code>	Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively.
<code>chr</code>	Character containing the chromosome to use.
<code>sex</code>	Character that is either FALSE or M indicating the sex. Only used on x chromosome.



**Details**

This function is a placeholder for a more sophisticated algorithm to be developed later. It is an effort to provide a set of starting transition probabilities for mapping populations other than the DO.

**Value**

A matrix of transition probabilities between genotype states.

**Note**

This function is still in development.

**Author(s)**

Daniel Gatti

**See Also**

[do.trans.probs](#), [cc.trans.probs](#)

**Examples**

```
## Not run: generic.trans.probs(states, snps, chr = 1, sex = "M")
```

---

genome.summary.plots    *Genome summary plots*

---

**Description**

These plots show summaries of the founder haplotype proportions across SNPs or samples.

**Usage**

```
genotype.by.sample.barplot(results)  
genotype.by.snp.barplot(results, snps)
```

**Arguments**

results	Data.frame as produced by <a href="#">summarize.by.snps</a> or <a href="#">summarize.by.samples</a> .
snps	Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively.

**Details**

Barplots across samples or SNPs are produced from the relevant summary files. This is intended to be run after DO genomes have been reconstructed for a large set of samples to verify that founder allele frequencies are consistent across SNPs and samples.

**Author(s)**

Daniel Gatti

**See Also**

[summarize.by.snps](#), [summarize.by.samples](#)

**Examples**

```
## Not run:  
load(url("ftp://ftp.jax.org/MUGA/muga_snps.Rdata"))  
results = summarize.by.samples(path = ".", snps = muga_snps)  
genotype.by.sample.barplot(results)  
  
## End(Not run)
```

---

get.chr.lengths

*Get chromosome lengths for the mouse*

---

**Description**

Using the org.Mm.eg.db package, get the chromosome lengths.

**Usage**

```
get.chr.lengths()
```

**Details**

Keeps only the lengths of the chromosomes, not random or unmapped.

**Value**

Named vector of chromosome lengths.

**Author(s)**

Daniel Gatti

**Examples**

```
get.chr.lengths()
```

---

get.do.states	<i>Get the 36 genotype states for the DO</i>
---------------	--

---

**Description**

Get the 36 genotype states for the DO

**Usage**

```
get.do.states()
```

**Value**

Character vector containing the two letter genotype codes for the 36 genotype states.

**Author(s)**

Daniel Gatti

**Examples**

```
get.do.states()
```

---

get.gene.name	<i>Get the gene symbol</i>
---------------	----------------------------

---

**Description**

Given an MGI ID, get the gene symbol from an MGI data.frame.

**Usage**

```
get.gene.name(value, mgi)
```

**Arguments**

value	Character string containing an MGI ID.
mgi	Data.frame containing MGI data from <a href="#">get.mgi.features</a> .

**Value**

Character string containing the gene symbol.

**Author(s)**

Daniel Gatti

**See Also**

[get.mgi.features](#)

**Examples**

```
## Not run: get.gene.name(value, mgi)
```

---

```
get.machine.precision Get the machine precision
```

---

**Description**

Get the machine precision from `.Machine$double.eps` on a log10 scale.

**Usage**

```
get.machine.precision()
```

**Value**

Numeric value containing the machine precision.

**Author(s)**

Daniel Gatti

**Examples**

```
get.machine.precision()
```

---

```
get.max.geno Get the genotype with the highest probability
```

---

**Description**

FALSE or each sample at each marker, return the genotype state with the highest probability.

**Usage**

```
get.max.geno(probs)
```

**Arguments**

`probs` Three dimensional numeric matrix containing the founder haplotype contributions.

**Details**

If the maximum probability is greater than 0.75, it is called homozygous. Otherwise, we take the two highest probabilities and call heterozygous.

**Value**

Character matrix containing the genotype with the highest probability at each locus.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run: get.max.geno(probs)
```

---

```
get.mgi.features      get.mgi.features
```

---

**Description**

Retrieve the MGI features within a genomic region. FALSE features include genes, gene models, non-coding RNA, etc., but not SNPs. This allows the user to filter by source and type of feature.

**Usage**

```
get.mgi.features(file = "ftp://ftp.jax.org/SNPtools/genes/MGI.20130703.sorted.txt.gz",
  chr = NULL, start = NULL, end = NULL, source = c("all", "MGI", "VEGA", "ENSEMBL",
  "Blat", "NCBI_Gene"), type = c("all", "gene", "pseudogenic_transcript",
  "pseudogenic_exon", "pseudogene", "match", "match-part", "transcript", "exon",
  "mRNA", "five_prime_UTR", "start_codon", "CDS", "stop_codon", "three_prime_UTR",
  "pseudogenic_mRNA", "pseudogenic_start_codon", "pseudogenic_CDS",
  "pseudogenic_stop_codon", "pseudogenic_five_prime_UTR", "pseudogenic_three_prime_UTR",
  "sequence_feature"))
```

**Arguments**

file	character, the full path to the Tabix indexed and zipped MGI feature file. Default = "http://cgd.jax.org/tools/SNPtools/MGI/MGI.20130305.sorted.txt.gz" for internal JAX use.
chr	Numeric vector, chr for each start and end position. Chr, start and end must all have the same length.
start	Numeric vector, start position in Mb or bp for each chr. Chr, start and end must all have the same length.
end	Numeric vector, end position in Mb or bp for each chr. Must be greater than or equal to the corresponding start value. Chr, start and end must all have the same length.
source	Character vector, the source of the annotation. Options are ("all", "MGI", "VEGA", "ENSEMBL", "Blat", "NCBI_Gene"). "all" returns all features.
type	Character vector, the type of feature. Options are ("all", "gene", "pseudogenic_transcript", "pseudogenic_exon", "pseudogene", "match", "match-part", "transcript", "exon", "mRNA", "five_prime_UTR", "start_codon", "CDS", "stop_codon", "three_prime_UTR", "pseudogenic_mRNA", "pseudogenic_start_codon", "pseudogenic_CDS", "pseudogenic_stop_codon", "pseudogenic_five_prime_UTR", "pseudogenic_three_prime_UTR", "sequence_feature"). "all" returns all features.

**Details**

This function is designed to return features from the MGI gene feature file (GFALSEFALSE). You can select multiple regions on different chromosomes.

**Value**

A list of data.frames for each region requested. Each data.frame will contain 14 columns; seqid, source, type, start, stop, score, strand, phase, ID, Name, Parent, Dbxref, mgiName, bioType. If there is only one requested region, a single data frame is returned.

**Author(s)**

Daniel Gatti

**References**

The MGI GFALSEFALSE file is at <ftp://ftp.informatics.jax.org/pub/mgigff/>.

**Examples**

```
## Not run:
genes = get.mgi.features(chr = 7, start = 103 end = 105 source = "MGI", type = "gene")

## End(Not run)
```

---

get.num.auto

*Get the number of autosomes*

---

**Description**

Given a data.frame of SNPs, return the number of autosomes by counting the number of numeric chromosomes.

**Usage**

```
get.num.auto(snps)
```

**Arguments**

snps	Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively.
------	---

**Value**

Numeric value containing the number of autosomes.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run: get.num.auto(snps)
```

---

get.pattern.variants    *get.pattern.variants*

---

## Description

Given a vector of strains names and a set of variants, return the variants for which the given strains have one allele and the remaining strains have the other allele.

