# Package 'COHCAP'

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lype Package	
<b>Title</b> CpG Island Analysis Pipeline for Illumina Methylation Array and Targeted BS-Seq Data	
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Depends WriteXLS, COHCAPanno	
Description This package provides a pipeline to analyze single-nucleotide resolution methylation data (Illumina 450k methylation array, targeted BS-Seq, etc.). It provides QC metrics, differential methylation for CpG Sites, differential methylation for CpG Islands, integration with gene expression data, and visualization of methylation values.	
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COHCAP.annotate

Annotation for CpG Sites

#### **Description**

Provides annotations (chromosome, location, gene, and CpG island) for CpG sites from a specified annotation file.

Annotations for common platforms are provided with COHCAP (with respect to hg19). Custom annotation files can also be provided.

Output files will be created in the "Raw\_Data" subfolder.

#### Usage

```
COHCAP.annotate(beta.file, project.name, project.folder,
platform, annotation.file = NULL,
output.format = "xls")
```

#### **Arguments**

beta.file Table of beta / percentage methylation values. CpG sites are represented in rows.

Samples are represented in columns.

Name for COHCAP project. This determines the names for output files. project.name

project.folder Folder for COHCAP output files

Annotation file to be used. Enter "450k-UCSC" for UCSC CpG Islands for 450k platform

array probes, "450k-HMM" for HMM CpG Islands for 450k array probes, "27k"

for UCSC CpG Islands for 27k array probes.

If none of these pre-defined annotations are acceptable, please enter "custom"

for the platform and provide an annotation file.

annotation.file

Annotation file to be used for a custom platform. This variable is not used for

common, pre-defined annotation files.

The annotation file should be a tab-delimited text file with the header "SiteID Chr Loc Gene Island", with columns respectively specifying the CpG identifier (must match beta / percent methylation file), chromosome for CpG site, position for CpG site (preferably in hg19 coordinates), nearest gene mapping for CpG

site, nearest CpG island mapping for CpG site.

Format for output tables: 'xls' for Excel file, 'txt' for tab-delimited text file output.format

## Value

Data frame of beta values (must be between 0 and 1) or percentage methylation values (must be between 0 and 100).

Just like the input table, the first column specifies the SiteID, CpG sites are represented on rows, samples are represented in samples (starting with the 6th column). Additionally, the 2nd column now specifies the CpG site chromosome, the 3rd column now specifies the CpG site position (in hg19 coordinates, for pre-defined annotation files), the 4th column lists the nearest gene mapping, and the 5th column lists the nearest CpG island mapping.

This data frame is used for quality control and differential methylation analysis.

#### See Also

COHCAP Discussion Group: http://sourceforge.net/p/cohcap/discussion/general/

## **Examples**

```
library("COHCAP")

dir <- system.file("extdata", package="COHCAP")
beta.file <- file.path(dir,"GSE42308_truncated.txt")
project.folder <- getwd()
project.name <- "450k_test"

beta.table <- COHCAP.annotate(beta.file, project.name, project.folder, platform="450k-UCSC")</pre>
```

COHCAP.avg.by.island CpG Island Differential Methylation Analysis (Average by Island Workflow).

## **Description**

Provides statistics for CpG islands as well as a list of differentially methylated sites. CpG Island statistics are calculated by averaging beta values among samples per site and comparing the average beta values across groups (considering the pairing between sites).

List of differentially methylated islands will be created in the "CpG\_Island" folder. Table of statistics for all CpG islands will be created in the "Raw\_Data" folder.

## Usage

```
COHCAP.avg.by.island(sample.file, site.table, beta.table, project.name, project.folder, methyl.cutoff=0.7, unmethyl.cutoff = 0.3, delta.beta.cutoff = 0.2, pvalue.cutoff=0.05, fdr.cutoff=0.05, num.groups=2, num.sites=4, plot.box=TRUE, paired=FALSE, ref="none", output.format = "xls", gene.centric=TRUE)
```