## Usage

```
get.pattern.variants(variants, strain.subset = NULL)
```

## Arguments

`variants`            Matrix of data.frame of variants with four header columns containing annotation and snps in the remaining columns.

`strain.subset`    character vector of strain names that comprise one subset.

## Details

Use this function to obtain a subset of SNPs for which one set of strains contain one allele and the rest contain the other allele. This might come up in the course of QTL mapping where several strains have a high allele and others have a low allele.

## Value

Data.frame of variants that match the requested pattern.

## Author(s)

Daniel Gatti

## See Also

[get.variants](#)

## Examples

```
## Not run:
vcf = readVcf("vcf_file.tar.gz", param)
snp.subset = get.pattern.variants(vcf,
  strain.subset = c("A/J", "NOD/ShiLtJ", "NZO/H1LtJ", "AKR/J"))

## End(Not run)
```

---

 get.strains

*get.strains*


---

### Description

Get available strain names for a set of strains from a variant file.

### Usage

```
get.strains(
  file = "http://cgd.jax.org/tools/SNPtools/Build38/sanger.snps.NCBI38.txt.gz")
```

### Arguments

`file` Character, full path to the variants file to use. Default is the file at the Center for Genome Dynamics at The Jackson Laboratory.

### Value

Character vector of strain names in the variant file.

### Author(s)

Daniel Gatti

### Examples

```
## Not run:
available.strains = get.strains()
strains = available.strains[c(2, 5, 8, 9, 13:15, 17)]
snps = get.variants(chr = 7, start = 103, end = 105, strains = strains)

## End(Not run)
```

---

 get.trans.probs

*Get the transition probabilities between markers.*


---

### Description

Based on the DO founders and the recombination fraction, calculate the transition probability between markers on one chromosome.

### Usage

```
get.trans.probs(r, do.gen, alpha, chr = c(1:19, "X"), sex = c("M", "F"))
```



**Arguments**

r	Double vector of recombination fractions between SNPs.
do.gen	Integer containing the outbreeding generation of DO.
alpha	Double vector, proportion of preCC progenitors at generation k. Generation numbers in the names.
chr	Character, one of 1:19, X.
sex	Character, either M or FALSE. Only used on X chromosome.

**Value**

Numeric three dimensional array of transition probabilities between each pair of markers (num.states by num.states by num.markers - 1).

**Author(s)**

Daniel Gatti

**References**

Haplotype probabilities in advanced intercross populations. Broman KW. G3 (Bethesda). 2012 FALSEeb;2(2):199-202. doi: 10.1534/g3.111.001818. Epub 2012 FALSEeb 1. PMID: 22384398  
 Genotype probabilities at intermediate generations in the construction of recombinant inbred lines. Broman KW. Genetics. 2012 FALSEeb;190(2):403-12. doi: 10.1534/genetics.111.132647. PMID: 22345609

**Examples**

```
## Not run: get.trans.probs(r, do.gen, alpha, chr = c(1:19, "X"), sex = c("M", "F"))
```

---

get.variants	<i>get.variants</i>
--------------	---------------------

---

**Description**

Get SNPs for a set of strains for a specific regions of chromosomes.

**Usage**

```
get.variants(file = "ftp://ftp.jax.org/SNPtools/variants/cc.snps.NCBI38.txt.gz",
  chr, start, end, type = c("snp", "indel", "sv"), strains, polymorphic = TRUE,
  quality)
```

**Arguments**

file	Character, full path to the SNP file to use. Default is the file at the Center for Genome Dynamics at The Jackson Laboratory.
chr	Numeric vector, chr for each start and end position. Chr, start and end must all have the same length.
start	Numeric vector, start position in Mb or bp for each chr. Chr, start and end must all have the same length.

end	Numeric vector, end position in Mb or bp for each chr. Must be greater than or equal to the corresponding start value. Chr, start and end must all have the same length.
type	Character, with one of "snp", "indel", "sv". Indicates the type of variantes being queried.
strains	Character vector, listing the strains to retrieve. The names can be obtained from <code>get.strains()</code> . Default returns all strains.
polymorphic	Boolean. Default = TRUE. If TRUE, retrieve only SNPs that are polymorphic among the requested strains. If FALSE, return all SNPs in the requested interval(s).
quality	Integer, denoting the confidence levels to return. This looks at the quality column of all requested strains and takes values greater than or equal to the given number. Look at the documentation for the SNP file you are using for details on the quality score. Default returns all SNPs.

### Value

A list of data.frames for each region requested. Each data.frame will contain five header columns; ID, CHROM, POS, REF & ALT corresponding to the SNP ID, chromosome, bp position, reference and alternate alleles. There follows two columns for each requested strain containing the allele calls and quality scores for each strain. If there is only one requested region, a single data frame is returned.

### Author(s)

Daniel Gatti

### See Also

[get.strains](#)

### Examples

```
## Not run:
available.strains = get.strains()
strains = available.strains[c(2, 5, 8, 9, 13:15, 17)]
snps = get.variants(chr = 7, start = 103, end = 105, strains = strains)

## End(Not run)
```

---

html.report

*Create an HTML report for a set of QTL*

---

### Description

Given a list of QTL objects, create a set of HTML pages summarizing the QTL.

### Usage

```
html.report(path, qtl, perms, assoc = FALSE)
```

**Arguments**

path	Character string with path to which to write.
qtl	List containing DOQTL objects as produced by <code>scanone</code> .
perms	Numeric vector containing permutation results for the phenotypes.
assoc	Boolean, if TRUE, look for corresponding *.Rdata files containing the names of the qtl in the current working directory and plot the association plots. If FALSE (default), do not plot association mapping analysis.

**Details**

This function summarizes a set of QTL scans made using `scanone`. For each QTL in the qtl list, it creates an HTML page with a QTL plot and coefficient plots for the significant QTL. The permutations are used to assess significance. It then summarizes all of these in a table and creates a summary HTML page and a \*.csv file with the QTL results.