## **Arguments**

sample.file	Tab-delimited text file providing group attributions for all samples considered for analysis.
beta.table	Data frame with CpG sites in columns (with DNA methylation represented as beta values or percentage methylation), samples in columns, and CpG site annotations are included (in columns 2-5).
	The COHCAP.annotate function automatically creates this file.
site.table	Data frame with CpG site statistics (one row per CpG site) and CpG site annotations (in columns 2-5).
	The COHCAP.site function automatically creates this file.
project.name	Name for COHCAP project. This determines the names for output files.
project.folder	Folder for COHCAP output files

methyl.cutoff Minimum beta or percentage methylation value to be used to define a methylated CpG site. Default is 0.7 (used for beta values), which would correspond to 70 Used for either 1-group or 2-group comparison.

unmethyl.cutoff

Minimum beta or percentage methylation value to be used to define an unmethylated CpG site. Default is 0.3 (used for beta values), which would correspond to 30 Used for either 1-group or 2-group comparison.

delta.beta.cutoff

fdr.cutoff

The minimum absolute value for delta-beta values (mean treatment beta - mean reference beta) to define a differentially methylated CpG site. Only used for 2-group comparison.

pvalue.cutoff Maximum p-value allowed to define a site as differentially methylated. Used only for comparisons with at least 2 groups (with 3 replicates per group)

Maximum False Discovery Rate (FDR) allowed to define a site as differentially

methylated. Used only for comparisons with at least 2 groups (with 3 replicates

per group)

num.groups Number of groups described in sample description file. COHCAP algorithm

differs when analysing 1-group, 2-group, or >2-group comparisons. COHCAP

cannot currently analyze continuous phenotypes.

num.sites Minimum number of differentially methylated sites to define a differentially

methylated CpG island.

ref Reference group used to define baseline methylation levels. Only used for 2-

group comparison

plot.box A logical value: Should box-plots be created to visualize CpG island differential

methylation?

paired A logical value: Is there any special pairing between samples in different groups?

If so, the pairing variable must be specified in the 3rd column of the sample description file. Used for p-value calculation, so this only applies to comparisons

with at least 2 groups.

output.format Format for output tables: 'xls' for Excel file, 'txt' for tab-delimited text file

gene.centric Should CpG islands not mapped to genes be ignored? Default: TRUE (Recom-

mended setting for integration with gene expession data)

#### Value

Data frame of average beta (or percentage methylation) values across differentially methylated sites within a differentially methylated CpG island.. This data frame can be used for integration analysis.

## See Also

COHCAP Discussion Group: http://sourceforge.net/p/cohcap/discussion/general/

```
library("COHCAP")

dir <- system.file("extdata", package="COHCAP")
beta.file <- file.path(dir,"GSE42308_truncated.txt")
sample.file <- file.path(dir,"sample_GSE42308.txt")
project.folder <- getwd()</pre>
```

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```
project.name <- "450k_avg_by_island_test"

beta.table <- COHCAP.annotate(beta.file, project.name, project.folder, platform="450k-UCSC")
filtered.sites <- COHCAP.site(sample.file, beta.table, project.name, project.folder, ref="parental")
filtered.islands <- COHCAP.avg.by.island(sample.file, filtered.sites, beta.table, project.name, project.folder, ref="parental")</pre>
```

COHCAP.avg.by.site

CpG Island Differential Methylation Analysis (Average by Site Workflow).

## **Description**

Provides statistics for CpG islands as well as a list of differentially methylated sites. CpG Island statistics are calculated by averaging beta values among samples per site and comparing the average beta values across groups (considering the pairing between sites).

List of differentially methylated islands will be created in the "CpG\_Island" folder. Table of statistics for all CpG islands will be created in the "Raw\_Data" folder.

#### Usage