**Value**

No value is returned. HTML report of QTL is written out to the specified directory.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run: html.report(path, qtl, perms, merge = FALSE)
```

---

intensity.plots	<i>Plot founders and F1 hybrids or genotype state means and variances on an intensity plot.</i>
-----------------	---

---

**Description**

Given the X and Y (or rho and theta) array intensities, plot the values for one SNP and color and mark the founders and FALSEs.

**Usage**

```
founder.F1.intensity.plot(theta, rho, s = 1, is.founder.F1, plotNew = TRUE, ...)
intensity.mean.covar.plot(s, states, theta, rho, r.t.means, r.t.covars, sample, ...)
```

**Arguments**

theta	Numeric matrix containing theta (or X) intensity values.
rho	Numeric matrix containing rho (or Y) intensity values.
s	Numeric integer containing the SNP index in rho and theta to plot.
is.founder.F1	Boolean vector containing TRUE for samples that are DO founders or FALSEs and FALSE for those that are not.
plotNew	Boolean that is TRUE if a new plot should be made and FALSE if the points should be added to the existing plot.

states	Character vector containing genotypes state letter codes.
r.t.means	Numeric three dimensional array containing genotype state means in rho and theta coordinates.
r.t.covars	Numeric three dimensional array containing genotype state variances in rho and theta coordinates.
sample	Numeric value containing the sample index into the rho and theta matrices to plot in orange.
...	Additional arguments to be passed to plot.

**Value**

No return value. A scatter plot is made of the rho and theta (or X and Y) intensities at the requested SNP. Founders and F1 hybrid samples are colored.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run: founder.F1.intensity.plot(theta, rho, s = 1, is.founder.F1, plotNew = TRUE, ...)
```

---

kinship.probs	<i>Create a kinship matrix.</i>
---------------	---------------------------------

---

**Description**

Read in the genotypes and produce a kinship matrix based on either allele sharing or haplotype contributions.

**Usage**

```
kinship.probs(probs, snps, bychr = FALSE)
kinship.alleles(geno)
```

**Arguments**

probs	Three dimensional numeric array containing the founder haplotype contributions. Num.samples by num.founder by num.markers.
snps	Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively.
bychr	Boolean that is true if the current chromosome should be excluded from the calculation of kinship for the current chromosome. See Details.
geno	Character matrix of allele calls.

**Details**

Allele based method: This simply calculates the mean allele sharing between two individuals. Intensity based method: FALSEor each pair of samples, at each marker, calculate the cosine of the angle between the two founder contribution vectors. Take the average across the genome.

When bychr = FALSE, we calculate one kinship matrix for all chromosomes. When bychr TRUE, we calculate a different kinship matrix for each chromosome. FALSEor each chromosome, we remove use only the SNPs on the remaining chromosomes to calculate the kinship matrix for that chromosome. This is motivated by the Cheng et.al. reference below.

**Value**

Numeric matrix, with rows and columns equal to the number of samples in the genotype file.

**Author(s)**

Daniel Gatti

**References**

Practical Considerations Regarding the Use of Genotype and Pedigree Data to Model Relatedness in the Context of Genome-Wide Association Studies. Cheng R, Parker CC, Abney M, Palmer AA. G3 (Bethesda). 2013 Oct 3;3(10):1861-1867. PMID: 23979941

**See Also**

[extract.raw.data](#), [filter.samples](#), [batch.normalize](#), [calc.genoprob](#)

**Examples**

```
## Not run:  
K = kinship.probs(probs)  
  
## End(Not run)
```

---

muga.snps.to.keep

*SNPs to use for genotyping and mapping on the MUGA*

---

**Description**

SNPs to use for genotyping and mapping on the MUGA. These are a combination of one culled by UNC due to poor performance or uncertain location and ones culled by JAX based on low intensity values.

**Usage**

```
muga.snps.to.keep
```

**Format**

A character vector with MUGA SNP IDs to use when genotyping or mapping.

**Examples**

```
muga.snps.to.keep
```

---

```
plot.doqtl
```

```
Plot a QTL
```

---

**Description**

Given a genome scan produced by scanone, create a plot of the LOD score across all chromosomes.

**Usage**

```
## S3 method for class 'doqtl'
plot(x, stat.name = c("lod", "neg.log10.p"), sig.thr = NULL, sig.col = "red", ...)
```

**Arguments**

x	DOQTL object containing the LOD score and model coefficients.
stat.name	Character string containing the name of the mapping statistic.
sig.thr	Numeric vector containing significance thresholds.
sig.col	Colors to use when plotting the significance thresholds above.
...	Additional arguments to pass to plot.

**Value**

Creates a QTL plot.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run:
  qtl = scanone(pheno = pheno, pheno.col = 1, probs = probs, snps = snps)
  plot(qtl)
## End(Not run)
```

---

plot.genoprobs                      *Plot the genome of a DO sample.*

---

### Description

This function plots the genotype of a DO sample by taking the genotype with the maximum probability at each marker.

### Usage

```
## S3 method for class 'genoprobs'
plot(x, snps, colors = "mm10", chrLen = "mm10", ...)
chr.skeletons(chr, genome = "mm10", ...)
genomic.points(chr = NULL, loc = NULL)
write.genoprob.plots(path = ".", snps)
```

### Arguments

x	Numeric matrix containing the 36 genotype state probabilities generated by <a href="#">calc.genoprob</a> .
snps	Data.frame containing the SNP locations in prsmth. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively.
colors	Character string containing "DO" for the DO. FALSE or all other crosses, a data.frame containing single letter codes, strain names and colors for the founders in columns 1 through 3, respectively.
chrLen	Character string containing the genome for the chromosome lengths. Default = mm10.
genome	Character string containing the genome for the chromosome lengths. Default = mm10.
chr	Vector of chromosome IDs.
path	Character string containing the path to the posterior genotype probability files in R binary format (i.e. *.Rdata).
loc	Numeric vector of genome locations in Mb of same length as chr.
...	Additional arguments to be passed to plot.

### Value

plot.genoprobs plots the reconstructed DO genome in terms of founder haplotypes. chr.skeletons plots the chromosome skeletons. This is useful for further plotting. genomic.points plots locations on a genome such as might be drawn by chr.skeletons. write.genoprob.plots loops through all of the files supplied in the argument and writes out genotype plots.

### Author(s)

Daniel Gatti

### See Also

[calc.genoprob](#)

**Examples**

```
## Not run: plot.genoprobs(prsmth, snps, main = "plot title")
```

pxg.plot

*Phenotype by genotype plot at a single marker.***Description**

This function plots the phenotype versus the most probable genotype at a single marker.

**Usage**

```
pxg.plot(pheno, pheno.col, probs, snp.id, snps, legend = TRUE, sex = NA, covar, ...)
```

**Arguments**

pheno	Data.frame containing the phenotype data with samples in rows and phenotypes in columns. Sample IDs in rownames and phenotype names in colnames.
pheno.col	Numeric or character vector: Either a vector of number that indicate columns to use or a set of column names in pheno.
probs	3D numeric array, containing the founder haplotype contributions or genotype probabilities. The sample IDs, founder letter codes and markers IDs must be in dimnames.
snp.id	Character string containing the marker to plot at. Must be in SNPs and dimnames(probs)[[3]].
snps	Data.frame containing 4 columns with marker location information. SNP ID, chr, Mb, cM in columns 1 through 4, respectively.
legend	Boolean that is true if a legend explaining the founder letter codes should be plotted.
sex	Character that is either FALSE or M, indicating sex. This is used only when the SNP ID is on the X chromosome and all of the samples are male. In this case, there are only 8 genotype states.
covar	Vector of categories for each sample (i.e. diet, treatment, etc.) which will be plotted as different symbols. Optional.
...	Additional arguments passed along to plot.

**Details**

The most probable genotype is inferred from the eight founder allelic contributions at the marker. If a founder has a value > 0.75, it is assumed to be homozygous.

**Value**

Creates a plot with the phenotype on the y-axis and the 36 DO genotypes on the x-axis.

**Author(s)**

Daniel Gatti



**See Also**

[plot.doqtl](#), [coefplot](#)

**Examples**