```
COHCAP.avg.by.site(site.table, project.name, project.folder, methyl.cutoff=0.7, unmethyl.cutoff = 0.3, delta.beta.cutoff = 0.2, pvalue.cutoff=0.05, fdr.cutoff=0.05, num.groups=2, num.sites=4, output.format = "xls")
```

#### **Arguments**

site.table Data frame with CpG site statistics (one row per CpG site) and CpG site anno-

tations (in columns 2-5).

The COHCAP.site function automatically creates this file.

project.folder Folder for COHCAP output files

methyl.cutoff Minimum beta or percentage methylation value to be used to define a methylated

 $\mbox{CpG}$  site. Default is 0.7 (used for beta values), which would correspond to 70

Used for either 1-group or 2-group comparison.

unmethyl.cutoff

Minimum beta or percentage methylation value to be used to define an unmethylated CpG site. Default is 0.3 (used for beta values), which would correspond to 30 Used for either 1-group or 2-group comparison.

delta.beta.cutoff

The minimum absolute value for delta-beta values (mean treatment beta - mean reference beta) to define a differentially methylated CpG site. Only used for

2-group comparison.

pvalue.cutoff Maximum p-value allowed to define a CpG island as differentially methylated.

fdr. cutoff Maximum False Discovery Rate (FDR) allowed to define CpG island as differ-

entially methylated.

num.groups	Number of groups described in sample description file. COHCAP algorithdiffers when analysing 1-group, 2-group, or >2-group comparisons. COHC cannot currently analyze continuous phenotypes.	
num.sites	Minimum number of differentially methylated sites to define a differentially methylated CpG island.	
output.format	Format for output tables: 'xls' for Excel file, 'txt' for tab-delimited text file	

#### Value

Data frame of average beta (or percentage methylation) statistics and/or p-value / false discovery rate statistics (per CpG island).

The content of the data frame depends upon the number of groups specified for analysis. All workflows provide p-values and FDR values. 1 and 2 group comparisons provide counts for methylated and unmethylated sites as well as an overall methylation status per island. >2 group comparisons only provide counts for the total number of differentially methylated sites.

This data frame can be used for integration analysis.

#### See Also

COHCAP Discussion Group: http://sourceforge.net/p/cohcap/discussion/general/

## **Examples**

```
library("COHCAP")

dir <- system.file("extdata", package="COHCAP")
beta.file <- file.path(dir,"GSE42308_truncated.txt")
sample.file <- file.path(dir,"sample_GSE42308.txt")
project.folder <- getwd()
project.name <- "450k_avg_by_site_test"

beta.table <- COHCAP.annotate(beta.file, project.name, project.folder, platform="450k-UCSC")
filtered.sites <- COHCAP.site(sample.file, beta.table, project.name, project.folder, ref="parental")
filtered.islands <- COHCAP.avg.by.site(filtered.sites, project.name, project.folder)</pre>
```

COHCAP.BSSeq.preprocess

Preprocessing for Targeted BS-Seq data

## **Description**

Creates custom annotation file as well as COHCAP input file (for COHCAP.annotate).

This function is not necessary for Illumina methylation array analysis.

Output files will be created in specified locations

#### Usage

```
COHCAP.BSSeq.preprocess(methyl.folder=getwd(),
cohcap.inputfile = file.path(getwd(),"BS_Seq_combined.txt"),
gene.table = file.path(getwd(),"GENCODE_Genes.bed"),
targeted.regions = file.path(getwd(),"UCSC_CpG_Islands.bed"),
annotation.file = file.path(getwd(),"COHCAP.targeted.BSSeq.anno.txt"),
shore.length=2000)
```

#### **Arguments**

methyl.folder Folder containing .bed files created using genome\_methylation\_bismark2bedGraph\_v3.pl (following Bismark alignment)

cohcap.inputfile

Output file containing a tab-delimited table of percentage methylation values.

This table will compatible with the custom annotation file created by this func-

tion (annotation.file)

gene.table .bed file containing gene names and coordinates

targeted.regions

.bed file containing regions selected for targeted BS-Seq

annotation.file

Custom annotation file providing gene and targeted region mappings for CpG

sites specifically covered in your Bismark alignment

shore.length Length of shores considered to be part of the CpG island (in bp upstream and

downstream of targeted region coordinates)

## Value

This function creates two tab-delimited text files.

One is to be used to as a custom annotation file (annotation.file).

The other is used to create an appropriate input file for COHCAP (cohcap.inputfile).

This function will likely take several hours to run. However, it only needs to be run once.

#### See Also

Useful Example Files: http://sourceforge.net/projects/cohcap/files/COHCAP\_BSSEQ\_anno.zip/download \*Default settings utilize these files (in current working directory) \*These files were created using the UCSC Genome Browser (build hg19)

Raw Data for Demo Dataset: http://www.ncbi.nlm.nih.gov/sra/SRX084504 Full, Formatted Demo Dataset (in standalone package): http://sourceforge.net/projects/cohcap/

Bismark: http://www.bioinformatics.babraham.ac.uk/projects/bismark/

UCSC Genome Browser: http://genome.ucsc.edu/cgi-bin/hgTables?command=start

```
library("COHCAP")

dir <- system.file("extdata", package="COHCAP")
bed.folder <- file.path(dir,"BSSeq")
gene.table <- file.path(dir,"GENCODE_Genes_truncated.bed")
targeted.regions <- file.path(dir,"UCSC_CpG_Islands_truncated.bed")</pre>
```

```
output.folder <- getwd()
annotation.file <- file.path(output.folder,"COHCAP.targeted.BSSeq.anno.txt")
cohcap.inputfile <- file.path(output.folder,"BS_Seq_combined.txt")

COHCAP.BSSeq.preprocess(bed.folder, cohcap.inputfile, gene.table,
targeted.regions,annotation.file)</pre>
```

```
COHCAP.integrate.avg.by.island
```

Integration with Gene Expression Data(Average by Island Workflow).

## **Description**

Provides lists of genes with a significant negative correlation between DNA methylation and gene expression data.

A table of normalizated intensity / expression values is provided in the gene expression table and a table of filtered beta values is provided for the DNA methylation data.

Lists of genes with negative expression trends will be created in the "Integrate" folder, along with scatter plots (if descired). All correlation stats are provided in the "Raw\_Data" folder.

#### Usage

```
COHCAP.integrate.avg.by.island(beta.table, project.name, project.folder, expr.file, sample.file, cor.pvalue.cutoff=0.05, cor.fdr.cutoff = 0.05, cor.cutoff = -0.2, plot.scatter=TRUE, output.format = "xls")
```

## Arguments

-		
beta.table	Data frame with beta values averaged across differentially methylated sites (islands in rows, samples in columns). This table is already filtered for differentially methylated CpG islands.  The COHCAP.avg.by.island function automatically creates this file.	
project.name	Name for COHCAP project. This determines the names for output files.	
project.folder	Folder for COHCAP output files	
expr.file	Table of normalized expression or intensity values (can be for either microarray or RNA-Seq data).	
	Sample IDs (listed in column header) must match the sample IDs used for the DNA methylation data (e.g. those listed in beta.table)	
sample.file	Tab-delimited text file providing group attributions for all samples considered for analysis.	
	Only used if plot.scatter=TRUE	
cor.cutoff	The minimum negative correlation coefficient to define a differentially expressed	
	gene.	
cor.pvalue.cutoff		

Maximum p-value allowed to define a gene as differentially expressed.

cor.fdr.cutoff Maximum False Discovery Rate (FDR) allowed to define a gene as differentially expressed.

```
A logical value: Create scatter plot for genes with a significant negative correlation?

output.format Format for output tables: 'xls' for Excel file, 'txt' for tab-delimited text file
```

#### See Also

COHCAP Discussion Group: http://sourceforge.net/p/cohcap/discussion/general/

#### **Examples**

```
library("COHCAP")

dir <- system.file("extdata", package="COHCAP")
beta.file <- file.path(dir,"GSE42308_truncated.txt")
sample.file <- file.path(dir,"sample_GSE42308.txt")
project.folder <- getwd()
expression.file <- file.path(dir,"expression-Average_by_Island_truncated.txt")
project.name <- "450k_avg_by_island_test"

beta.table <- COHCAP.annotate(beta.file, project.name, project.folder,
platform="450k-UCSC")
filtered.sites <- COHCAP.site(sample.file, beta.table, project.name,
project.folder, ref="parental")
filtered.islands <- COHCAP.avg.by.island(sample.file, filtered.sites,
beta.table, project.name, project.folder, ref="parental")
COHCAP.integrate.avg.by.island(filtered.islands, project.name,
project.folder, expression.file, sample.file)</pre>
```