```
## Not run: pxg.plot(pheno, pheno.col, founder.probs, snp.id, snps)
```

---

qtl.heatmap	<i>Plot a Heatmap of all QTL</i>
-------------	----------------------------------

---

**Description**

This function accepts LOD scores and SNP locations and plots a heatmap of the QTL. Each QTL is scaled to between 0 and 1 and the QTL are hierarchically clustered. A tile plot is then created with QTL colored from white (strong) to black (weak). The motivation for this plot is to visualize QTL that may be coincident across different phenotypes.

**Usage**

```
qtl.heatmap(lod, ...)
```

**Arguments**

lod	Data frame, containing the SNP names, chromosome and dbp locations in columns 1 through 3 and LOD scores for all phenotyped in the remaining columns.
...	Additional arguments to be passed to plot.

**Value**

No value is returned. A tile plot is plotted from the data.

**Note**

The QTL that appear in white are *not* necessarily significant. They are simply the strongest QTL for each phenotype.

**Author(s)**

Daniel Gatti

**See Also**

[plot.doqtl](#), [scanone](#), [scanone.perm](#)

**Examples**

```
## Not run:  
qtl.heatmap(lod)  
  
## End(Not run)
```

---

qtl.LRS *QTL mapping with no kinship.*

---

### Description

These are two functions that perform QTL mapping without a kinship matrix. They use the QR decomposition to speed up the computation. Other than for a quick screen or for assessing significance thresholds, we do not recommend mapping without a kinship matrix. They are included for historical reasons

### Usage

```
qtl.LRS(pheno, probs, snps, addcovar = NULL)
permutations.qtl.LRS(pheno, probs, snps, addcovar, nperm = 1000,
return.val = c("lrs", "p"))
```

### Arguments

pheno	Data.frame containing the phenotype data with samples in rows and phenotypes in columns. Sample IDs in rownames and phenotype names in colnames.
probs	Numeric three dimensional array, containing the founder haplotype contributions or genotype probabilities. The sample IDs, founder letter codes and markers IDs must be in dimnames.
snps	data.frame containing 4 columns with marker location information. SNP ID, chr, Mb, cM in columns 1 through 4, respectively.
addcovar	data.frame or numeric matrix, containing any additive covariates. Sample IDs must be in rownames.
nperm	Numeric value containing the number of permutations to run.
return.val	Character string containing either "LRS" or "p", indicating the type of return statistic.

### Details

The function performs Haley-Knott regression at the markers using the founder haplotype contributions in probs.

### Value

List containing two elements. LRS: a data.frame containing the LOD scores. coef: numeric matrix containing model coefficients.

### Author(s)

Daniel Gatti

### See Also

[scanone](#)

### Examples

```
## Not run: qtl.LRS(pheno, probs, snps, addcovar = NULL)
```

---

`qtl.qtlrel`*Use QTLRel to map a set of traits*

---

**Description**

This function accepts phenotypes, genotype probabilities and a sample kinship matrix and maps the requested traits using an eight state linear model. FALSEixed covariates may be passed in as well. The output is written to two files: \*.LOD.txt (containing the LOD scores for each SNP) and \*.coef.txt. (containing the model coefficient at each SNP).

**Usage**

```
qtl.qtlrel(pheno, probs, K, addcovar, intcovar, snps)
```

**Arguments**

pheno	Data frame, containing the sample IDs, phenotype data and covariates.
probs	3D numeric array, containing the genotype probabilities for all samples at each SNP. Dimensions must be samples by states by SNPs and all dimensions must be named.
K	Numeric matrix, containing the kinship between individuals as computed by QTLRel.
addcovar	Numeric matrix containing additive covariates.
intcovar	Numeric matrix containing covariates that interact with the QTL.
snps	Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively.

**Value**

A list containing two elements:

lod	Data.frame containing the SNP locations and LOD and p-values.
coef	Data.frame containing the model coefficients.

**Author(s)**

Daniel Gatti

**See Also**

[plot.doqtl](#), [scanone](#), [scanone.perm](#)

**Examples**

```
## Not run:
  qtl.qtlrel(pheno, prob, K, covar = NULL, pheno.name = "")
## End(Not run)
```

---

qtl.simulate

*Simulate a QTL in the DO*


---

### Description

Using a set of real DO genomes, simulate a QTL with a given minor allele frequency (MAF), sample size and effect size.

### Usage

```
qtl.simulate(probs, snps, K, sample.size = dim(probs)[[1]], effect.size = 1,
  maf = 4, num.poly = 18, num.sims = 1000)
```

### Arguments

probs	Numeric three dimensional array, containing the founder haplotype contributions or genotype probabilities. The sample IDs, founder letter codes and markers IDs must be in dimnames.
snps	Data.frame containing 4 columns with marker location information. SNP ID, chr, Mb, cM in columns 1 through 4, respectively.
K	Numeric matrix, containing the additive kinship matrix. The samples IDs must be in rownames and colnames.
sample.size	Numeric vector sample sizes. Must be less than or equal to the number of samples in probs.
effect.size	Numeric vector containing the effect sizes as the number of standard deviations from the phenotype mean.
maf	Numeric value containing the minor allele frequency as the number of founders. FALSE or DO, the value must be between 1 and 4.
num.poly	Numeric value containing the number of autosomes on which to simulate a polygenic background.
num.sims	Numeric value containing the number of simulations to run.

### Details

This function will simulate a phenotype with a QTL. It will output a phenotype vector with sample IDs and a data.frame called qtl describing the simulated loci.

### Value

Writes pheno and qtl lists to an R binary file.

### Author(s)

Daniel Gatti

### See Also

[scanone](#)

**Examples**

```
## Not run: qtl.simulate(probs, snps, K, sample.size = dim(probs)[[1]],
  effect.size = 1, maf = 4, num.poly = 18, num.sims = 1000)
## End(Not run)
```

---

rankZ	<i>Rank Z transformation</i>
-------	------------------------------

---

**Description**

This is also called the inverse normal transformation. We rank the data in  $x$ , divide by  $n-1$ , and take quantiles from the normal distribution using `qnorm`.

**Usage**

```
rankZ(x)
```

**Arguments**

$x$  Numeric vector of values to be transformed.

**Details**

We often use this when there are hundreds or thousands of phenotypes. This allows us to calculate permutation derived thresholds once and use them for all phenotypes.

**Value**

Numeric vector with a normal distribution.

**Author(s)**

Daniel Gatti

**Examples**

```
rankZ(rlnorm(20))
```

---

read.vcf                      *Read and parse VCFALSE data*

---

### Description

\*\*\* CURRENTLY UNDER CONSTRUCTION \*\*\*. This function is designed to read in the Sanger VCFALSE files for SNPs, Indels and structural variants.

### Usage