```
COHCAP.integrate.avg.by.site
```

Integration with Gene Expression Data(Average by Site Workflow).

## **Description**

Provides lists of genes with an inverse CpG island methylation trend (Methylation Down, Expression Up and Methylation Up, Expression Down).

Lists of genes with negative expression trends will be created in the "Integrate" folder.

The "Average by Site" workflow requires that genes already have fold-change, p-value, and FDR values calculated. There many tools available for this type of analysis (limma, sRAP, DEseq, edgeR, etc.)

This function will only work for 2-group comparisons.

## Usage

```
COHCAP.integrate.avg.by.site(island.table, project.name, project.folder, expr.file, expr.pvalue=0.05, expr.fdr = 0.05, expr.fc = 1.5, output.format = "xls")
```

#### **Arguments**

island.table Data frame with CpG island statistics (one row per CpG island) for differentially methylated CpG islands. The COHCAP.avg.by.site function automatically creates this file. Name for COHCAP project. This determines the names for output files. project.name project.folder Folder for COHCAP output files Table of differential expression statistics. Gene symbols must be in the first expr.file column, fold-change values must be in the second column, p-values must be in the third column, and false discovery rate (FDR) values must be in the fourth column. These statistics must be calculated outside of COHCAP. Duplicate gene symbols are OK - statistics will be averaged among duplicate gene symbols. expr.fc The minimum absolute value for fold-change values (treatment versus reference) to define gene as differentially expressed (from gene expression table). Only used for 2-group comparison. Fold-change is expected to be on a linear scale. Maximum p-value allowed to define a gene as differentially expressed (from expr.pvalue gene expression table). Maximum False Discovery Rate (FDR) allowed to define a gene as differentially expr.fdr expressed (from gene expression table) output.format Format for output tables: 'xls' for Excel file, 'txt' for tab-delimited text file

## See Also

COHCAP Discussion Group: http://sourceforge.net/p/cohcap/discussion/general/sRAP: http://www.bioconductor.org/packages/release/bioc/html/sRAP.html

```
library("COHCAP")

dir <- system.file("extdata", package="COHCAP")
beta.file <- file.path(dir, "GSE42308_truncated.txt")
sample.file <- file.path(dir, "sample_GSE42308.txt")
project.folder <- getwd()
expression.file <- file.path(dir, "expression-Average_by_Site_truncated.txt")
project.name <- "450k_avg_by_site_test"

beta.table <- COHCAP.annotate(beta.file, project.name, project.folder, platform="450k-UCSC")
filtered.sites <- COHCAP.site(sample.file, beta.table, project.name, project.folder, ref="parental")
filtered.islands <- COHCAP.avg.by.site(filtered.sites, project.name, project.folder)
COHCAP.integrate.avg.by.site(filtered.islands, project.name, project.folder, expression.file)</pre>
```

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COHCAP.qc DNA Methylation Quality Control Statistics	
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## **Description**

Provides descriptive statistics (median, top/bottom quartiles, mininum,maximum), sample histograms, sample dendrogram, principal component analysis plot.

Output files will be created in the "QC" subfolder.

#### Usage

```
COHCAP.qc(sample.file, beta.table, project.name, project.folder,
plot.legend=TRUE, color.palette = c("red","blue",
    "green","orange","purple","cyan","pink","maroon",
    "yellow","grey","black",colors()))
```

## **Arguments**

sample.file	Tab-delimited text file providing group attributions for all samples considered for analysis.
beta.table	Data frame with CpG sites in columns (with DNA methylation represented as beta values or percentage methylation), samples in columns, and CpG site annotations are included (in columns 2-5).
	The COHCAP.annotate function automatically creates this file.
project.name	Name for COHCAP project. This determines the names for output files.
project.folder	Folder for COHCAP output files
plot.legend	A logical value: Should legend be plotted within QC figures?
color.palette	Colors for primary variable (specified in the second column of the sample file).
	Remember, COHCAP can only analyze discrete variables categoried with groups (preferably with replicates).

## See Also

COHCAP Discussion Group: http://sourceforge.net/p/cohcap/discussion/general/