```
read.vcf(vcf.file, chr = 1, start = 4, end = 4.5, strains,
        return.val = c("allele", "number"), return.qual = TRUE, csq = FALSE)
```

### Arguments

vcf.file	Character string containing full path to a Sanger SNP, Indel or structural variant VCFALSE file.
chr	Character or numeric vector containing mouse chromosome IDs to query.
start	Numeric vector containing the start position on each chromosome to query. Must be the same length as chr. If the value is less than 200, it is assumed to be in Mb. Values greater than 200 are assumed to be in bp.
end	Numeric vector containing the end position on each chromosome to query. Must be the same length as chr. If the value is less than 200, it is assumed to be in Mb. Values greater than 200 are assumed to be in bp.
strains	Character vector containing strain names, as listed in the vcf.file, to retrieve. If missing, then all strains are returned. Use <a href="#">get.vcf.strains</a> to get the strain names from the VCFALSE file.
return.val	Character string that is either "allele", which returns the nucleotide allele calls, or "number", which returns numeric values.
return.qual	Boolean that is TRUE if the quality columns should be returned.
csq	Boolean that is TRUE if the consequence column should be returned.

### Details

There is a very nice package called vcf2geno, but it is designed to work with the 1000 Genomes VCFALSE format. The Sanger format is slightly different, hence the need for this function. FALSEu-ture plans include the addition of variant consequence selection. The Sanger variant files have 'snp', 'indel' or 'SV' in them and we use this to determine the type of file being processed.

### Value

If one chromosome location is requested, a data.frame containing the requested variants and quality scores. The variant locations and reference alleles are in the first 5 columns and the variants and quality scores for each strain are in the remaining columns. If more than one location is requested, a list of data.frames containing the variants.

### Note

!!! This function is still under development !!!

**Author(s)**

Daniel Gatti

**References**

The [Sanger Mouse Genomes Project](#) generated these variants. Raw data is at the [Sanger FALSETP site](#).

Next-generation sequencing of experimental mouse strains. Yalcin B, Adams DJ, FALSElint J and Keane TM Mammalian genome : official journal of the International Mammalian Genome Society 2012;23;9-10;490-8 PUBMED: 22772437

Sequencing and characterization of the FALSEVB/NJ mouse genome. Wong K, Bumpstead S, Van Der Weyden L, Reinholdt LG, Wilming LG, Adams DJ and Keane TM Genome biology 2012;13;8;R72 PUBMED: 22916792

The fine-scale architecture of structural variants in 17 mouse genomes. Yalcin B, Wong K, Bhomra A, Goodson M, Keane TM, Adams DJ and FALSElint J Genome biology 2012;13;3;R18 PUBMED: 22439878

The genomic landscape shaped by selection on transposable elements across 18 mouse strains. Nellaker C, Keane TM, Yalcin B, Wong K, Agam A, Belgard TG, FALSElint J, Adams DJ, FALSEr-ankel WN and Ponting CP Genome biology 2012;13;6;R45 PUBMED: 22703977

High levels of RNA-editing site conservation amongst 15 laboratory mouse strains. Danecek P, Nellaker C, McIntyre RE, Buendia-Buendia JE, Bumpstead S, Ponting CP, FALSElint J, Durbin R, Keane TM and Adams DJ Genome biology 2012;13;4;26 PUBMED: 22524474

Mouse genomic variation and its effect on phenotypes and gene regulation. Keane TM, Goodstadt L, Danecek P, White MA, Wong K, Yalcin B, Heger A, Agam A, Slater G, Goodson M, FALSEur-lotte NA, Eskin E, Nellaker C, Whitley H, Cleak J, Janowitz D, Hernandez-Pliego P, Edwards A, Belgard TG, Oliver PL, McIntyre RE, Bhomra A, Nicod J, Gan X, Yuan W, van der Weyden L, Steward CA, Bala S, Stalker J, Mott R, Durbin R, Jackson IJ, Czechanski A, Guerra-Assuncao JA, Donahue LR, Reinholdt LG, Payseur BA, Ponting CP, Birney E, FALSElint J and Adams DJ Nature 2011;477;7364;289-94 PUBMED: 21921910

Sequence-based characterization of structural variation in the mouse genome. Yalcin B, Wong K, Agam A, Goodson M, Keane TM, Gan X, Nellaker C, Goodstadt L, Nicod J, Bhomra A, Hernandez-Pliego P, Whitley H, Cleak J, Dutton R, Janowitz D, Mott R, Adams DJ and FALSElint J Nature 2011;477;7364;326-9 PUBMED: 21921916

**Examples**

```
## Not run: read.vcf(vcf.file =
  "ftp://ftp-mouse.sanger.ac.uk/current_snps/mgp.v3.snps.rsIDdbSNPv137.vcf.gz",
  chr = 1, start = 5, end = 5.5)
## End(Not run)
```

scanone

*Perform a genome scan.***Description**

This function is the main QTL mapping function for point mapping at each marker. The user must supply phenotypes, genotype probabilities and marker locations. Optional kinship, additive and interactive covariates may be passed in. Scanone regresses the phenotype on the genotype probabilities and produces a LOD score and founder allele effects at each marker.

**Usage**

```
scanone(pheno, pheno.col = 1, probs, K, addcovar, intcovar, snps, model =
c("additive", "full"))
```

**Arguments**

pheno	data.frame containing phenotype data. Required. rownames must contain sample IDs and there must be a column labelled 'sex' to perform correct mapping on the X chromosome.
pheno.col	numeric or character vector: Either a vector of number that indicate columns to use or a set of column names in pheno.
probs	Numeric three dimensional array, containing the founder haplotype contributions or genotype probabilities. The sample IDs, founder letter codes and markers IDs must be in dimnames.
K	numeric matrix, containing the additive kinship matrix. The samples IDs must be in rownames and colnames.
addcovar	data.frame or numeric matrix, containing any additive covariates. Sample IDs must be in rownames.
intcovar	data.frame or numeric matrix, containing any covariates that interact with the QTL. Sample IDs must be in rownames.
snps	data.frame containing 4 columns with marker location information. SNP ID, chr, Mb, cM in columns 1 through 4, respectively.
model	character string, containing one of "additive" or "full", indicating the type of model to fit.

**Details**

We require a sex argument in addcovar for mapping on the X chromosome. This is true even if all of your samples are of the same sex. It also means that mapping on the autosomes will occur with sex as an additive covariate. See A Guide to QTL Mapping with R/qtl by Karl Broman, pages 108-118.

**Value**

A list containing two elements:

lod	Data.frame with nine columns containing the marker locations, LOD scores and p-values for each phenotype. Column names are SNP_ID, Chr, Mb_NCBI38, cM, perc.var, lrs, lod, p and neg.log10.p.
coef	Numeric matrix containing the founder allele effects at each locus. Colnames are the additive covariates plus the founder terms.

**Author(s)**

Daniel Gatti

**See Also**

[scanone.perm](#)

**Examples**

```
## Not run: scanone(pheno, pheno.col = 1, probs)
```



---

scanone.eqtl                      *Mapping using the Matrix EQTL algorithm.*

---

### Description

MatrixEQTL uses a series of matrix operations to greatly accelerate QTL mapping. It can accommodate additive covariates and a common kinship matrix for all phenotypes.

### Usage

```
scanone.eqtl(expr, probs, K, addcovar, snps, sex)
```

### Arguments

expr	Numeric matrix of phenotype values with samples in rows and phenotypes in columns. Rownames must contain sample IDs and colnames must contain phenotype names.
probs	Numeric three dimensional array containing the founder haplotype contributions. Num.samples by num.founders by num.markers.
K	Numeric matrix of kinship values for all samples. Num.samples by num.samples.
addcovar	Numeric matrix of additive covariates.
snps	Data.frame containing 4 columns with marker location information. SNP ID, chr, Mb, cM in columns 1 through 4, respectively.
sex	Character vector containing either "M" or F, indicating the sex of each sample. Used for mapping on the X chromosome.

### Details

Matrix EQTL centers and rotates the phenotype and genotype matrices using matrix operations. It only calculates the LOD score at each marker and does not provide coefficients.

### Value

Numeric matrix of LOD scores for all phenotypes and markers.

### Author(s)

Daniel Gatti

### References

Matrix eQTL: ultra fast eQTL analysis via large matrix operations. Shabalin AA. Bioinformatics. 2012 May 15;28(10):1353-8.

### See Also

[scanone](#)

### Examples

```
## Not run: scanone.eqtl(expr, probs, K, addcovar, snps)
```

---

scanone.perm	<i>Perform a genome scan.</i>
--------------	-------------------------------

---

### Description

This function is the main QTL mapping function for point mapping at each marker. The user must supply phenotypes, genotype probabilities and marker locations. Optional kinship, additive and interactive covariates may be passed in. scanone.perm regresses the phenotype on the genotype probabilities and produces a LOD score and founder allele effects at each marker.

### Usage

```
scanone.perm(pheno, pheno.col = 1, probs, addcovar, intcovar, snps, model =
c("additive", "full"), path = ".", nperm = 1000)
```

### Arguments

pheno	data.frame containing phenotype data. Required. rownames must contain sample IDs and there must be a column labelled 'sex' to perform correct mapping on the X chromosome.
pheno.col	numeric or character vector: Either a vector of number that indicate columns to use or a set of column names in pheno.
probs	3D numeric array, containing the founder haplotype contributions or genotype probabilities. The sample IDs, founder letter codes and markers IDs must be in dimnames.
addcovar	data.frame or numeric matrix, containing any additive covariates. Sample IDs must be in rownames.
intcovar	data.frame or numeric matrix, containing any covariates that interact with the QTL. Sample IDs must be in rownames.
snps	data.frame containing 4 columns with marker location information. SNP ID, chr, Mb, cM in columns 1 through 4, respectively.
model	Character string, containing one of "additive" or "full", indicating the type of model to fit.
path	Character string containing the location where permutation results should be written.
nperm	Numeric, containing the number of permutations to run.

### Value

A list containing two elements:

lod	Data.frame with seven columns containing the marker locations and LOD scores for each phenotype.
coef	Numeric matrix containing the founder allele effects at each locus.

### Author(s)

Daniel Gatti

**See Also**[scanone.perm](#)**Examples**

```
## Not run: scanone.perm(pheno, pheno.col = 1, probs)
```

---

`sdp.plot`*Plot association mapping results.*

---

**Description**

Plot the founder strain distribution patterns (SDP) of the SNPs in results.

**Usage**

```
sdp.plot(results, ...)
```

**Arguments**

<code>results</code>	Data.frame containing output from <a href="#">assoc.map</a> .
<code>...</code>	Additional arguments passed to plot.

**Details**

Given the output from [assoc.map](#), plot the SDPs for the SNPs in results. This will show which strains have the minor allele at each SNP in results. You may also add this plot to the top of [assoc.plot](#).

**Value**

Produces a plot. There is no return value.

**Author(s)**

Daniel Gatti

**See Also**[assoc.map](#), [assoc.plot](#)**Examples**

```
## Not run:
  results = assoc.map(pheno = pheno, pheno.col = 1, probs = probs, K = K, addcovar = addcovar,
snp = snps, chr = 1, start = 40, end = 45)
  sdp.plot(results[results[,12] > 3,])

## End(Not run)
```

---

sex.predict	<i>Determine the sex of each sample</i>
-------------	---

---

### Description

This function uses the mean X and Y chromosome intensities to predict the sex of each sample. Linear discriminant analysis from the MASS package is used.

### Usage

```
sex.predict(x, y, snps, plot = FALSE)
```

### Arguments

x	Numeric matrix, num.samples x num.snps, containing X intensities for all samples.
y	Numeric matrix, num.samples x num.snps, containing Y intensities for all samples.
snps	Data.frame containing SNP IDs, chromosomes, Mb and cM locations in columns 1 through 4, respectively.
plot	Boolean that will create a plot of mean X and Y Chr intensities if TRUE. Default = FALSE.

### Value

Character vector with sex assignments.

### Author(s)

Daniel Gatti

### Examples

```
## Not run:
  data(snps)
  sex.predict(x, y, sex = rep(F, ncol(x)))

## End(Not run)
```

---

snp.plot	<i>snp.plot</i>
----------	-----------------

---

### Description

Lower level function that makes a tile plot the given SNPs with the major allele colored dark and the minor allele light. Optionally, add SNPs that match a certain pattern and genes and a QTL score.

**Usage**

```
snp.plot(variants, col = c("black", "grey50", "white"), cluster = TRUE, ref, highlight,
pattern.snps, mgi, qtl)
```

**Arguments**

variants	Data.frame with variants as returned by <a href="#">get.variants</a> .
col	Color vector with SNP colors for no call, alternate allele and reference allele.
cluster	Boolean that indicates if the strains should be clustered.
ref	Character that is the reference strain to use. Must be present in the strains in variants.
highlight	Character vector with strain names to highlight in the plot. Strain names must be present in the strains in variants.
pattern.snps	Data.frame with SNPs that match some pattern. These will be plotted below the SNPs.
mgi	Data.frame with genes in the interval to be plotted.
qtl	data.frame with QTL values containing the chromosome, bp position and QTL score in column 1 to 3. Default = NULL.

**Details**

Different strains can be used as the reference strain by using the `ref` argument. Otherwise, the major allele is plotted dark and the minor allele lighter. The QTL values will be scaled within the plotting interval and drawn with black (low) and red (high). If a strain pattern is provided, the SNPs matching the pattern are plotted in orange and any genes that they intersect with are also colored orange. Otherwise, genes are colored blue.

**Value**

Produces a tile plot with the locations along the horizontal axis and the strains in the SNP matrix along the vertical axis. Also, if a set of strains is given in the `pattern` argument, the SNPs that match that pattern are returned, categorized according to which gene they lie within.

**Author(s)**

Daniel Gatti

**See Also**

[variant.plot](#), [get.mgi.features](#), [categorize.variants](#)

**Examples**

```
## Not run:
data(qtl)
strains = get.strains()
variants = get.variants(chr = 7, start = 103, end = 105,
strains = strains[c(2,4,8,10,15:18)])
pattern.snps = get.pattern.variants(variants, strain.subset = c("C57BL/6J", "NOD/ShiLtJ",
"NZO/H1LtJ"))
mgi = get.mgi.features(chr = 7, start = 103, end = 105, source = "MGI", type = "gene")
variants = convert.variants.to.numeric(variants)
```