```
library("COHCAP")

dir <- system.file("extdata", package="COHCAP")
beta.file <- file.path(dir,"GSE42308_truncated.txt")
sample.file <- file.path(dir,"sample_GSE42308.txt")
project.folder <- getwd()
project.name <- "450k_test"

beta.table <- COHCAP.annotate(beta.file, project.name, project.folder, platform="450k-UCSC")
COHCAP.qc(sample.file, beta.table, project.name, project.folder)</pre>
```

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CpG Site Differential Methylation Analysis

## **Description**

Provides statistics for CpG sites as well as a list of differentially methylated sites. Can also provide .wig files for visualization in IGV, UCSC Genome Browser, etc.

List of differentially methylated sites and .wig files will be created in the "CpG\_Site" folder. Table of statistics for all CpG sites will be created in the "Raw\_Data" folder.

## Usage

```
COHCAP.site(sample.file, beta.table, project.name, project.folder,
methyl.cutoff=0.7, unmethyl.cutoff = 0.3,
delta.beta.cutoff = 0.2, pvalue.cutoff=0.05,
fdr.cutoff=0.05, ref="none", num.groups=2,
create.wig = TRUE, paired=FALSE, output.format = "xls")
```

#### **Arguments**

sample.file	Tab-delimited text file providing group attributions for all samples considered for analysis.	
beta.table	Data frame with CpG sites in columns (with DNA methylation represented as beta values or percentage methylation), samples in columns, and CpG site annotations are included (in columns 2-5).  The COHCAP annotate function automatically creates this file.	
	The COHCAF annotate function automatically creates this me.	
project.name	Name for COHCAP project. This determines the names for output files.	
project.folder	Folder for COHCAP output files	
methyl.cutoff	Minimum beta or percentage methylation value to be used to define a methylated CpG site. Default is 0.7 (used for beta values), which would correspond to 70 Used for either 1-group or 2-group comparison.	
unmethyl.cutoff		
	Minimum beta or percentage methylation value to be used to define an unmethy	

Minimum beta or percentage methylation value to be used to define an unmethylated CpG site. Default is 0.3 (used for beta values), which would correspond to 30 Used for either 1-group or 2-group comparison.

delta.beta.cutoff

fdr.cutoff

ref

The minimum absolute value for delta-beta values (mean treatment beta - mean reference beta) to define a differentially methylated CpG site. Only used for 2-group comparison.

pvalue.cutoff Maximum p-value allowed to define a site as differentially methylated. Used only for comparisons with at least 2 groups (with 3 replicates per group)

Maximum False Discovery Rate (FDR) allowed to define a site as differentially methylated. Used only for comparisons with at least 2 groups (with 3 replicates

per group)

Reference group used to define baseline methylation levels. Only used for 2-

group comparison

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num.groups Number of groups described in sample description file. COHCAP algorithm differs when analysing 1-group, 2-group, or >2-group comparisons. COHCAP cannot currently analyze continuous phenotypes.

create.wig A logical value: Create .wig files (using average beta and delta-beta values)?

In the standalone version of COHCAP, this was only an option when using the "Average by Site" workflow (because that was the only situation where the analysis method matched the visualization). .wig files are defined with respect to hg19 (for pre-defined annotation files) and can be visualized using IGV, UCSC

Genome Browser, etc.

paired A logical value: Is there any special pairing between samples in different groups?

If so, the pairing variable must be specified in the 3rd column of the sample description file. Used for p-value calculation, so this only applies to comparisons

with at least 2 groups.

output.format Format for output tables: 'xls' for Excel file, 'txt' for tab-delimited text file

#### Value

Data frame of average beta (or percentage methylation) statistics and/or p-value / false discovery rate statistics.

The content of the data frame depends upon the number of groups specified for analysis (avg.beta only for 1-group; avg.beta, delta.beta, p-value, and FDR for 2-group; p-value and FDR only for >2 groups).

This data frame is used for CpG island analysis.

#### See Also

COHCAP Discussion Group: http://sourceforge.net/p/cohcap/discussion/general/

```
library("COHCAP")

dir <- system.file("extdata", package="COHCAP")
beta.file <- file.path(dir,"GSE42308_truncated.txt")
sample.file <- file.path(dir,"sample_GSE42308.txt")
project.folder <- getwd()
project.name <- "450k_test"

beta.table <- COHCAP.annotate(beta.file, project.name, project.folder, platform="450k-UCSC")
filtered.sites <- COHCAP.site(sample.file, beta.table, project.name, project.folder, ref="parental")</pre>
```

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