```
snp.plot(variants = variants, pattern.snps = pattern.snps, mgi = mgi, qtl = qtl)

## End(Not run)
```

---

```
summarize.genotype.transitions
```

*Summarize the genotype data output by the genotyping HMM.*

---

## Description

These functions read in all of the individual genotype data files and summarizes the founder allele frequency by sample and SNP.

## Usage

```
summarize.genotype.transitions(path = ".", snps)
summarize.by.snps(path = ".", snps)
summarize.by.samples(path = ".", snps)
num.recomb.plot(results, gen)
```

## Arguments

path	Character, full path to the genotype directory where the *.genotype.probs.Rdata files are stored.
snps	Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively.
results	Data.frame containing results as summarized by summarize.genotype.transitions.
gen	Numeric vector containing DO outbreeding generation for all samples in results. Must have sample IDs in names.

## Value

Data.frame with six columns: sample, proximal genotype, distal genotype, chr, proximal Mb, distal Mb.

## Author(s)

Daniel Gatti

## Examples

```
## Not run: geno.sum = summarize.genotype.transitions()
```

---

variant.plot	<i>variant.plot</i>
--------------	---------------------

---

## Description

Convenience function that makes a tile plot the given SNPs with the major allele colored dark and the minor allele light. Optionally, add SNPs that match a certain pattern and genes and a QTL score. This calls [sdp.plot](#) after gathering all of the data.

## Usage

```
variant.plot(var.file = "ftp://ftp.jax.org/SNPtools/variants/cc.snps.NCBI38.txt.gz",
            mgi.file = "ftp://ftp.jax.org/SNPtools/genes/MGI.20130703.sorted.txt.gz", chr,
            start, end, type = c("snp", "indel", "sv"), strains = c("A/J", "C57BL/6J",
            "129S1/SvImJ", "CAST/EiJ", "NOD/ShiLtJ", "NZO/H1LtJ", "PWK/PhJ", "WSB/EiJ"),
            ref = "C57BL/6J", pattern, qtl)
```

## Arguments

<code>var.file</code>	Character string with the full path to the tabix indexed Sanger/UNC SNP file. Default is a file at the Center for Genome Dynamics at the Jackson Laboratory.
<code>mgi.file</code>	Character string with the full path to the tabix indexed MGI feature file. Default is a file at the Center for Genome Dynamics at the Jackson Laboratory.
<code>chr</code>	Character with the chromosome ID to plot on. Required.
<code>start</code>	Numeric value with the start position to plot at. May be in bp or Mb. If the value is 200 or less, it is assumed to be in Mb. Otherwise it is assumed to be in bp. Required.
<code>end</code>	Numeric value with the end position to plot at. May be in bp or Mb. If the value is 200 or less, it is assumed to be in Mb. Otherwise it is assumed to be in bp. Required.
<code>type</code>	Character indicating the type of variant to retrieve. One of "snp", "indel", or "sv". Default = "snp".
<code>strains</code>	Character vector with the strains to use. Must be from the set of strains returned by <code>get.strains{get.strains}</code> . Default = Collaborative Cross/Diversity Outbred founders.
<code>ref</code>	Character string with the reference strain. Must be one of the names in the strains vector argument. Default = C57BL/6J.
<code>pattern</code>	Character vector with strain names in the strains argument. We search for SNPs for which these strains have one allele and all other strains have the opposite allele. A more sophisticated pattern matching method may be included in future releases. Default = NULL.
<code>qtl</code>	data.frame with QTL values containing the chromosome, bp position and QTL score in column 1 to 3. Default = NULL.

**Details**

Different strains can be used as the reference strain by using the `ref` argument. Otherwise, the major allele is plotted dark and the minor allele lighter. The QTL values will be scaled within the plotting interval and drawn with black (low) and red (high). If a strain pattern is provided, the SNPs matching the pattern are plotted in orange and any genes that they intersect with are also colored orange. Otherwise, genes are colored blue.

**Value**

Produces a tile plot with the locations along the horizontal axis and the strains in the SNP matrix along the vertical axis. Also, if a set of strains is given in the `pattern` argument, the SNPs that match that pattern are returned, categorized according to which gene they lie within.

**Author(s)**

Daniel Gatti

**See Also**

[convert.variants.to.numeric](#), [get.mgi.features](#), [categorize.variants](#), [snp.plot](#)

**Examples**

```
## Not run:
data(qtl)
strains = get.strains()[c(2,4,8,10,15:18)]
variant.plot(chr = 7, start = 103, end = 105, strains = strains,
pattern = c("C57BL/6J", "NOD/ShiLtJ", "NZO/HiLtJ"), qtl = qtl)

## End(Not run)
```

---

`write.founder.genomes` *Write out the genotypes of DO samples*

---

**Description**

Given a directory containing files generated by the DOQTL HMM (i.e. that end with "genotype.probs.Rdata"), write out two files for each sample containing the founder haplotype blocks.

**Usage**

```
write.founder.genomes(filenamees = dir(path = ".",
pattern = "genotype.probs.Rdata"), snps)
```

**Arguments**

<code>filenamees</code>	Character vector of posterior genotype probability files in R binary format.
<code>snps</code>	Data.frame containing the marker locations. SNP ID, chromosome, Mb and cM locations in columns 1 through 4, respectively.



**Details**

FALSEor each sample, we take the genotype state with the highest probability and write it out.

**Value**

No value is returned. FALSEiles are written to the current working directory.

**Author(s)**

Daniel Gatti

**Examples**

```
## Not run: write.founder.genomes(filenamees = dir(path = ".",  
pattern = "genotype.probs.Rdata"))  
## End(Not run)
```

# Index

- \*Topic **Diversity Outbred**
  - summarize.genotype.transitions, 70
- \*Topic **HMM**
  - calc.genoprob, 11
  - calc.genoprob.alleles, 13
  - calc.genoprob.intensity, 14
  - coefplot, 18
  - condense.model.probs, 20
  - create.genotype.states, 24
  - do.trans.probs, 27
  - get.trans.probs, 48
  - summarize.genotype.transitions, 70
- \*Topic **HTML**
  - html.report, 50
- \*Topic **HTNL**
  - create.html.page, 25
- \*Topic **MGI**
  - convert.genes.to.GRanges, 22
  - get.gene.name, 43
- \*Topic **MUGA**
  - batch.normalize, 9
  - calc.genoprob, 11
  - calc.genoprob.alleles, 13
  - calc.genoprob.intensity, 14
  - coefplot, 18
  - extract.raw.data, 34
  - filter.samples, 38
  - kinship.probs, 52
- \*Topic **QTLRel**
  - qtl.heatmap, 57
  - qtl.qtlrel, 59
- \*Topic **QTL**
  - bayesint, 10
  - coef.doqtl, 18
  - fast.qtlrel, 35
  - filter.geno.probs, 37
  - html.report, 50
  - plot.genoprob, 55
  - pxg.plot, 56
  - qtl.heatmap, 57
  - qtl.LRS, 58
  - qtl.qtlrel, 59
  - qtl.simulate, 60
  - scanone, 63
  - scanone.eqtl, 65
  - scanone.perm, 66
- \*Topic **Sanger**
  - do2sanger, 28
  - read.vcf, 62
- \*Topic **VCFALSE**
  - read.vcf, 62
- \*Topic **\textasciitildekw1**
  - create.Rdata.files, 26
- \*Topic **\textasciitildekw2**
  - create.Rdata.files, 26
- \*Topic **alleles**
  - convert.allele.calls, 21
  - convert.genotypes, 22
- \*Topic **association**
  - assoc.map, 6
  - assoc.plot, 8
  - do2sanger, 28
  - sdp.plot, 67
- \*Topic **autosomes**
  - get.num.auto, 46
- \*Topic **call rate**
  - filter.samples, 38
- \*Topic **chromosome**
  - get.chr.lengths, 42
- \*Topic **cluster**
  - estimate.cluster.params, 30
- \*Topic **datasets**
  - do.colors, 26
  - do.states, 27
  - example.genes, 31
  - example.pheno, 32
  - example.qtl, 33
  - example.snps, 34
  - muga.snps.to.keep, 53
- \*Topic **genome**
  - genome.summary.plots, 41
- \*Topic **genotypes**
  - filter.geno.probs, 37
  - get.do.states, 43
  - write.founder.genomes, 72
- \*Topic **genotype**

- create.genotype.states, 24
  - get.max.geno, 44
- \*Topic **genotyping**
  - calc.genoprob, 11
  - calc.genoprob.alleles, 13
  - calc.genoprob.intensity, 14
  - coefplot, 18
  - summarize.genotype.transitions, 70
- \*Topic **haplotypes**
  - write.founder.genomes, 72
- \*Topic **haplotype**
  - plot.genoprob, 55
- \*Topic **imputation**
  - do2sanger, 28
- \*Topic **intensities**
  - intensity.plots, 51
- \*Topic **intensity**
  - batch.normalize, 9
  - extract.raw.data, 34
- \*Topic **kinship**
  - kinship.probs, 52
- \*Topic **log**
  - colSumsLog, 19
- \*Topic **mapping**
  - qtl.heatmap, 57
  - qtl.qtlrel, 59
- \*Topic **normalization**
  - batch.normalize, 9
  - filter.samples, 38
- \*Topic **precision**
  - get.machine.precision, 44
- \*Topic **qtl**
  - assoc.map, 6
  - assoc.plot, 8
  - sdp.plot, 67
- \*Topic **rankZ**
  - rankZ, 61
- \*Topic **sanger**
  - assoc.map, 6
- \*Topic **sdp**
  - sdp.plot, 67
- \*Topic **sex**
  - sex.predict, 68
- \*Topic **snps**
  - fill.in.snps, 36
- \*Topic **summary**
  - genome.summary.plots, 41
- \*Topic **transition**
  - cc.trans.probs, 16
  - do.trans.probs, 27
  - generic.trans.probs, 40
  - get.trans.probs, 48
- \*Topic **variants**
  - read.vcf, 62
- add.missing.F1s, 3
- add.slash, 4
- addLog, 4, 5, 20
- addLogVector, 5, 5
- assoc.map, 6, 8, 9, 30, 67
- assoc.plot, 8, 8, 67
- batch.normalize, 9, 35, 38, 53
- bayesint, 10
- calc.genoprob, 11, 21, 53, 55
- calc.genoprob.alleles, 13
- calc.genoprob.intensity, 14
- categorize.variants, 15, 69, 72
- cc.trans.probs, 16, 41
- chr.skeletons (plot.genoprob), 55
- cluster.strains, 17
- coef.doqtl, 18
- coefplot, 18, 57
- colSumsLog, 19
- condense.model.probs, 6, 20
- convert.allele.calls, 21
- convert.genes.to.GRanges, 22
- convert.genotypes, 22
- convert.variants.to.GRanges, 23
- convert.variants.to.numeric, 17, 23, 72
- create.genotype.states, 24
- create.html.page, 25
- create.Rdata.files, 26
- do.colors, 26
- do.states, 27
- do.trans.probs, 27, 41
- do2sanger, 28
- estimate.cluster.params, 30
- example.genes, 31
- example.pheno, 32, 33
- example.qtl, 33
- example.snps, 34
- extract.raw.data, 10, 34, 38, 53
- fast.qtlrel, 35
- fill.in.snps, 36
- filter.geno.probs, 37
- filter.samples, 10, 35, 38, 53
- find.overlapping.genes, 38
- founder.F1.intensity.plot  
(intensity.plots), 51
- gene.plot, 39

- generic.trans.probs, 40
- genome.summary.plots, 41
- genomic.points (plot.genoprobs), 55
- genotype.by.sample.barplot
  - (genome.summary.plots), 41
- genotype.by.snp.barplot
  - (genome.summary.plots), 41
- get.additive (condense.model.probs), 20
- get.chr.lengths, 42
- get.do.states, 43
- get.dominance (condense.model.probs), 20
- get.full (condense.model.probs), 20
- get.gene.name, 43
- get.machine.precision, 44
- get.max geno, 44
- get.mgi.features, 22, 39, 40, 43, 45, 69, 72
- get.num.auto, 46
- get.pattern.variants, 47
- get.strains, 48, 50, 71
- get.trans.probs, 48
- get.variants, 15, 16, 47, 49, 69
- get.vcf.strains, 62
- get.vcf.strains (read.vcf), 62
  
- hmm.intensity, 31
- html.report, 25, 50
  
- intensity.mean.covar.plot
  - (intensity.plots), 51
- intensity.plots, 51
  
- keep.homozygotes
  - (estimate.cluster.params), 30
- kinship.alleles (kinship.probs), 52
- kinship.probs, 52
  
- matrizeqtl.snps (scanone.eqtl), 65
- muga.snps.to.keep, 53
  
- normalize.batches (batch.normalize), 9
- num.recomb.plot
  - (summarize.genotype.transitions), 70
  
- permutations.qtl.LRS (qtl.LRS), 58
- plot.doqtl, 19, 54, 57, 59
- plot.genoprobs, 55
- prsmth.plot (plot.genoprobs), 55
- pxg.plot, 56
  
- qtl.heatmap, 57
- qtl.LRS, 58
- qtl.qtlrel, 59
- qtl.simulate, 60
  
- quantilenorm (batch.normalize), 9
  
- rankZ, 61
- read.vcf, 62
- rowSumsLog (colSumsLog), 19
  
- scanone, 11, 18, 19, 36, 37, 51, 57–60, 63, 65
- scanone.eqtl, 65
- scanone.perm, 11, 37, 57, 59, 64, 66, 67
- sdp.plot, 67, 71
- sex.predict, 68
- snp.plot, 68, 72
- summarize.by.samples, 41, 42
- summarize.by.samples
  - (summarize.genotype.transitions), 70
- summarize.by.snps, 41, 42
- summarize.by.snps
  - (summarize.genotype.transitions), 70
- summarize.genotype.transitions, 70
  
- variant.plot, 17, 69, 71
  
- write.founder.genomes, 72
- write.genoprob.plots (plot.genoprobs), 55